# ASMS 2004

Eric S. Grumbach, Diane M. Diehl and Jeffrey R. Mazzeo Waters Corporation, 34 Maple St. Milford MA, 01757

#### **Overview**

Many LC/ESI-MS assays use 2 mm i.d. columns packed with 5 µm particles. This situation is a result of the widespread availability of these columns, the compatibility of the optimal flow rates of these columns with electrospray ionization interfaces, and the ability of modern HPLC instrumentation to run these columns with minimal loss in performance due to extra-column bandspreading. Utilizing 1.7 µm particles and 1 mm i.d. columns should lead to an improvement in sensitivity leading to lower limits of detection. These improvements are dependant on having an LC system that operates at higher pressures and has minimal system volume.

#### **Methods**

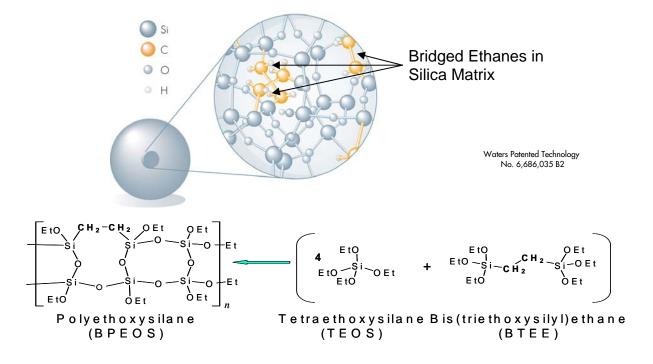
LC/ESI-MS sensitivity of a traditional HPLC and UPLC<sup>™</sup> (Ultra Performance LC<sup>™</sup>) were compared using a 2.1 mm i.d. column packed with 5 µm bridged ethyl-siloxane hybrid (BEH) particles to demonstrate the benefit of using a low dispersion LC as an MS inlet in order to gain sensitivity.

LC/ESI-MS sensitivity was further compared on the UPLC<sup>™</sup> using 2.1 mm i.d. columns packed with 5 µm and 1.7 µm BEH particles to establish the benefit of using sub-2 µm particles for improving sensitivity when performing LC/MS<sup>n</sup>.

Additionally, a 1 mm i.d. column packed with 1.7 µm BEH particles was used to demonstrate improved limits of detection compared to traditional 2.1 mm i.d. columns packed with 5 µm particles.

#### A New Generation of Hybrid Packings

Optimal linear velocities of sub-2 µm particles require operations at higher pressures. Inorganic-organic hybrid materials have demonstrated increased mechanical stability compared to traditional silica based materials\*1. A 1.7 µm bridged ethyl-siloxane hybrid (BEH) particle was designed to meet the demands of operation at pressures as high as 15,000 PSI, as well as mobile phase pH in the range of 1-12.



# Utilizing a Low Dispersion UPLC<sup>TM</sup> System with Conventional 5 µm Particles as a Mass Spectrometer Inlet

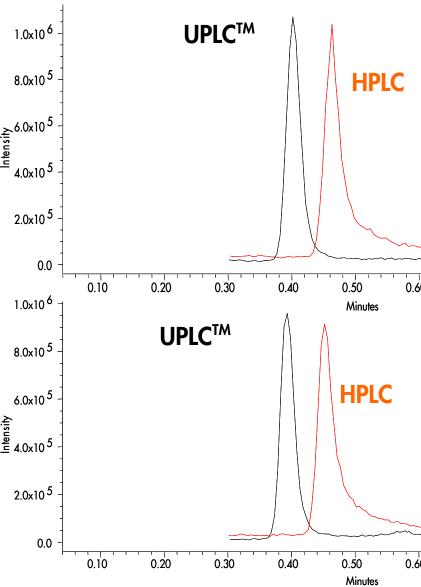
#### **Chromatographic Conditions:**

Column: BEH C<sub>18</sub> 2.1 x 50 mm, 5.0 µm Mobile Phase A: 0.1% NH<sub>4</sub>OH in H<sub>2</sub>O Mobile Phase B: 0.1% NH₄OH in MeOH Flow Rate: 0.6 mL/min Gradient: Profile Time (min) %A %B 20 80 5 95 0.75 Injection Volume: 20 µL (HPLC); 15 µL (UPLC™) Sample Diluent: 70:30 MeOH: H<sub>2</sub>O with 0.1% NH<sub>4</sub>OH Sample Concentration: 0.5 ng/mL

Temperature: 38 °C

Instrument:

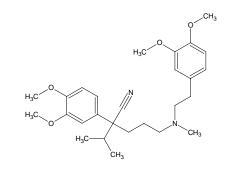
HPLC: Alliance<sup>®</sup> HT System, 2795 Separations Module with Waters ZQ<sup>TM</sup> **UPLC<sup>™</sup>:** ACQUITY UPLC<sup>™</sup> with Waters ZQ<sup>™</sup>



sensitivity can be observed by utilizing a smaller particle size.

