

# Achieving fastest analyses with the Agilent 1200 Series Rapid Resolution LC system and 2.1-mm id columns

## Application Note

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### **Abstract**

The need to increase the daily throughputs of LC systems is a constant desire. Now, with the Agilent 1200 Series Rapid Resolution LC system highest throughputs are possible, and in combination with the Agilent ZORBAX RRHT columns and the increased pressure and temperature range of the LC system, excellent chromatographic resolution can be achieved even at run times below one minute.

This Application Note describes the correct set-up of the instrument which is the key for optimal results with narrow bore columns, such as a 2.1 mm x 50 mm column packed with sub two micron particles. Peak capacities in the range of fifty in analysis times as short as 24 seconds and peak widths as narrow as 200 milliseconds are shown. The well-balanced use of all possible module options to achieve shortest cycle times with throughputs far beyond 1500 samples per day is described.



**Agilent Technologies**

## Introduction

Particularly analytical service laboratories in the pharmaceutical industry, responsible for analyzing chemical libraries<sup>1</sup> or performing MS based quantifications of certain ADME-properties and drug metabolism studies of drug candidates<sup>2</sup> are faced with the challenge to increase their throughput, but also to maintain a high chromatographic resolution. In 2003 Agilent Technologies introduced sub two micron particles in their RRHT column series. Because of the small particle size, the chromatographic resolution obtainable with these columns is superior to standard particle sizes such as 3.5  $\mu\text{m}$  or even 5  $\mu\text{m}$ . Due to a unique silica manufacturing process, Agilent ZORBAX RRHT columns show a significantly reduced backpressure, if compared to similar column dimensions of other manufacturers. Excellent chromatographic results are achieved in a very short analysis time with the Agilent 1200 Series Rapid Resolution LC system, which facilitates an increased pressure range and flow rates from 0.05 up to 5 mL/min using column diameters ranging from 2.1-mm id up to 4.6-mm id. This Application Note will focus on 2.1-mm id columns only. Not only are the run times of the analyses important for high throughput, but also the overhead time. The Agilent 1200 Series Rapid Resolution LC system can be optimized to achieve highest throughputs with exceptionally good overall system performance.

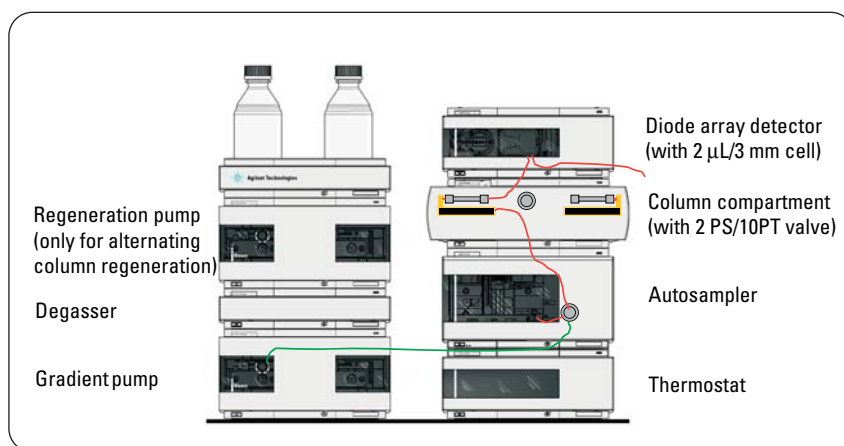
## Experimental

An important issue when dealing with narrow bore columns, especially in gradient mode where smallest peak widths can be achieved, is to have small extra column volumes. This also includes any volumes in front of the sampling device, because any volume after the solvent mixing point will increase the time for the gradient composition to reach the column. This results in an increased run time. The Agilent 1200 Series Rapid Resolution LC system can be reconfigured within a few minutes to provide appropriate system volumes for different column ids. Here, the pumps are set-up in the low delay volume configuration with an internal volume of approximately 120  $\mu\text{L}$ . All other modules are optimized for lowest delay volumes by using the low delay volume capillary kit (G1316-68744). Consequently, only capillaries of 0.12 mm id are used beyond the injection valve. In the Agilent 1200 Series thermostatted column compartment SL the newly introduced low dispersion

