

# Waters

**Application of LC/MS(MS) to  
metabonomics studies**

**Robert Plumb, 8<sup>th</sup> Oct 2002 CPSA2002**



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MS TECHNOLOGIES

- *Metabonomics*
- *Advantages and disadvantages of LC/MS vs NMR*
- *LC/MS methodology*
- *Data analysis a test case*
- *Identification of biomarkers*
- *What do we do with polar compounds*
- *Conclusions and Acknowledgements*

- *Study of endogenous metabolites rather than xenobiotics*
- *Primary aim is the identification and quantitation of small molecules in a biological system*
- *Down-regulation and Up-regulation of these metabolites are indicative of biological insults due to disease, toxicity, genetic modification or environmental factors*
- *Knowledge of these metabolites and possible biomarkers, may be used for diagnosis and screening*

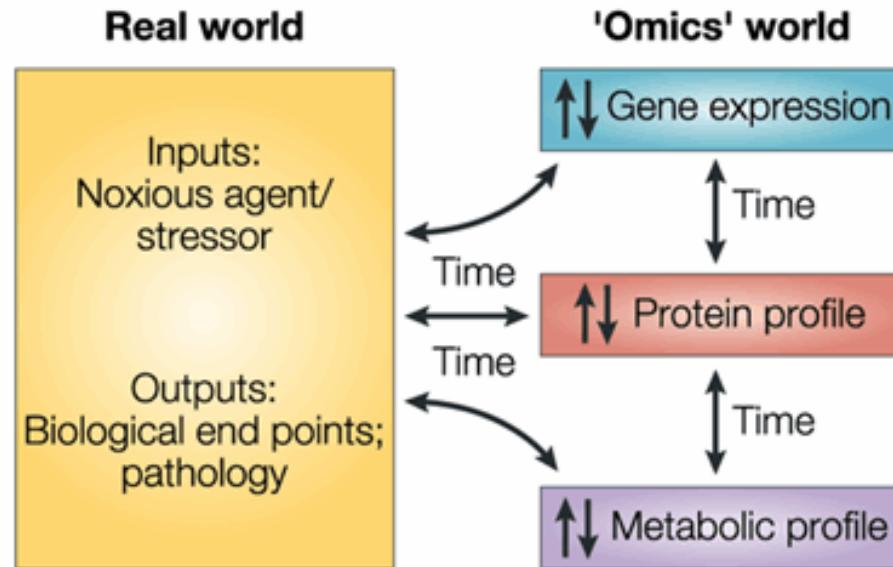
## Metabonomics

**“Quantitative measurement of time-related multiparametric metabolic responses of multicellular systems to pathophysiological stimuli or genetic modification”.**

## Metabolomics

**“The measurement of metabolite concentrations and fluxes in isolated (and usually identical) cell systems or cell complexes”.**

# Real World vs the "Omics" World

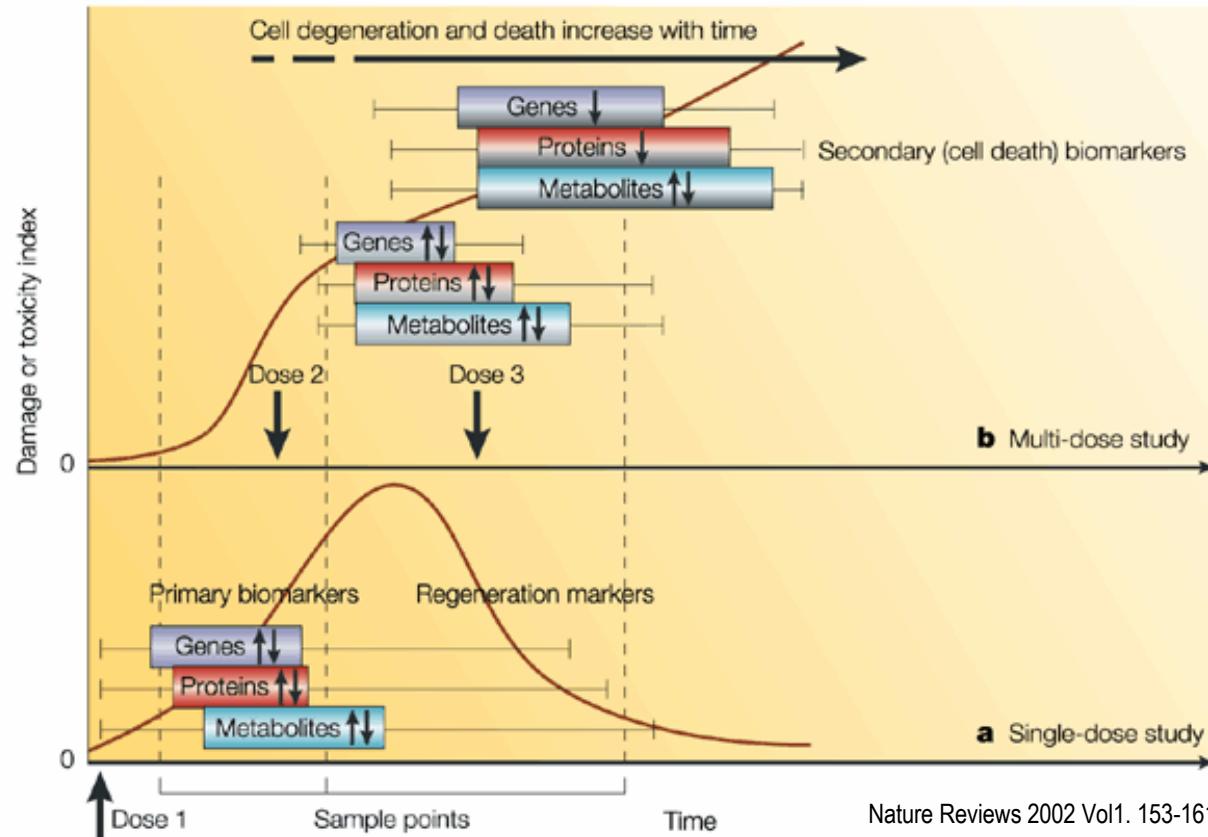


Nature Reviews 2002 Vol1. 153-161

*"Metabolic changes are real world end points, whilst gene expressions indicate the potential for an end-point change"*

(J Nicholson Nature 2002)

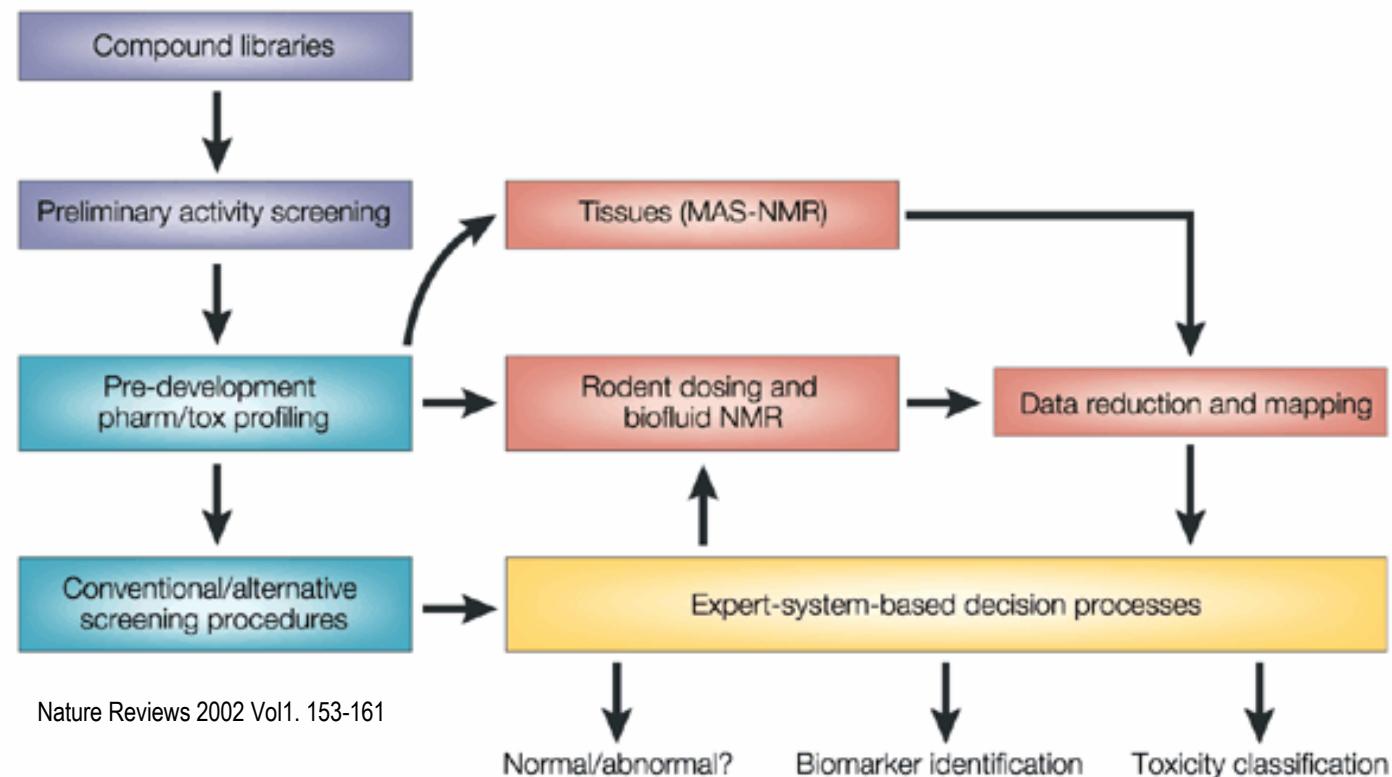
# Effect of Time in Toxicological Studies