#### Mass Spectrometer Conditions (ESI+):

Capillary (kV): 3.0 Cone (V): 35 (MV), 30 (V) Extractor: 3 V RF Lens: 0.5 V Source Temperature (°C): 150 Desolvation Temperature (°C): 350 Cone Gas Flow (L/Hr): 60 Desolvation Gas Flow (L/Hr): 500 SIR m/z: 455.45 verapamil (V); 485.45 methoxyverapamil (MV) Dwell Time: 50 milliseconds Interchannel Delay: 50 milliseconds Interscan Delay: 50 milliseconds



Verapami

0.80 0.60 0.70 0.90 Methoxyverapamil 0.80 0.60 0.70 0.90

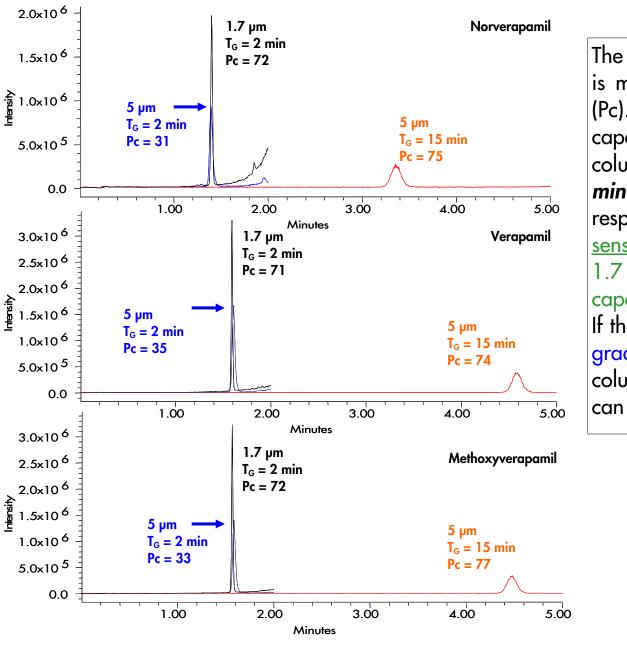
## Improved ESI-MS Response Utilizing 2.1 mm i.d. Columns Packed with 1.7 µm Particles

#### Chromatographic Conditions:

Column: ACQUITY UPLC<sup>™</sup> BEH C<sub>18</sub> 2.1 x 50 mm, 1.7 µm, BEH C<sub>18</sub> 2.1 x 50 mm, 5.0 μm Mobile Phase A: 0.1% NH<sub>4</sub>OH in H<sub>2</sub>O Mobile Phase B: 0.1% NH₄OH in MeOH Flow Rate: 0.5 mL/min Profile Gradient: Time %A %B 50 50 5 95 15/2.0 Injection Volume: 20 µL Sample Diluent: 70:30 MeOH: H<sub>2</sub>O with 0.1% NH<sub>4</sub>OH Sample Concentration: 1.0 ng/mL Temperature: 38 °C

# Instrument: ACQUITY UPLC<sup>TM</sup> with Waters ZQ<sup>TM</sup>

Capillary (kV): 3.0 Cone (V): 35 (NV, MV), 30 (V) Extractor: 3 V RF Lens: 0.5 V Source Temperature (°C): 150 Desolvation Temperature (°C): 350 Cone Gas Flow (L/Hr): 60 Desolvation Gas Flow (L/Hr): 500 SIR m/z: 441.4 norverapamil (NV); 455.45 verapamil (V); 485.45 methoxyverapamil (MV) Dwell Time: 50 milliseconds Interchannel Delay: 50 milliseconds Interscan Delay: 50 milliseconds



An improvement in peak shape is observed due to the lower dispersion volume of the ACQUITY UPLC<sup>TA</sup> system. Although the true benefits of UPLC<sup>™</sup> cannot be reached unless sub-2 µm particles are used, the ACQUITY UPLC<sup>™</sup> system can be used as a low dispersion LC for conventional particles. Greater sensitivity can be achieved due to the low dispersion characteristics of the UPLC<sup>™</sup> system. A further improvement in