heat exchangers with 1.6  $\mu\text{L}$  internal volume were used. In some experiments, the Agilent 1200 Series Rapid Resolution LC is set up for alternating column regeneration to achieve highest throughput using the ACR-capillary kit (G1316-68721) and 2.1-mm id columns<sup>3</sup>. The high pressure rated 2-position/10-port valve in the thermostatted column compartment was only placed into the flow path if alternating column regeneration was used indeed.

The instrument set-up is as follows (figure 1):

- Agilent 1200 Series binary pump SL with the new Agilent 1200 Series micro vacuum degasser
- Agilent 1200 Series high performance autosampler SL
- Agilent 1200 Series thermostatted column compartment SL, equipped with a high pressure, 2-position/10-port valve, facilitating alternating column regeneration
- Agilent 1200 Series diode-array detector SL with a 2- $\mu\text{L}$ /3-mm cell
- ZORBAX SB C18, 2.1 mm id x 50 mm, 1.8  $\mu\text{m}$



**Figure 1**  
System setup with low delay volume for high speed applications using 2.1-mm id columns with lengths from 20 to 50 mm.

The Agilent 1200 Series binary pump SL is designed to fulfill the demands for high throughput, highest performance, optimum resolution and low-pump ripple. The pump hardware is significantly different from the standard binary pump. In the Agilent 1200 Series binary pump SL the pressure transducer is separate from the damper which has been modified to have a lower delay volume (pressure dependent ranging from 80–280  $\mu\text{L}$ ). In this study the pumps were used in the low delay volume configuration without the mixer and damper in the flow path. In contrast to the standard binary pump the pump heads of the binary pump SL have an additional damping coil (500  $\mu\text{L}$  volume each) to allow damping in the low delay volume configuration. This does not add to the gradient delay volume because it is before the mixing point. Anyhow, pressure ripples are also strongly suppressed by the Electronic Damping Control (EDC). The pressure range of the pump and all other modules is increased to 600 bar.

Only one sample, the so-called “phenone-mix”, was used in the course of this study to keep variations low. The sample consists of nine compounds: acetanilid, acetophenone, propiophenone, butyrophenone, benzophenone, valerophenone, hexanophenone, heptanophenone and octanophenone. Unless otherwise stated, the concentration was 0.1  $\mu\text{g}/\mu\text{L}$  for each compound except butyrophenone which was 0.2  $\mu\text{g}/\mu\text{L}$ . The solvent was water-acetonitril 2:1.

## Results and discussion

The most frequently sold particle size in chromatographic columns today is 5  $\mu\text{m}$ . Of course, fast and ultra fast LC is also possible with columns packed with particles of these larger diameters – the reduced