*The “Omics” response varies as cell degeneration increases*

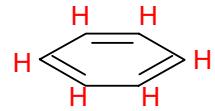
(J Nicholson Nature 2002)

# Metabonomic Integration for Toxicity in DMPK



*"Integration of data types will also pave the way to understanding the relationship between gene function and metabolic control in health and disease"*

## AROMATICS

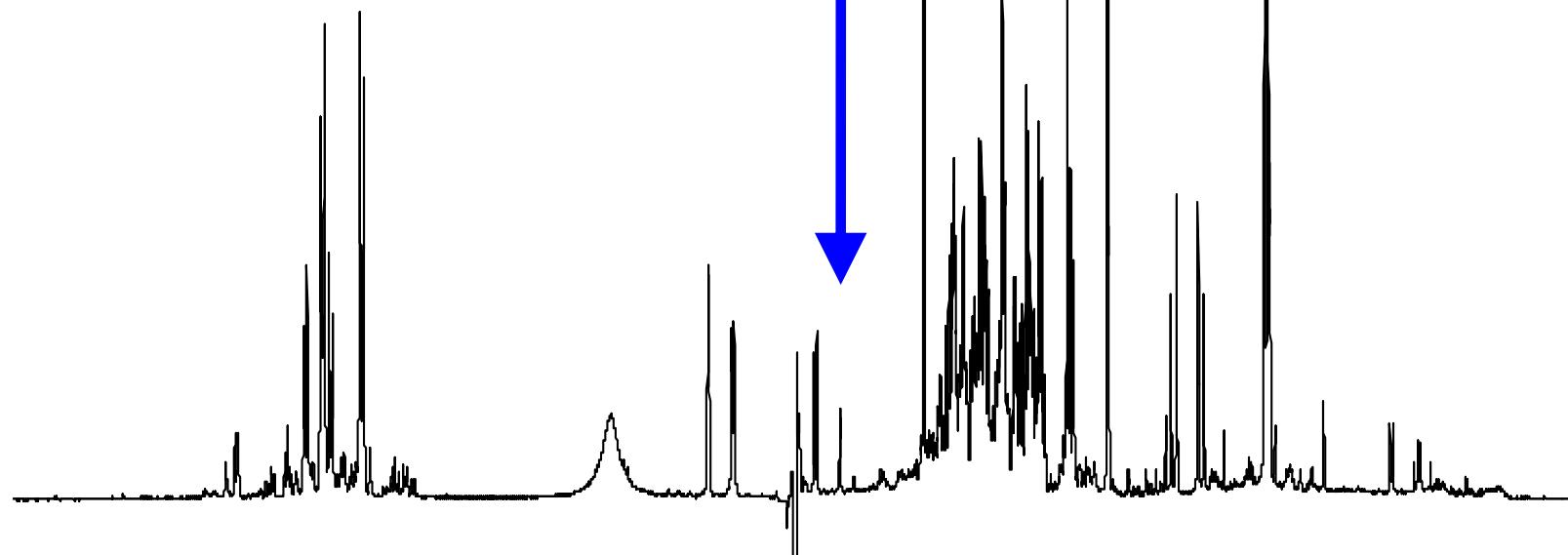
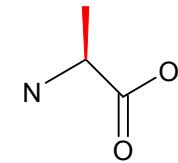


Xenobiotic

## SUGARS



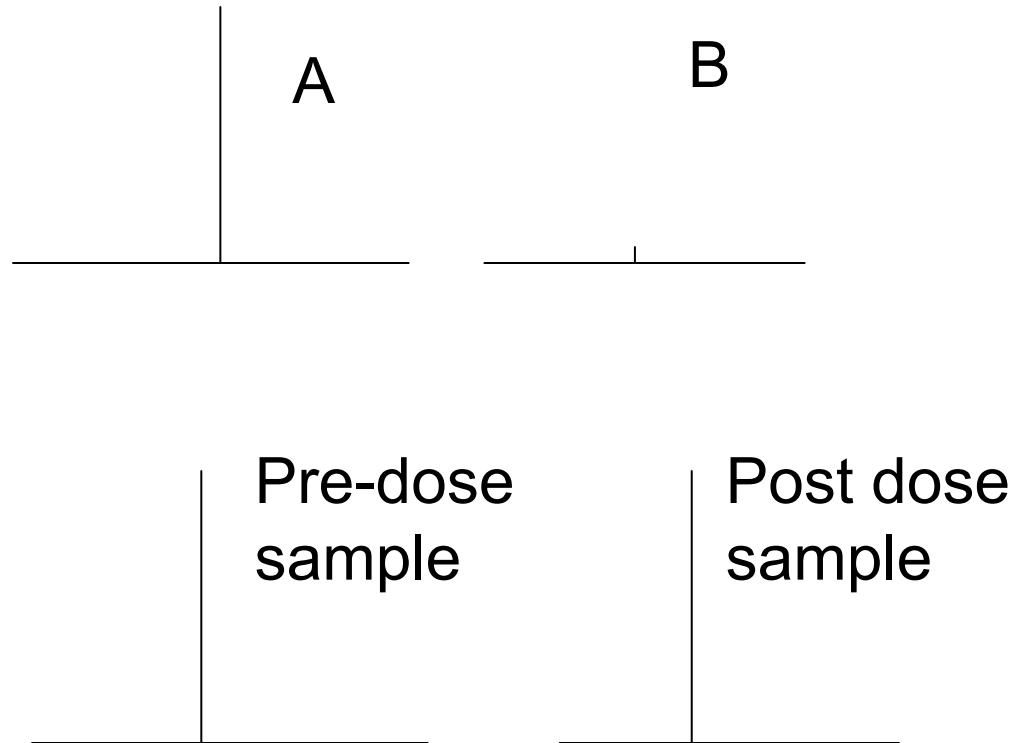
## ALIPHATICS



- When removing xenobiotics NMR data is lost. This can be replaced with archive control data.
- But what if there was a biomarker is in this region that had been up/down regulated ?
- This also takes time and assumes that you have an archive of data for every dosing vehicle combination, DMSO, PEG, Tween 80 etc..

