

M. Wood¹, G De Boeck², N. Samyn², V. Maes³, M. Morris¹¹ Waters Corporation, Floss Road, Wythenshawe, Manchester M23 9LZ, UK.² National Institute of Criminalistics and Criminology (NICC), Section Toxicology, Vilvoordsesteenweg 98, 1120 Brussels, Belgium³ Department of Clinical Chemistry-Toxicology, Academic Hospital, Free University of Brussels, BelgiumPresented at TIAFT, Paris, France, 26th - 30th August, 2002**OBJECTIVE**

To develop and validate a rapid and sensitive LC-MS/MS method for the simultaneous quantitation of benzodiazepines in plasma. The method should require minimal sample preparation.

INTRODUCTION

- Benzodiazepines are the most widely prescribed psychoactive drugs in the world for the symptomatic treatment of anxiety and sleep disorders.
- Here we describe the development of a rapid and sensitive LC-MS/MS method for the quantification of 10 benzodiazepines. Limits of detection 0.4 µg/L or better were achieved when just 50 µL plasma was used.
- Results were compared with those obtained using a commercial ELISA kit and a validated reference HPLC-DAD method.

EXPERIMENTAL**Validation Samples**

Plasma samples were obtained from the Hospital Emergency Department.

HPLC-DAD

Plasma samples were initially screened for a broad range of medicinal drugs by HPLC-DAD. Briefly, after a liquid extraction of 1 mL of plasma at pH 9.2 with 1-chlorobutane, the organic phase was evaporated and redissolved in 150 µL of mobile phase (phosphate buffer pH 3.8- acetonitrile 67/33 (v/v)). Fifty microlitres was injected on a Waters Symmetry C₁₈ (3.9 x 150 mm, 5 µm) column at 33°C.

ELISA

Plasma samples were also screened using a commercial ELISA-kit (Cozart, UK) according to the manufacturer's instructions. Negative and positive calibrators (oxazepam in the range 0-500 µg/L) were pipetted (in duplicate) onto the same microtitre plates as the unknown samples.

LC-MS/MS conditions**HPLC**

HPLC System: Waters Alliance 2690

Column: Zorbax SB-phenyl column
(2.1 x 150mm, 5 µm)Mobile phase : A = 10:10:80
acetonitrile:methanol: 20mM
ammonium acetate
B = 95:5 acetonitrile: 20mM
ammonium acetate

Time (min)	A (%)	B (%)	Curve number
0	100	0	1
0.5	75	25	1
8	40	60	7 (concave)
11	40	60	6 (linear)
12	100	0	1
15	100	0	1

Flow rate: 0.25 mL/min

Injection volume: 10 µL

MS conditionsMass spectrometer: Micromass Quattro Ultima
(Figure 1)

Ionisation Mode: ES positive ion

Capillary voltage : 3kV

MS/MS: Collision gas Argon at 2.5 x
10⁻³ mbar

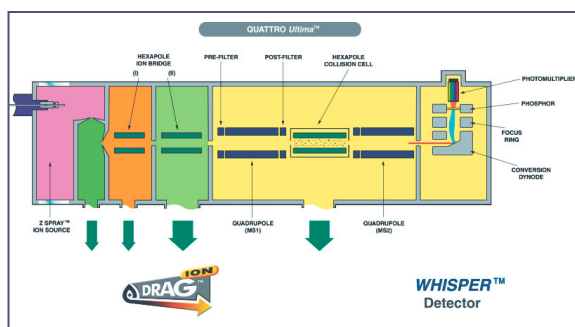


Figure 1. The Quattro Ultima triple quadrupole mass spectrometer incorporating 'Z' Spray source

RESULTS AND DISCUSSION

Figure 2 shows the MS and MS/MS spectra for a selection of the benzodiazepines. The MRM transitions and conditions used for the measurement of the benzodiazepines and their respective deuterated analogues are summarised in Table 1. The latter were used as internal standards for quantification purposes.