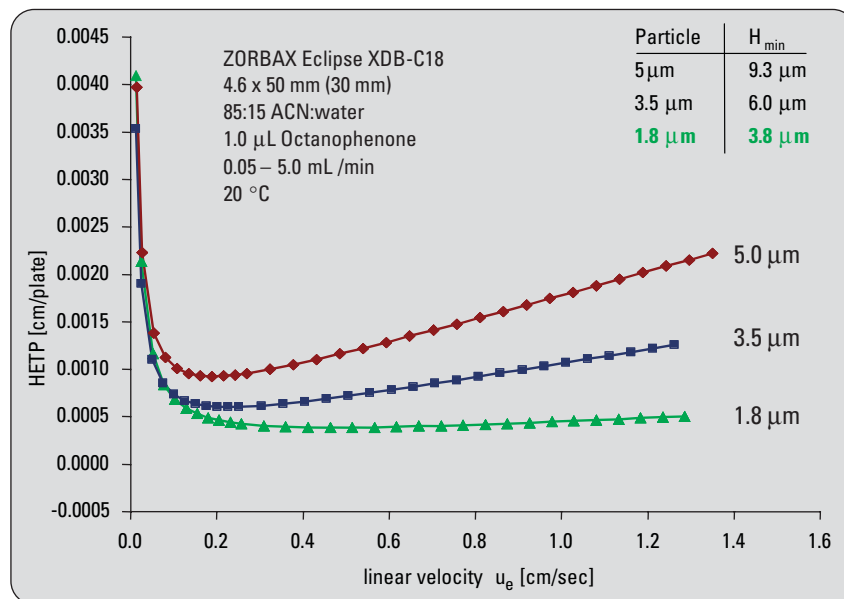


Figure 2  
Van Deemter curves of columns packed with 1.8  $\mu\text{m}$ , 3.5  $\mu\text{m}$  and 5.0  $\mu\text{m}$  particles.

back pressure is even beneficial to allow higher flow rates. However, resolution will be sacrificed because conditions are usually far on the right side of the van-Deemter-optimum. Here, the big advantage of the RRHT columns with particles of less than 2  $\mu\text{m}$  diameter is proven. The van Deemter optimum is shifted further to the right and the curve is much flatter at the onset because the “resistance of mass transfer” term is diminished (figure 2). In figure 3 the analysis on a 2.1-mm id column with 1.8- $\mu\text{m}$  particles is compared to the linear scaled analysis on the same stationary phase but on 5  $\mu\text{m}$  particles packed in a 4.6-mm id-column. The gain in resolution is obvious – from  $R_s = 2.1$  up to  $R_s = 3.5$  for the critical pair which matches the theoretically expected value of a 1.66 fold increase in resolution. Also note that there is a saving in solvent consumption of 8.6 mL in the “standard” HPLC analysis and only 1.8 mL in the ultra fast HPLC analysis.

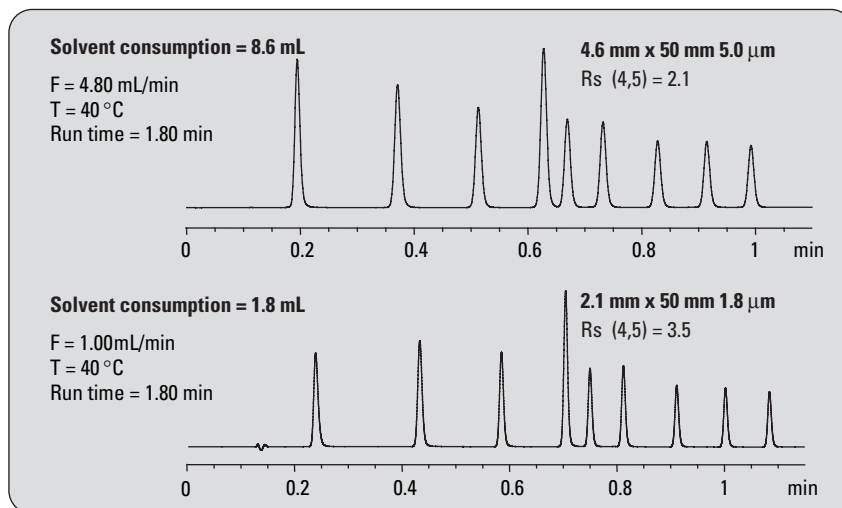
For gradient separation the dependencies of the capacity factor can be expressed as:

$$k^* = 0.87 \cdot tg \cdot \frac{F}{V_m \cdot \Delta\%B \cdot S}$$

( $tg$  = gradient time,  $F$  = flow rate,  $V_m$  = column void volume,  $\Delta\%B$  = gradient steepness,  $S$  = solvent and solute dependent factor)

If the product of the gradient time and flow rate, the so-called gradient volume, is kept constant together with all other parameters, the gradient time might be decreased while the flow rate is increased. Thus, the capacity factors of two compounds will stay constant and if no large alteration of the plate height occurs, the resolution will not change significantly, either. The final point is the big advantage of the sub two micron particles – the van-Deemter curve is nearly flat on the right side of the minimum (figure 2) and flow rates can be increased with only little increase in plate heights. However, the equation is an empirical one and deviations may occur especially under extreme conditions.