- Scenario:-

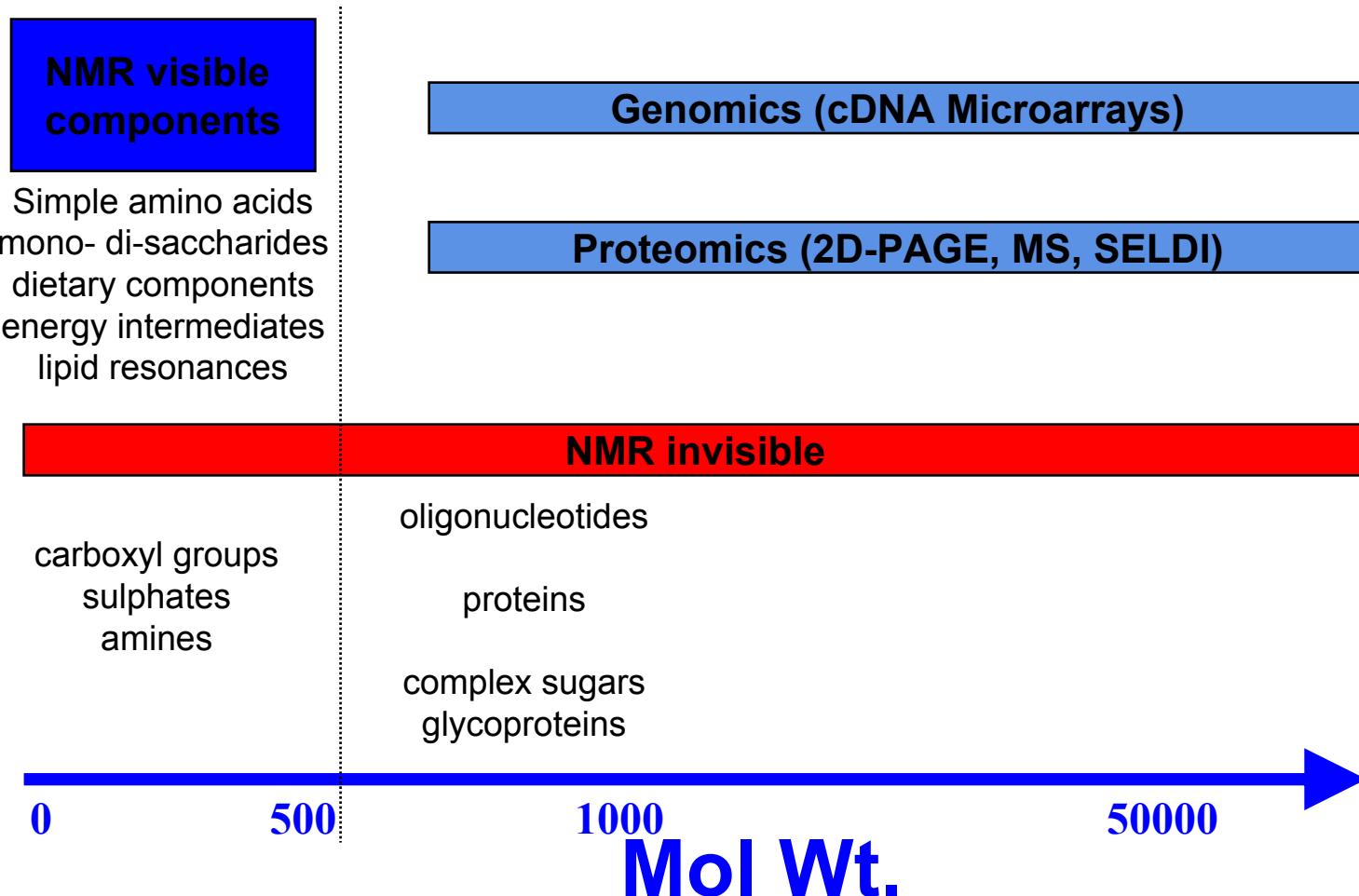
- Small abundance component appears in the same region of the NMR spectra as a large concentration component.
- The peak height ratio is 1:10
- Natural variation in large component is +/- 15%
- After dosing B is 50% down regulated.



- Result:-

- Variation in B never observed

# NMR is the Current Method of Choice, but with Limitations.



Provided by John Haselden (GSK)

- *20 rat urine samples*
- *Two time points: 0-8 hr and 8-24hr*
- *3 x 2 controls*
- *3 compounds, 3 samples per time point, 2 time points*
- *Simple Sample Prep: Samples centrifuged then diluted 1:4 with distilled water prior to analysis by LC/MS*

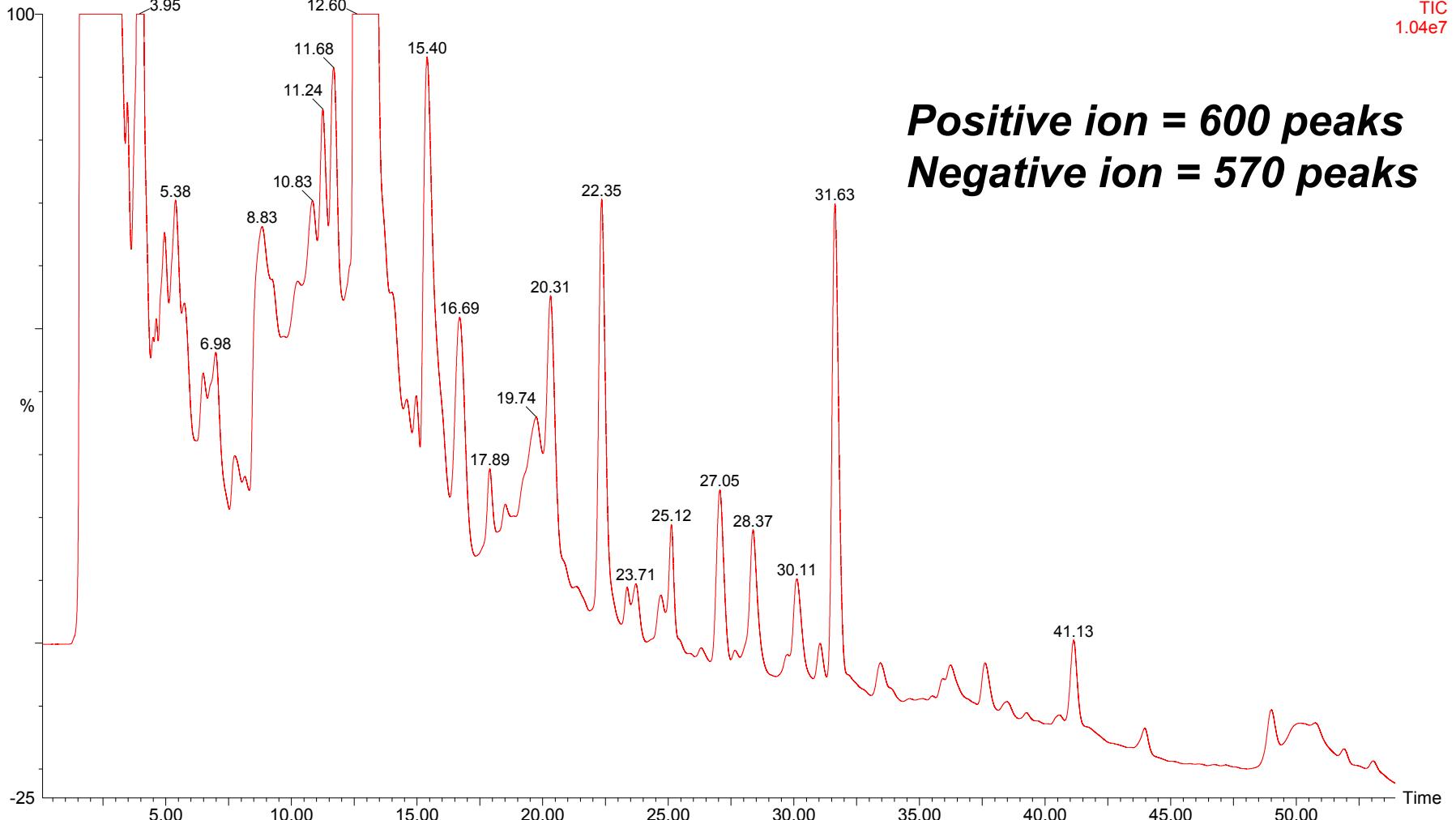
- *Chromatography:*

- Column: Waters Xterra<sup>®</sup> C18 100 x 2mm 3.5µm
  - Eluent: Reverse Phase
  - Flow Rate: 600 uL/min (slower for LC/MS/MS)

- *Mass spectrometry*

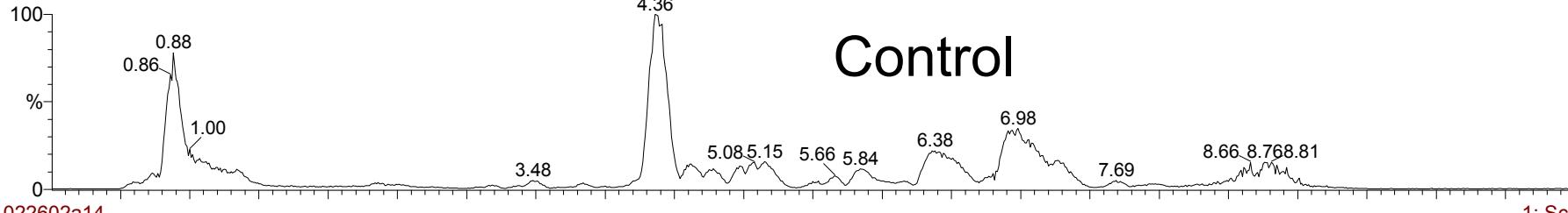
- Quattro micro<sup>TM</sup> or Q-Tof micro<sup>TM</sup>
  - Scan range 100-1000m/z
  - Data collected from 0-10mins (30 minutes for LC/MS/MS)