Compound	Precursor ion (<i>m/z</i>)	Product ion (<i>m/z</i>)	Cone voltage (V)	Collision energy (eV)
Alprazolam	308.8	280.9	70	25
Alprazolam-d5	313.8	285.8	100	25
Clonazepam	315.8	269.8	80	25
Clonazepam-d4	319.9	273.8	100	25
Diazepam	284.9	154.0	60	25
Diazepam-d5	289.8	153.7	80	25
Flunitrazepam	313.9	267.9	80	25
Flunitrazepam-d7	320.8	274.8	80	25
Lorazepam	320.8	274.7	60	23
Lorazepam-d4 [†]	326.8	280.8	80	23
Nordiazepam	270.9	139.8	80	25
Nordiazepam-d5	275.9	139.8	60	25
Oxazepam	287.0	240.8	60	26
Oxazepam-d5	291.7	245.8	80	26
Prazepam	324.9	270.9	80	25
Prazepam-d5	330.0	276.0	80	25
Temazepam	300.9	255.0	60	25
Temazepam-d5	305.8	259.8	60	25
Triazolam	342.9	307.7	60	25
Triazolam-d4 [†]	349.0	313.9	60	25

Table 1. MRM transitions and conditions for the measurement of 10 benzodiazepines. [†]For these 2 compounds the isobaric nature between the analogue and their respective non-deuterated compounds meant that an alternative precursor ion was necessary.

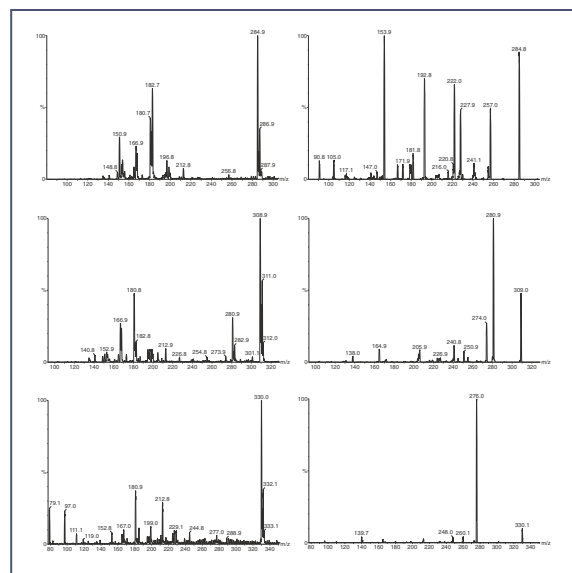


Figure 2. MS and MS/MS spectra of (top to bottom) diazepam, alprazolam and the deuterated internal standard for prazepam (d5).

A series of calibrators (1, 10, 40, 100, 200, 400 and 800 µg/L) were prepared by adding the benzodiazepines to drug-free plasma. Plasma samples were isolated from the matrix using a simple acetonitrile clean up procedure which also incorporates the addition of the internal standards.

Figure 3 shows the MRM chromatograms of the benzodiazepines obtained with a 10 µL injection of the 10 µg/L plasma calibrator. Quantification was performed by integration of the area under the specific MRM chromatograms. Figure 4 shows a typical standard curve for diazepam in plasma. Responses were linear, in all cases, over the range investigated (Coefficient of Determination > 0.99).

Table 2 compares the results for 20 plasma samples which were analysed using HPLC-DAD, LC-MS/MS and ELISA.

Plasma sample	HPLC-DAD	LC-MS/MS	ELISA *
1	Lora (24), Mida (25)	Lora (23)	negative (35)
2	D (59), ND (<20), Lora (24)	D (53), ND (7), Lora (29)	positive (566)
3	Alp (45)	Alp (48)	positive (506)
4	D (150), ND (34), Clon (<10)	D (129), ND (28), Clon (12), Ox (3), Tem (4)	positive (708)
5	Lora (30)	Lora (35)	negative (12)
6	D (1785), ND (1350)	D (1682), ND (1262), Ox (73), Tem (62)	positive (746)
7	ND (575) + Praz intox	ND (567), Ox (57), Pra (13)	positive (692)
8	D (850), ND (470)	D (763), ND (398), Ox (10), Tem (41)	positive (742)
9	Alp (15)	Alp (13)	positive (396)
10	D (280), ND (<20)	D (250), ND (5), Tem (3)	positive (681)
11	Brom (390), Lora (24)	Lora (30), D (10)	positive (353)
12	Alp (420)	Alp (364)	positive (767)
13	Lora (103)	Lora (117), D (17), ND (11)	positive (612)
14	Brom (200)	D (72), ND (20)	positive (589)
15	Ox (1300)	Clon (8), Flu (11), Ox (>800)	positive (792)
16	Alp (140)	Alp (147)	positive (648)
17	Brom (176), D (1290), ND (710)	D (1196), ND (655), Ox (47), Tem (46)	positive (806)
18	Alp (20), Lora (32)	Alp (19), Lora (34)	positive (294)
19	Flu (34)	Flu (26)	negative (6)
20	Brom (8300), D (3380), ND (2250)	D (3117), ND (2086), Ox (187), Tem (305)	positive (805)

Table 2. Benzodiazepine quantification in 20 plasma samples using HPLC-DAD, LC-MS/MS and ELISA.