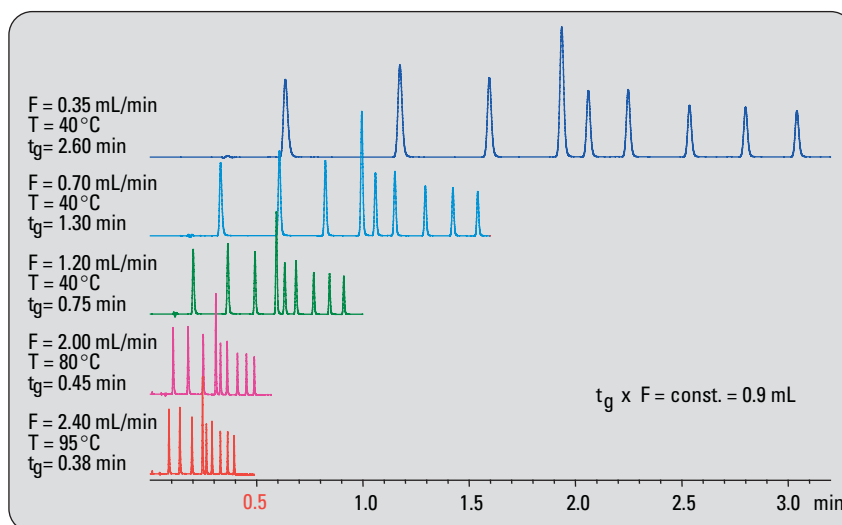
With a two-step approach, highest gradient speeds with virtually no loss or only little loss in resolution can be achieved. In the first step, start from a medium temperature and begin to increase the flow rate up to the pressure maximum. Subsequently the temperature should be increased to lower the viscosity of the solvent and then the flow rate is increased again. It may be worthwhile to check the resolution with two identical gradients but with different temperatures to see the influence of the temperature change on the resolution which may be very compound dependent. In figure 4 the result of this approach is shown. A nearly 7-fold increase in separation speed could be achieved with still base-line separation of the critical pair before meeting the pressure and temperature limit (the maximum temperature is a function of flow, temperature, number of controlled Peltier elements and of the heat capacity of the solvent used).



**Figure 3**  
Analysis with 1.8-µm particle column vs. 5.0 µm particle column.

<b>Conditions:</b>	4.6-mm id column used on standard Agilent 1200 system	
Solvent:	A = Water, B = ACN	
Temperature:	40 °C	
<b>Column:</b>	<b>2.1 mm x 50 mm, 1.8 µm</b>	<b>4.6 mm x 50 mm, 5.0 µm</b>
Flow:	1.0 mL/min	4.8 mL/min (scaled from 2.1 mm col.)
Gradient:	0.00 min 35 %B 0.90 min 95 %B 1.10 min 95 %B 1.11 min 35 %B 1.15 min	0.00 min 35 %B 0.90 min 95 %B 1.10 min 95 %B 1.11 min 35 %B 1.15 min
Stoptime:	0.70 min	0.70 min
Wavelength:	245 nm (8), ref. 450 nm (100)	245 nm (8), ref. 450 nm (80)
Peakwidth:	>0.0025 min (0.05 s res.time), 80 Hz	>0.01 min (>0.2 s), 20 Hz
Injection volume:	1 µL	5 µL (not scaled)