## Typical urine 60 minute chromatogram

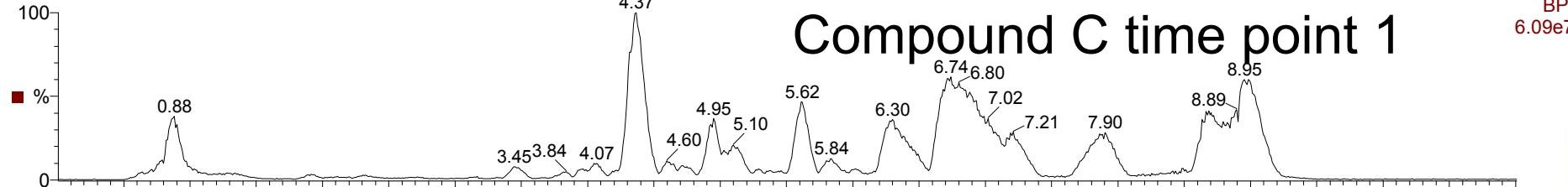
Rat urine  
D072902a09

## LC/MS TIC data: Qualitative Differences

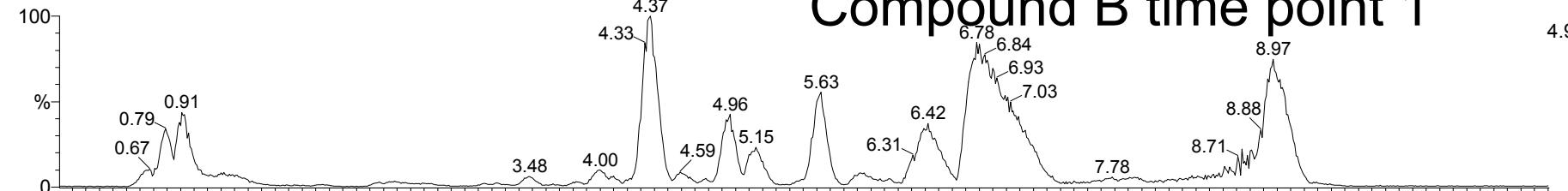
022602a07



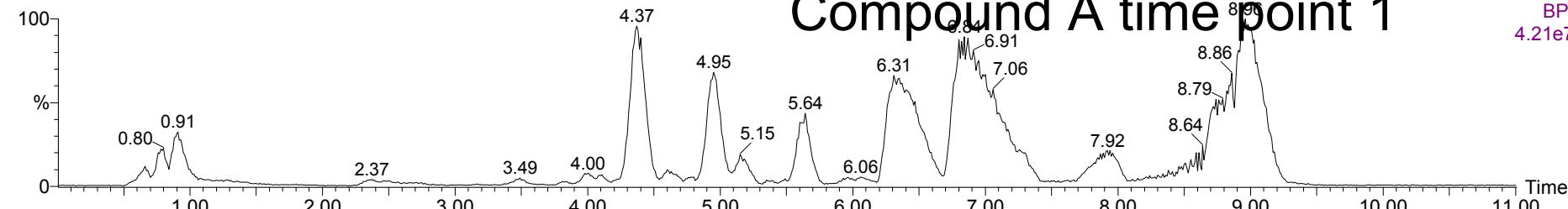
022602a14



022602a12

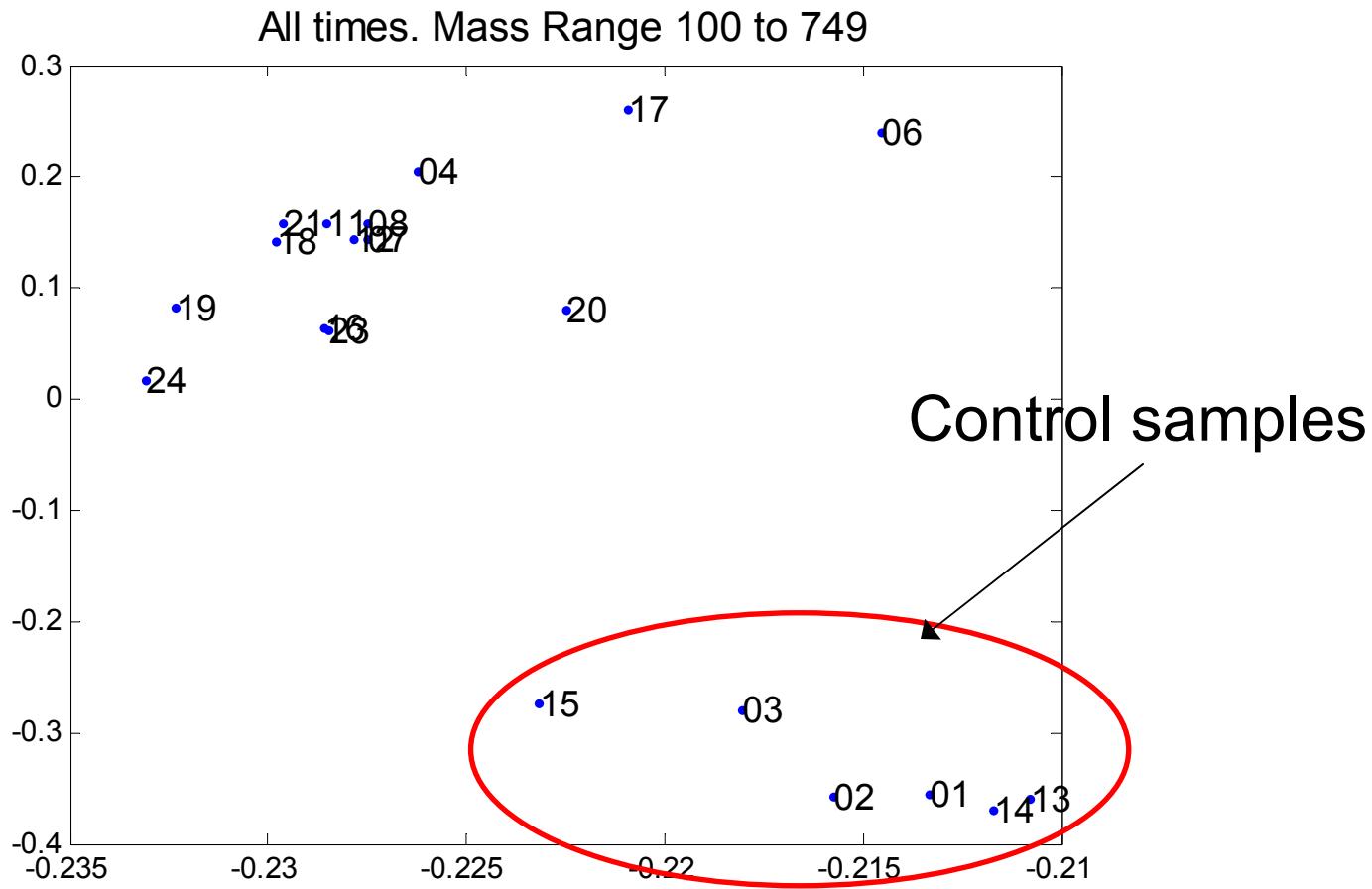


022602a10

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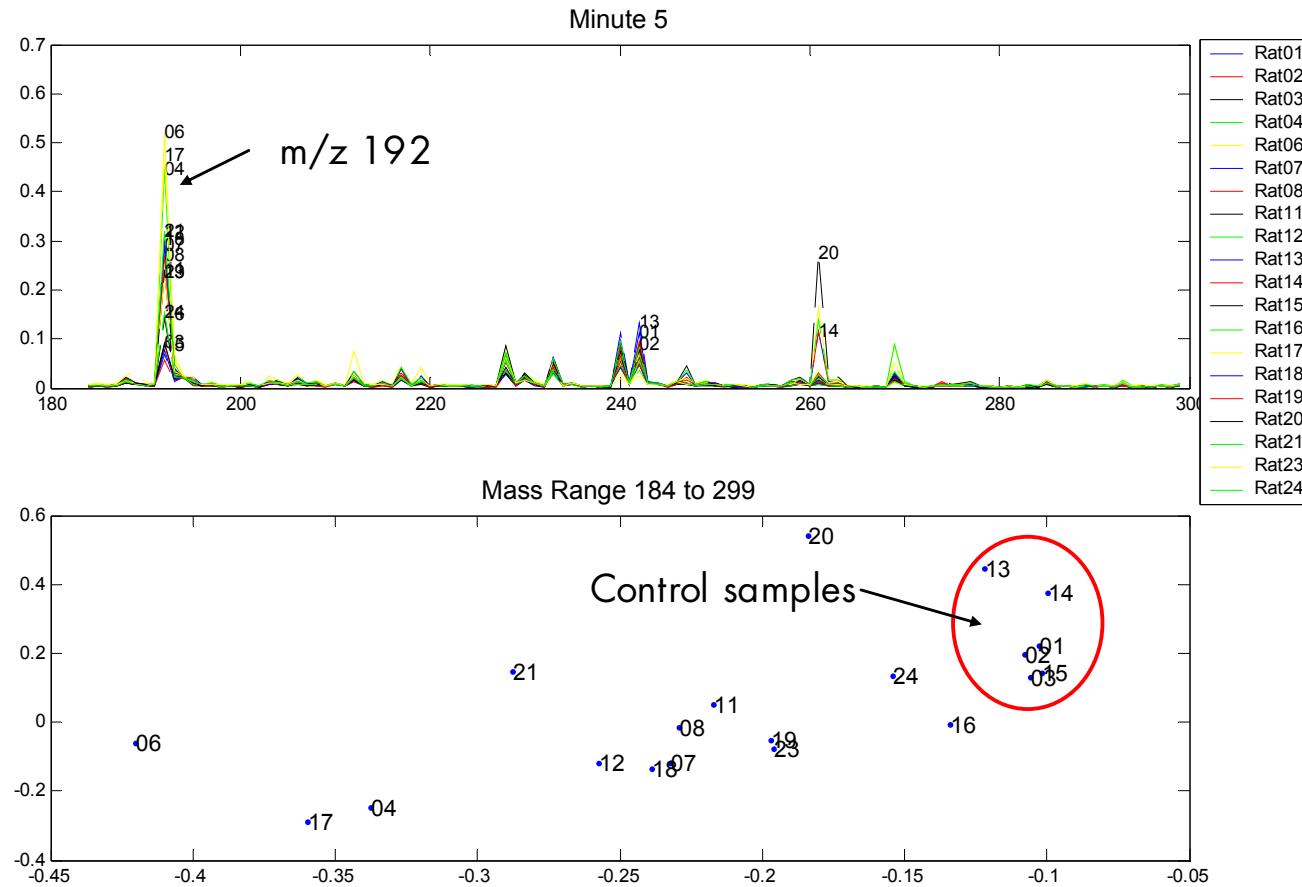
- *BioMarkers can be found by analyzing the Mass Spectra one-by-one. However this is cumbersome.*