Key: D=Diazepam, ND=nordiazepam, Tem=temazepam, Ox=oxazepam, Lora=lorazepam, Alp=alprazolam, Pra=prazepam, Mida=midazolam, Clon=clonazepam, Brom=bromazepam. Bracketed values are drug concentration in µg/L.

* A cut-off value of 100µg/L was applied to the ELISA results.

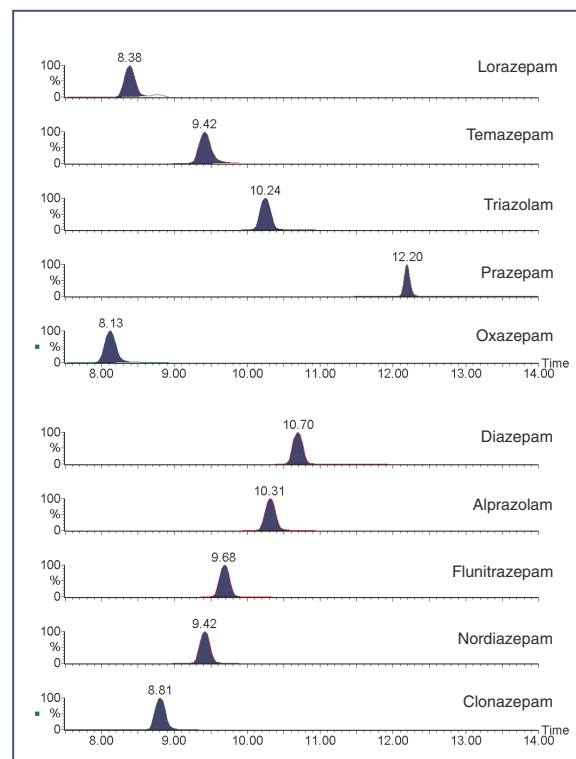


Figure 3. MRM chromatograms for (top to bottom): lorazepam, temazepam, triazolam, prazepam, oxazepam, diazepam, alprazolam, flunitrazepam, nordiazepam and clonazepam. Responses were obtained with a 10µL injection of the 10µg/L plasma calibrator

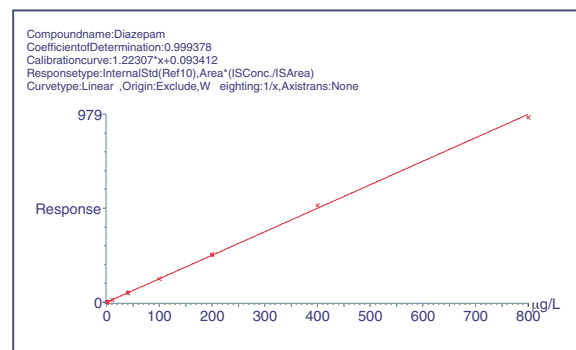


Figure 4. Typical response for plasma containing diazepam. Diazepam spiked plasma was firstly extracted using acetonitrile prior to analysis using HPLC/MRM. Benzodiazepines were quantified by reference to their deuterated internal standards

CONCLUSION

LC-MS/MS

We have developed a simple, rapid method that allows the simultaneous quantification of 10 benzodiazepines in a single chromatographic run. Only 50µL sample is required. The procedure involves a simple protein precipitation step with acetonitrile followed by HPLC/MS analysis and is less time-consuming and labour-intensive than the existing GC/MS and LC-DAD methods. The developed method has been successfully applied to the analysis of plasma samples collected from current benzodiazepine users and compared with the results of a validated reference HPLC-DAD screening method.

ELISA

A cut-off value of 100 µg/L was applied to the data which resulted in an accuracy of 89 %, a sensitivity of 87 % and a specificity of 100 % for a total of 62 samples. Lowering the cut-off to 50 µg/L does not change the outcome significantly. When a cut-off of 10 µg/L is applied, the accuracy is 90 %, sensitivity 93 % and specificity only 71 %. A lower cut-off value leads to a number of false positives, also due to the difference in the nature of the matrix of the samples. False negatives were only obtained for low concentrations of lorazepam and flunitrazepam.

FUTURE AIMS

- To validate the method for alternative specimens.
- To apply the method to insects recovered from decomposing human remains (forensic entomology) and post-mortem samples (hair, body fluids, tissue).

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