Several factors contribute to the improved response of these analytes at low ng/mL concentrations on a single quadrupole mass spectrometer. The largest contributing factors being the particle size and mobile phase composition. As the particle size is decreased, the peak width decreases and the peak height increases, thus yielding higher analyte response. Additionally, the use of basic pH under ESI<sup>+</sup> conditions has demonstrated a significant signal enhancement effect compared to equivalent concentrations of acidic limits of detection (0.025 ng/mL) mobile phases for a majority of basic analytes\*<sup>2</sup>. The high organic content of the mobile phase also contributes to enhanced analyte response allowing for efficient mobile phase desolvation and compound Wyndham, K.D., O' Gara, J.E., Walter, T.H., Glose, K.H., Lawrence, N.L., Alden, B.A., Izzo, G.S., Hudalla, C.J. and Iraneta, P.C. Anal, Chem. 2003, 75, 6781-6788 ionization. All of these factors allow for lower limits of detection compared to traditional HPLC analysis. Mallet, C.R., Lu, Z. and Mazzeo, J.R., Rapid Commun. Mass Spectrom. 2004; 18: 49-58

Mass Spectrometer Conditions (ESI+):

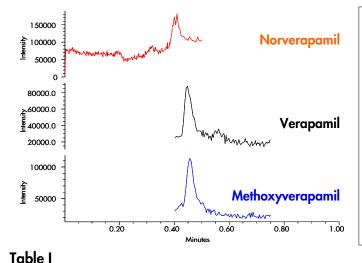
The efficiency of a gradient separation is measured in terms of peak capacity (Pc). In order to achieve equivalent peak capacity on both the 1.7 µm and 5 µm columns, a gradient duration ( $T_G$ ) of **2** minutes and 15 minutes must be used, respectively. A <u>5X improvement in</u> Table I sensitivity can be observed on the 1.7 µm column at the same peak capacity and within <u>8X faster</u> run times. If the 5 µm column is run with the same gradient duration time as the 1.7 µm column, a 2X improvement in sensitivity can be observed on the 1.7 µm column

> 5 µm  $PSI_{MAX} = 2,300$

1.7 µm  $PSI_{MAX} = 11,100$ 

## Single Quadrupole Linearity and Limits of Detection at Sub-ng/mL Analyte Concentrations

An ACQUITY UPLC<sup>™</sup> BEH C<sub>18</sub>, 1.0 x 50 mm, 1.7 µm column was used to demonstrate the low limits of detection achievable when using 1.0 mm i.d. columns packed with 1.7 µm particles. Verapamil and related compounds were analyzed over the working range of 0.01-10.0 ng/ mL, using linear regression to determine the Limits of Detection (LOD) and Limits of Quantitation (LOQ). Limits of detection were determined to be 0.025 ng/mL for V, NV and MV based on a signal-to-noise ratio of 3:1. Limits of quantitation were determined to be 0.05 ng/mL for V, NV and MV based on a signal-to-noise ratio of 10:1. Six replicates were performed to yield correlation coefficients of 0.9993, 0.9962 and 0.9997 for verapamil, norverapamil and methoxyverapamil, respectively. Linearity of the mass spectrometer over 3 orders of magnitude for each individual standard is shown in Table 1.



An extremely low LOD of 0.025 ng/ mL for V, NV and MV is achievable on a single quadrupole mass spectrometer by the utilization of 1.0 mm i.d. columns packed with 1.7 µm particles. Typical values using single quadrupole MS under single ion recording mode are 10 ng/mL (<u>+</u>10X).