- *Principal Component Analysis is a Multi-variate analysis technique that provides a Global View of the Data! (Potentially this could be faster).*

MassLynx -> Excel -> MatLab (PCA)



Data from March 2002

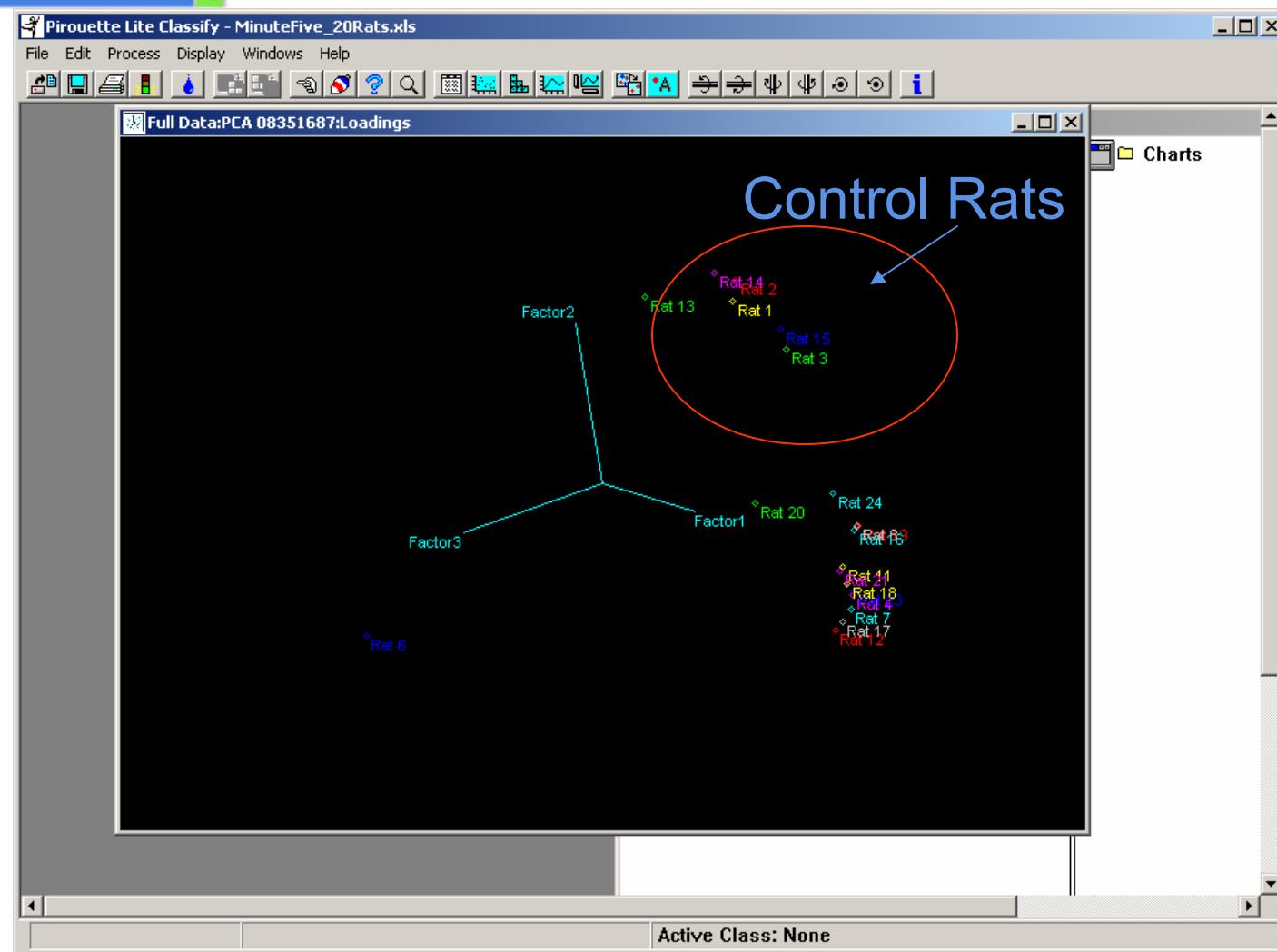
## PCA investigation from Minute 5 of Chromatogram



Note: LC Retention Information is Utilized!

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# Pirouette Loadings Plot: Minute 5