<b>Conditions:</b>	
Solvent:	A = water, B = ACN
Temp.:	40 °C, 80 °C, 95 °C
Flow:	0.35, 0.70, 1.20, 2.00, 2.40 mL/min
Gradient:	0.00 min 35 %B 2.60 min 95 %B 3.20 min 95 %B 3.21 min 35 %B
	<i>Time values for F = 0.35 mL/min. For all other flow rates times are scaled so that (tg x F) = 0.90 mL</i>
Stop time:	3.20 min
Post time:	2.00 min
Wavelength:	245 nm (8), Ref. 450 nm (100)
Peak width:	>0.0025 min (0.05 s response time), 80 Hz



**Figure 4**  
Increasing separation speed by increasing temperature and flow rate while decreasing gradient time.

The last chromatogram is enlarged in figure 5 and reveals the details of this separation. The first peak is eluted after only five seconds and peaks with a width at half height of less than 200 ms are achievable. Within twenty-four seconds nine compounds are separated with a peak capacity in the range of fifty.

#### Retention time precision at highest analysis speed

High analysis speed is meaningless without precision. One basic performance criteria for HPLC pumps is the precision of gradient formation measured by the precision of retention times of repeated gradients. However, the stability of the column temperature must also be taken into consideration, because temperature fluctuations will also influence the retention times of a given sample. In table 1 and figure 6 the results from the 10-fold repeated analysis of a standard sample are listed and since the deviation between individual runs is so small, the octanophenone peak is enlarged in a separate window. This sample contains compounds that are both not retained and refer to isocratically eluted compounds found at the starting conditions of the gradient, as well as highly unpolar and strongly retained compounds. The analyses

#### Conditions:

Solvent: A = Water, B = ACN  
 Temp.: 40 °C, 80 °C  
 Flow: 0.35 mL/min, 1.20 mL/min, 2.0 mL/min  
 Gradient: 0.00 min 35%B  
 2.60 min 95%B  
 3.20 min 95%B  
 3.21 min 35%B  
*Time values for F = 0.35 mL/min.  
 For all other flow rates times are scaled so that (time x flow) = 0.90 mL*  
 Stop time: 3.20 min  
 Post time: 2.00 min  
 Injection vol.: 1.0 µL