of Cal	ibration	Standa	rds (N =	= 6)					
0.01	0.025	0.05	0.10	0.25	0.50	1.00	2.50	5.00	10.00
		0.05	0.10	0.25	0.53	1.09	2.58	5.09	9.70
0.00	6.15	0.00	0.00	0.00	4.00	5.84	2.19	1.25	2.19
0.01	0.025	0.05	0.10	0.25	0.50	1.00	2.50	5.00	10.00
0.01	0.025	0.04	0.09	0.26	0.53	0.99	2.59	5.43	9.55
0.00	2.95	17.68	7.86	2.72	4.00	0.71	2.46	5.60	3.33
mL)									
0.01	0.025	0.05	0.10	0.25	0.50	1.00	2.50	5.00	10.00
0.01	0.024	0.05	0.10	0.24	0.51	1.07	2.60	5.01	9.82
0.00	2.95	0.00	0.00	2.95	1.39	4.63	2.72	0.14	1.30
	0.01 0.01 0.00 0.01 0.01 0.00 <i>mL)</i> 0.01 0.01	0.01 0.025 0.01 0.024 0.00 6.15 0.01 0.025 0.01 0.025 0.00 2.95	0.01 0.025 0.05 0.01 0.024 0.05 0.00 6.15 0.00 0.01 0.025 0.05 0.01 0.025 0.04 0.00 2.95 17.68 mL) 0.01 0.025 0.05 0.01 0.025 0.05 0.01 0.024 0.05	0.01 0.025 0.05 0.10 0.01 0.024 0.05 0.10 0.00 6.15 0.00 0.00 0.01 0.025 0.05 0.10 0.01 0.025 0.04 0.09 0.00 2.95 17.68 7.86 mL) 0.01 0.025 0.05 0.10 0.01 0.024 0.05 0.10	0.01 0.024 0.05 0.10 0.25 0.00 6.15 0.00 0.00 0.00 0.01 0.025 0.05 0.10 0.25 0.01 0.025 0.04 0.09 0.26 0.00 2.95 17.68 7.86 2.72 mL) 0.01 0.025 0.05 0.10 0.25 0.01 0.024 0.05 0.10 0.24	0.01 0.025 0.05 0.10 0.25 0.50 0.01 0.024 0.05 0.10 0.25 0.53 0.00 6.15 0.00 0.00 0.00 4.00 0.01 0.025 0.05 0.10 0.25 0.50 0.01 0.025 0.04 0.09 0.26 0.53 0.00 2.95 17.68 7.86 2.72 4.00 <i>mL)</i> 0.01 0.025 0.05 0.10 0.25 0.50 0.01 0.025 0.05 0.10 0.24 0.51	0.01 0.025 0.05 0.10 0.25 0.50 1.00 0.01 0.024 0.05 0.10 0.25 0.53 1.09 0.00 6.15 0.00 0.00 0.00 4.00 5.84 0.01 0.025 0.05 0.10 0.25 0.50 1.00 0.01 0.025 0.04 0.09 0.26 0.53 0.99 0.00 2.95 17.68 7.86 2.72 4.00 0.71 mL) 0.01 0.025 0.05 0.10 0.25 0.50 1.00 0.01 0.025 0.05 0.10 0.25 1.00	0.01 0.025 0.05 0.10 0.25 0.50 1.00 2.50 0.01 0.024 0.05 0.10 0.25 0.53 1.09 2.58 0.00 6.15 0.00 0.00 0.00 4.00 5.84 2.19 0.01 0.025 0.05 0.10 0.25 0.50 1.00 2.50 0.01 0.025 0.04 0.09 0.26 0.53 0.99 2.59 0.00 2.95 17.68 7.86 2.72 4.00 0.71 2.46 mL) 0.01 0.025 0.05 0.10 0.25 0.50 1.00 2.50 0.01 0.025 0.05 0.10 0.25 0.50 1.00 2.50 0.01 0.025 0.05 0.10 0.24 0.51 1.07 2.60	0.01 0.025 0.05 0.10 0.25 0.50 1.00 2.50 5.00 0.01 0.024 0.05 0.10 0.25 0.53 1.09 2.58 5.09 0.00 6.15 0.00 0.00 0.00 4.00 5.84 2.19 1.25 0.01 0.025 0.05 0.10 0.25 0.50 1.00 2.50 5.00 0.01 0.025 0.04 0.09 0.26 0.53 0.99 2.59 5.43 0.00 2.95 17.68 7.86 2.72 4.00 0.71 2.46 5.60 mL) 0.01 0.025 0.05 0.10 0.25 0.50 1.00 2.50 5.00 0.01 0.025 0.05 0.10 0.25 0.50 1.00 2.50 5.00 0.01 0.024 0.05 0.10 0.24 0.51 1.07 2.60 5.01

#### **Conclusions**

Improved LC/ESI-MS sensitivity can be observed when utilizing a UPLC<sup>™</sup> as an MS inlet with traditional 5 µm particles. Sensitivity can be further improved upon when using 1.7 µm particles yielding a 5X improvement in sensitivity at the same peak capacity and is achieved 8X faster compared to traditional 5 µm particles. Additionally, the utilization of 1.0 mm i.d. columns packed with 1.7 µm particles yields significant improvements in the capabilities of reaching extremely low