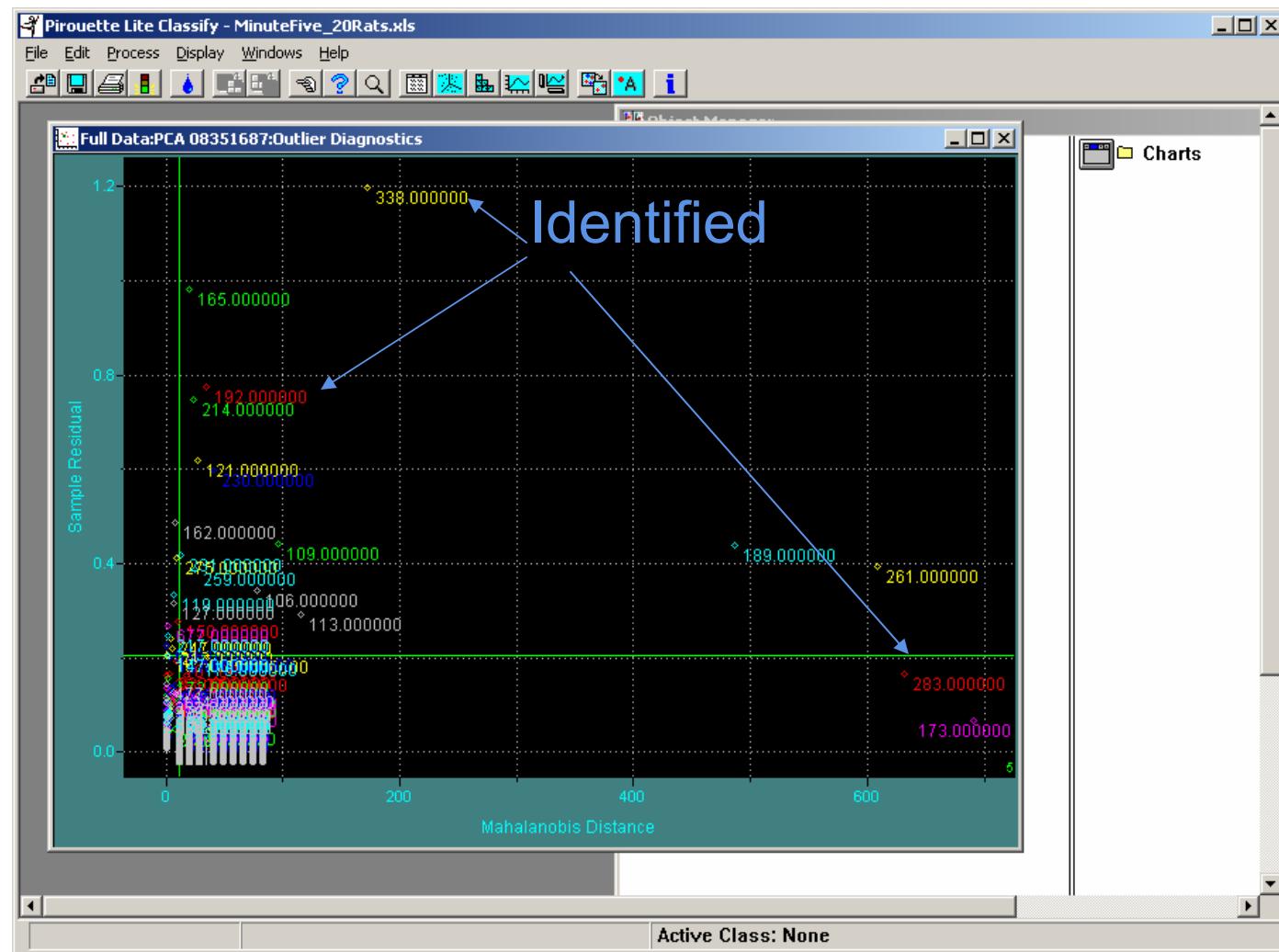


## Ions identified as responsible for PCA separation

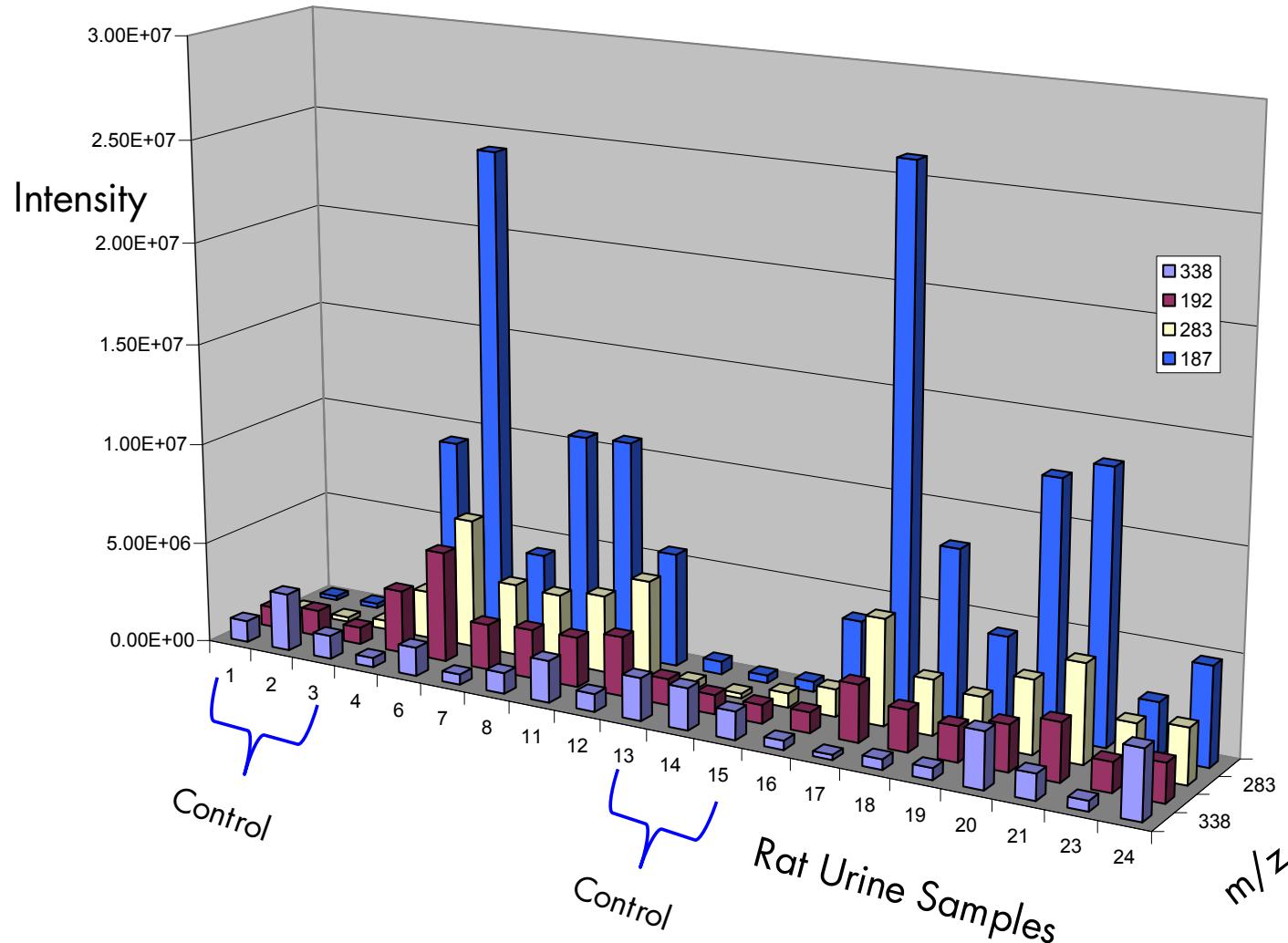
Compound dosed	Analyte m/z value	Change
A	283	10 fold increase
A	461	5 fold increase
A	187	10 fold increase
B	338	2 fold reduction
B	283	10 fold increase
B	461	10 fold increase
B	187	10 fold increase
C	283	20 fold increase
C	187	30 fold increase
A,B,C	192	3 fold increase

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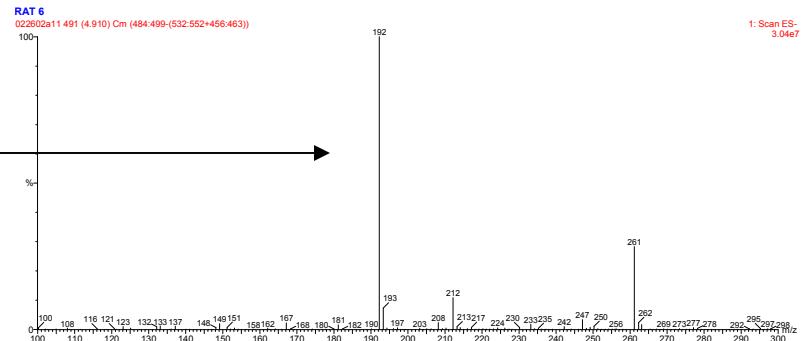
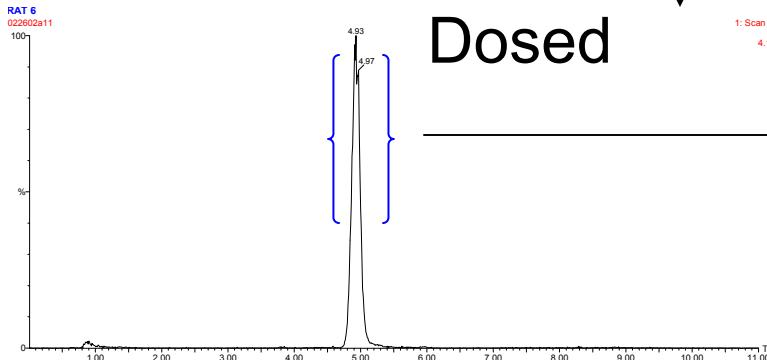
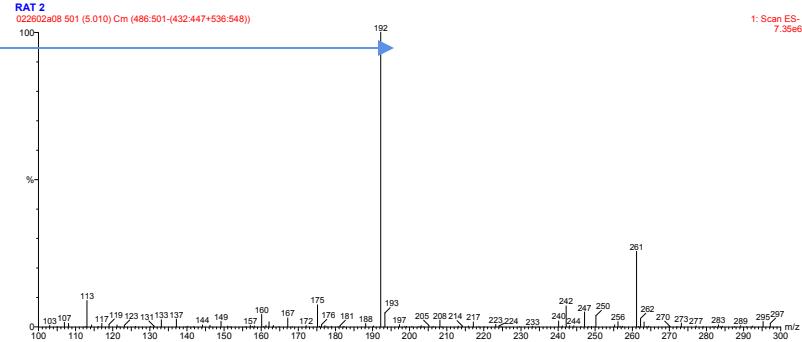
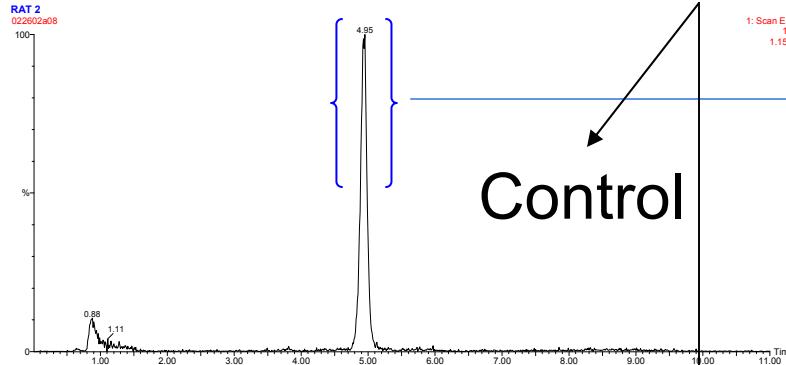
BioMarkers Indicated!