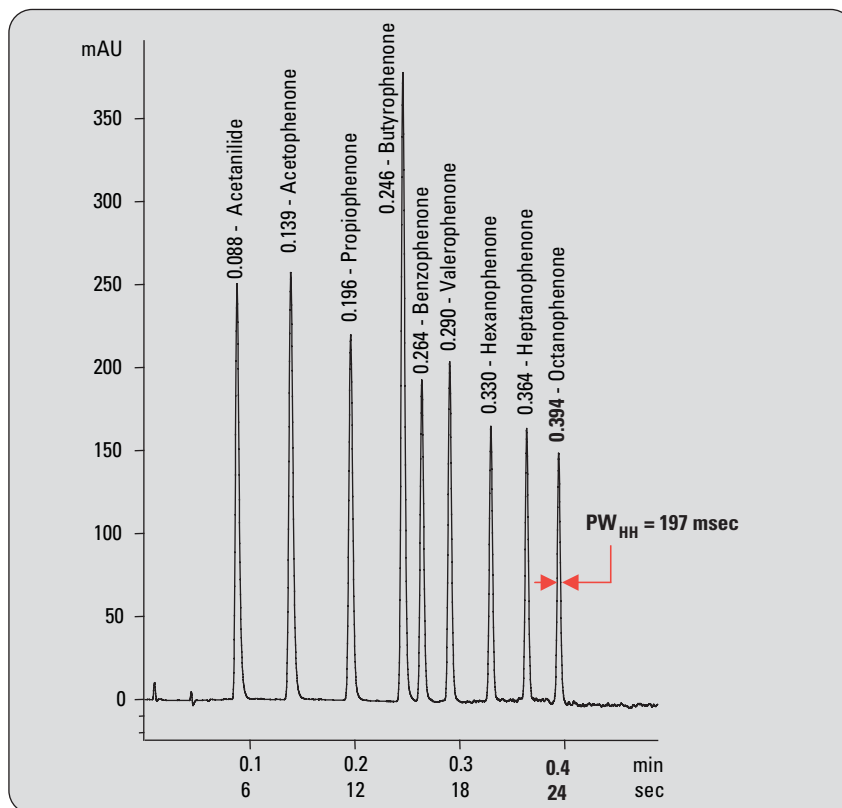


Figure 5  
Separation of a nine compound mixture under ultra fast conditions.

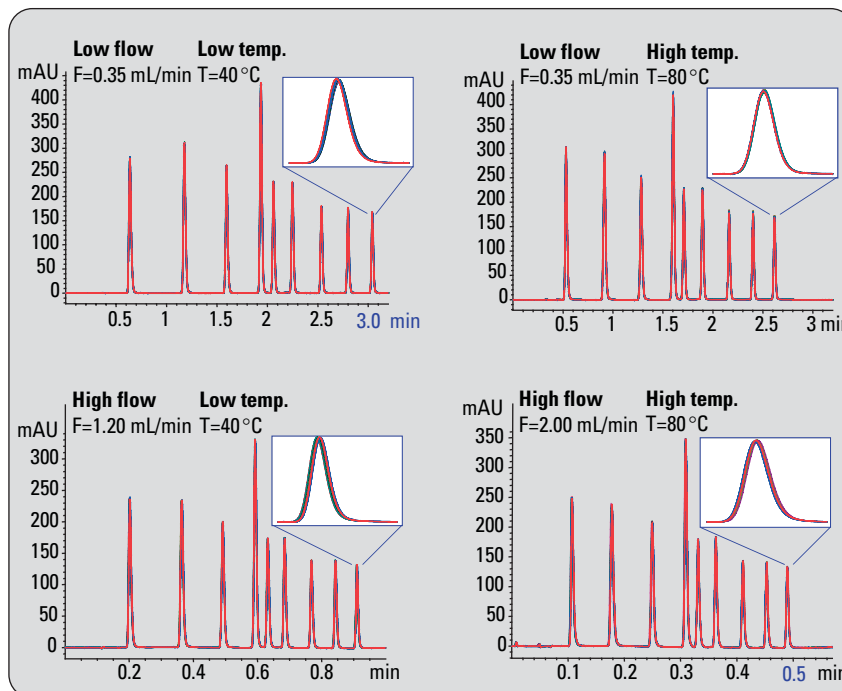


Figure 6  
Overlaid chromatograms of the repeated analysis of a 9 compound mixture under various conditions.



were done at high and low flow rates as well as with high and low temperatures as in the examples shown earlier. In all cases the mean retention time precision is below 0.3 % RSD, which was the specification of the Agilent 1100 Series LC system. Of course, the results are also in line with the specifications for the new Agilent 1200 Series Rapid Resolution LC system which is < 0.07 % RSD or < 0.02 min SD, whichever is met first. At these high gradient speeds, the SD criteria are always met. The RSD criteria are also met for both fast-LC gradients of 2.6 min duration (0.35 mL/min flow rate). Even at ultra-fast gradient speeds, the retention time precisions are still below or only slightly higher than 0.1% RSD (table 1).