# Ion intensity comparison of PCA identified ions ( $m/z$ 187, 192, 283, 338) in all 20 rat urine samples.

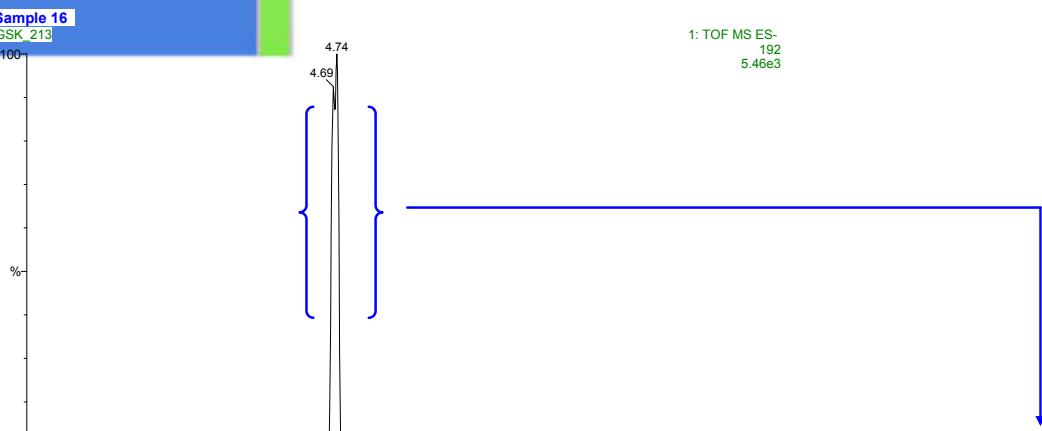


## Endogenous Metabolites

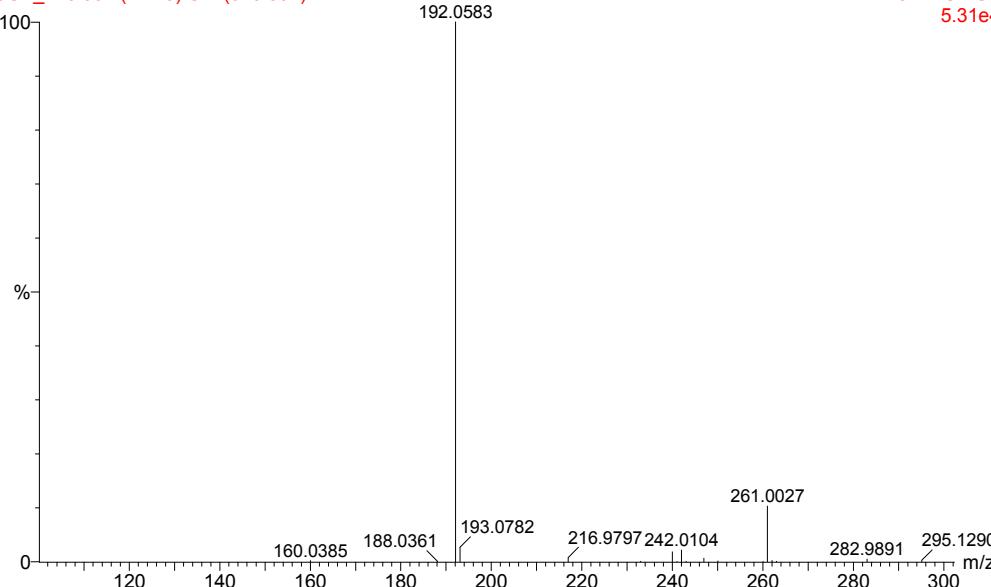


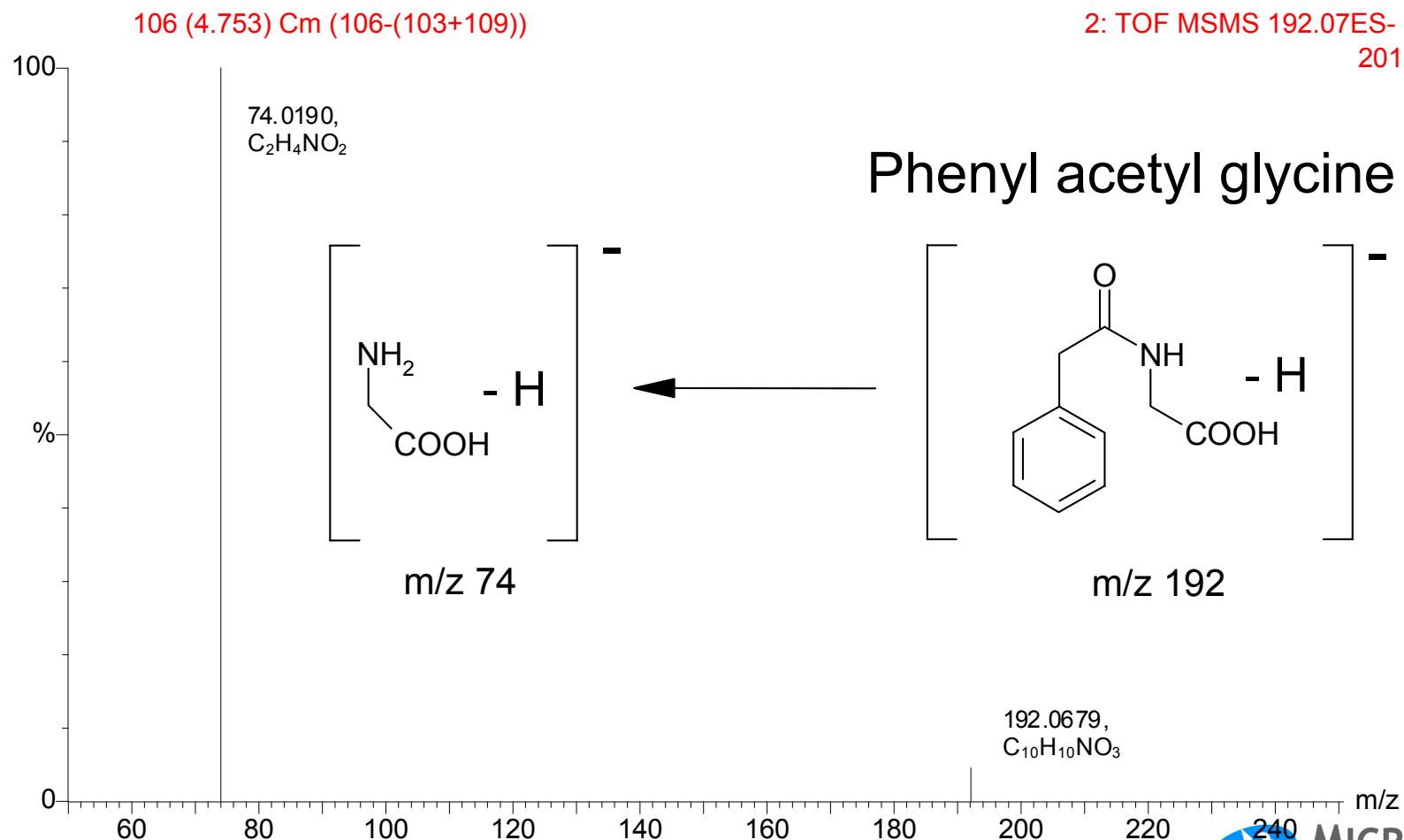