### Improving the cycle-time

Not only is the gradient speed important when dealing with high-throughput analysis but furthermore the over all cycle time of the entire system, which is the time between two consecutive analyses. A good method to measure the cycle time is by using the time stamp the data file is assigned by the operating system of the computer. Clearly, optimizing the cycle time has some drawbacks. For example, extensive needle cleaning procedures are in contradiction with a high sampling speed. Table 2 gives an overview of important parameters influencing the cycle time. Using 1.8- $\mu$ m particle size columns together with an optimized HPLC system very short run times can be achieved without sacrificing chromatographic resolution. Combining short run times together with low overhead times will result in a high daily throughput. In figure 7 the cycle time and daily throughput is shown for two

	0.35 mL/min, 40°C		0.35 mL/min, 80°C		1.20 mL/min, 40°C		2.00 mL/min, 80°C	
	SD	% RSD	SD	% RSD	SD	% RSD	SD	% RSD
Average	0.00107	0.067	0.00084	0.070	0.00048	0.098	0.00031	0.134

**Table 1**  
Standard deviations (mAU) and %RSD (n=10) of the retention times under different chromatographic conditions in temperature and flow.

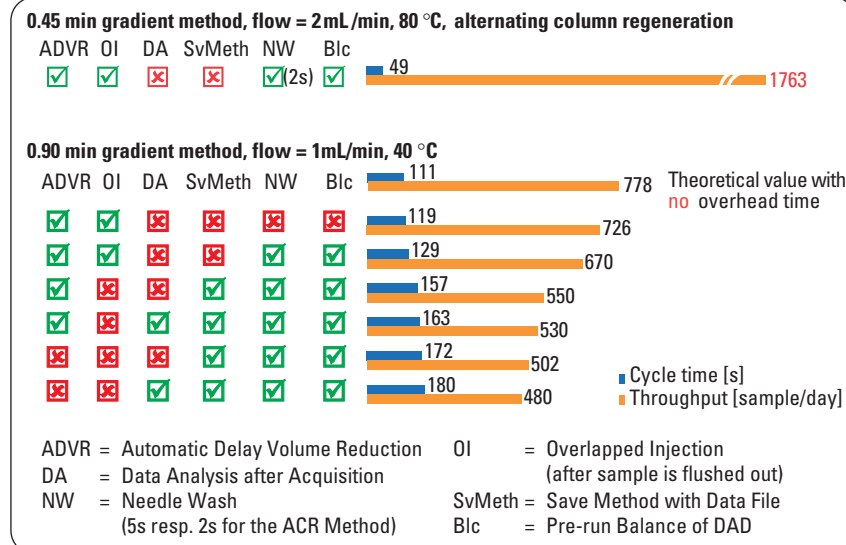
Module	Parameter	Effect on cycle time	Other effects
Pump	Low delay volume setting	Reduced retention times, run time can be shortened, reduced cycle time	Increased pressure ripple, slightly increased mixing noise if modifiers such as TFA are used.
	Automatic Delay Volume Reduction (ADVR) – activated	Reduced delay volume, reduced retention times, run time can be shortened, reduced cycle time	Increased carry-over
	ADVR activated and Overlapped Injection (OI)	Enables parallel sampling, thus reduces the cycle time independently of the below listed settings (as long as the overall sampling speed does not exceed the gradient and post time)	Increased carry-over
	no OI – Needle Wash	Increased sampling time with increasing wash time	Reduced carry-over with longer needle wash time
	no OI – Equilibration time	Increased sampling time with increased equilibration time	Better injection precision with longer equilibration time
Column compartment	no OI – Draw/Eject speed	Low speed causes increased sampling time	Low speed results in better injection precision
	Alternating column regeneration	Saves column wash-out and equilibration time, reduces cycle time enormously	Additional hardware required, slightly increased extra column volume, slightly different retention times between columns possible
Detector	Pre-run and/or post-run balance	Increased cycle time	Baseline drifts possible if not applied
	Spectral data acquisition with high data rate, small band width and broad wavelength range large data files	Depending on computer power and additional processes running might increase cycle time because of writing speed	Reduced information content if no spectral data acquired or with lower resolution
Software	Data analysis with acquisition	Increased cycle time, depending on computer power and number of peaks	Data analysis has to be done offline is no set
	Save method with data	Slightly increased cycle time	Information is missing if method is not saved
	Execution of pre-run or post-run macros	Increased cycle time, depending on macro	Depending on macro
System	LC controlled over local network between computer and LC (and MS) only	Faster data and method transfer between computer and LC because of reduced net work traffic reduced cycle time	Additional hardware might be necessary (use independent acquisition computer)
	Number of detectors	More detectors produce a higher data amount and lower the data transfer speed, resulting in higher cycle times	More detectors higher information content

**Table 2**  
Influence of various parameters on the overall cycle time.

different methods – both giving virtually the same resolution. The first method (0.45 min gradient) utilizes alternating column regeneration and high temperatures to allow high flow rates and speed optimized settings. A cycle time of 49 s could be achieved, resulting in a theoretical daily throughput of more than 1700 samples per day. The second method (0.90 min gradient) does not use high temperatures or alternating column regeneration and the time saving of some simple and often forgotten method options are shown. By optimizing these parameters the real cycle time gets as close to 8 s to the run time (stop time plus post time) and allows a daily throughput of more than 700 samples per day. By sub-optimal method set up this can easily drop to below 500 samples per day if options like automatic delay volume reduction, overlapped injection or offline data-analysis are not used.

## Conclusion

The Agilent 1200 Series Rapid Resolution LC system is a powerful tool to achieve highest chromatographic resolutions and also highest throughputs. The extended pressure range allows the usage of columns packed with stationary phases with particles sizes below 2 µm, for example, Agilent RRHT columns with particle sizes of 1.8 µm. These columns not only allow an increase in linear flow rates with virtually no loss in resolution but also have an inherently higher resolution compared to 3.5 µm or even 5.0 µm particle sizes. The possibility to switch the pump into its low delay volume configuration allows the use of the entire bandwidth of today's widely used column ids – from 4.6 mm



**Figure 7**  
Cycle time and daily throughput optimization.

### Chromatographic conditions:

#### Alternating Column Regeneration Method

Solvent: A = Water, B = ACN  
 Temp.: 80 °C  
 Flow: 2.0 mL/min  
 ADVR: Yes  
 Gradient:

#### Gradient-Pump

0.00 min 35 %B  
 0.45 min 95 %B  
 0.46 min 35 %B  
 0.57 min 35 %B

#### Regeneration-Pump

0.00 min 35 %B  
 0.01 min 95 %B  
 0.11 min 95 %B  
 0.12 min 35 %B

Stoptime: 0.57 min  
 Posttime: off  
 Wavelength: 245 nm (8), ref. 450 nm (100)  
 Peak width: > 0.0025 min (0.05 s response time), 80 Hz  
 Spectra: none  
 Injection volume: 1.0 µL  
 Injector: Overlapped injection, 2 s needle wash, sample flush-out factor = 10, draw/eject speed = 100 µL/min  
 Valve: next position

#### No Alternating Column Regeneration Method

Solvent: A = Water, B = ACN  
 Temp.: 40 °C  
 Flow: 1.0 mL/min  
 ADVR: Yes  
 Gradient:

0.00 min 35 %B  
 0.90 min 95 %B  
 1.10 min 95 %B  
 1.11 min 35 %B  
 1.15 min

No

0.00 min 35 %B  
 0.90 min 95 %B  
 1.10 min 95 %B  
 1.11 min 35 %B

Stoptime: 1.15 min  
 Posttime: 0.70 min  
 Wavelength: 245 nm (8), ref. 450 nm (100)  
 Peak width: > 0.0025 min (0.05 s response time), 80 Hz  
 Spectra: all, 190-500 nm, BW = 1 nm  
 Injection volume: 1.0 µL  
 Injector: See figure 7, 2 s equilibration time

1.40 min (add. 300 µL extra column volume, increased retention times)  
 0.70 min

down to 2.1 mm and even 1.0 mm. As illustrated above, the system has uncompromised performance

characteristics even at highest gradient speeds.

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