# LC/MS of 192 ion using a Q-Tof

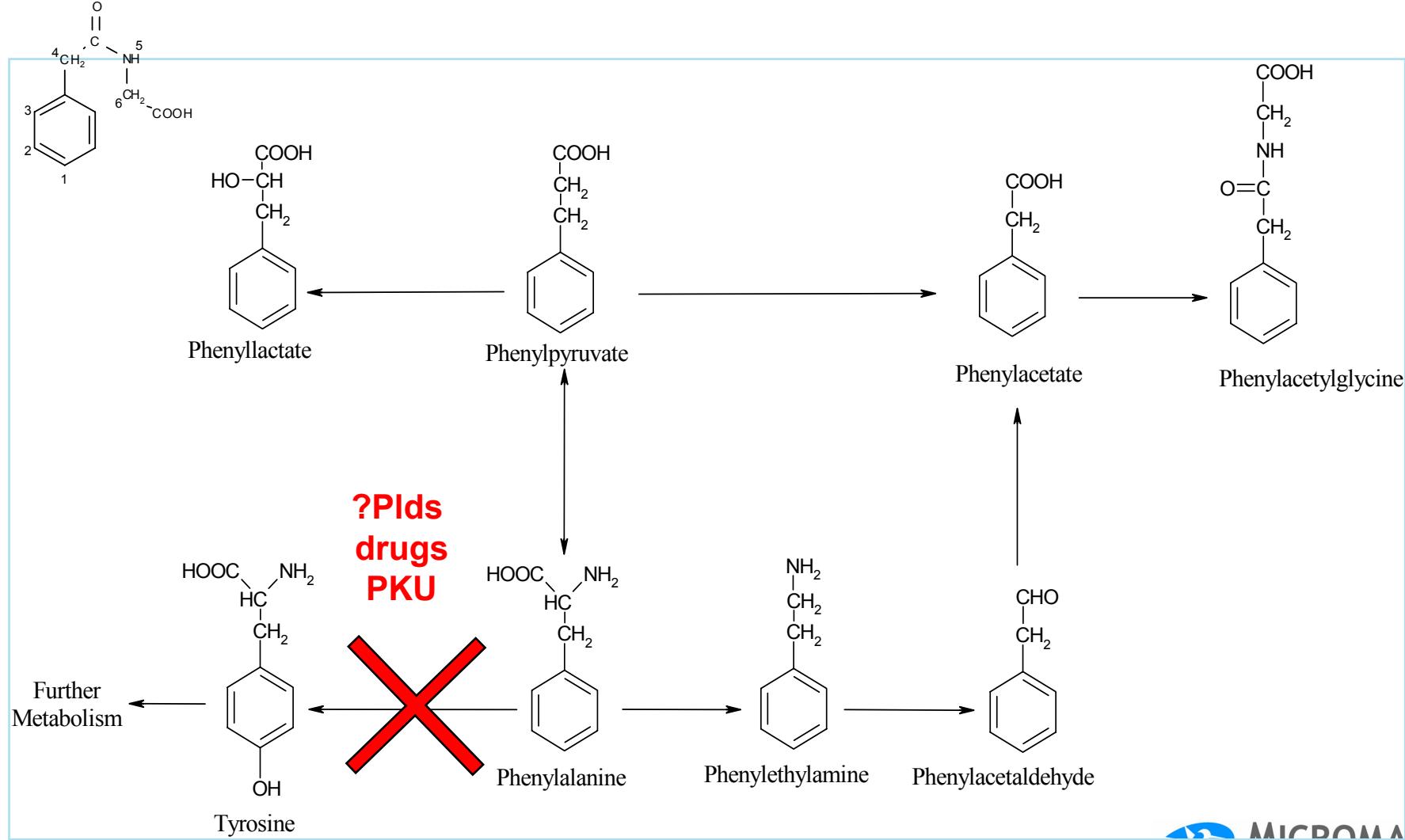
Accurate mass information obtained for the 192 ion was used to determine the elemental composition  
 $C_{10}H_{10}NO_3$ .

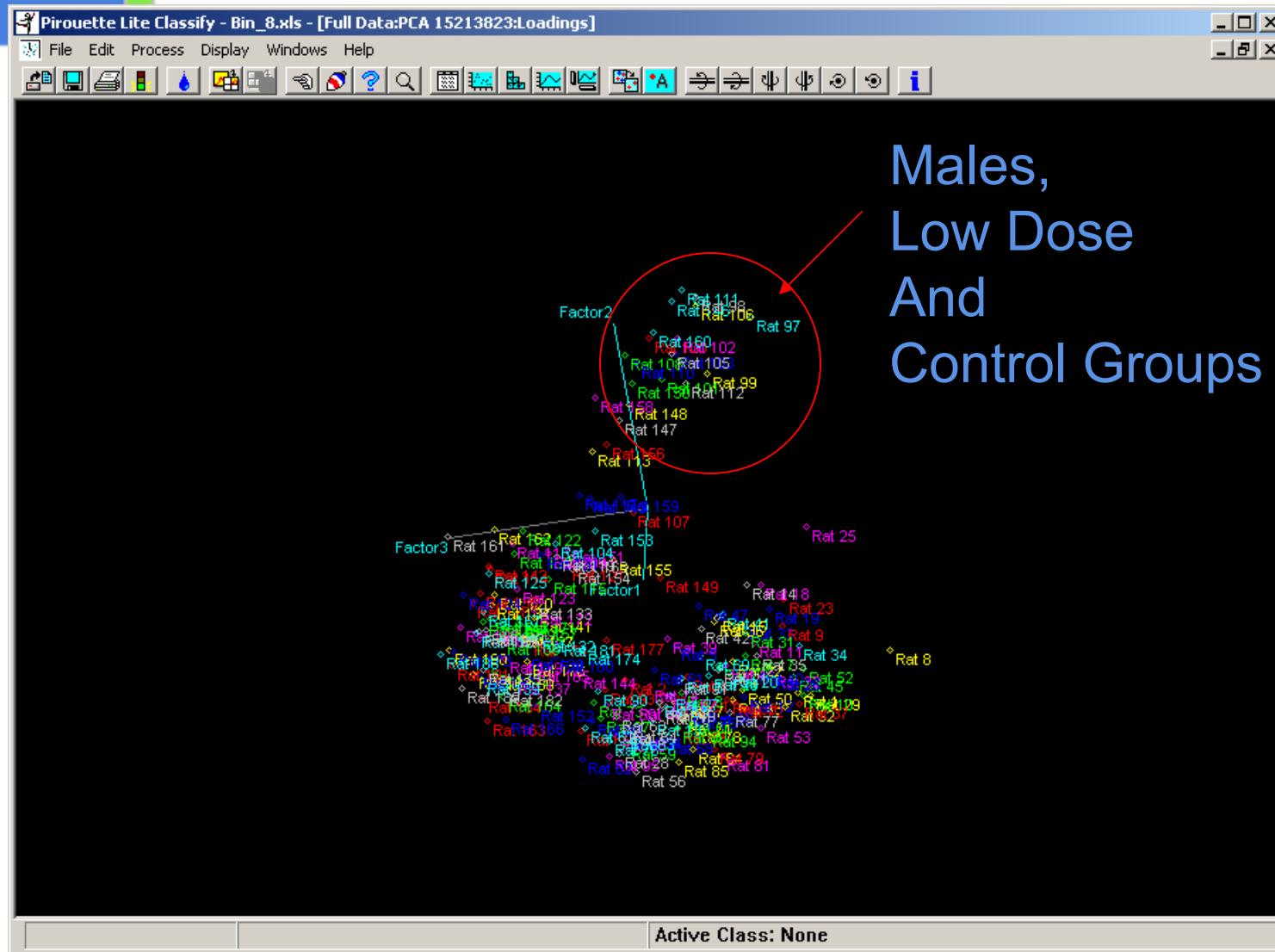


Sample 16  
GSK\_213.mz (4.743) Cm (370:392)



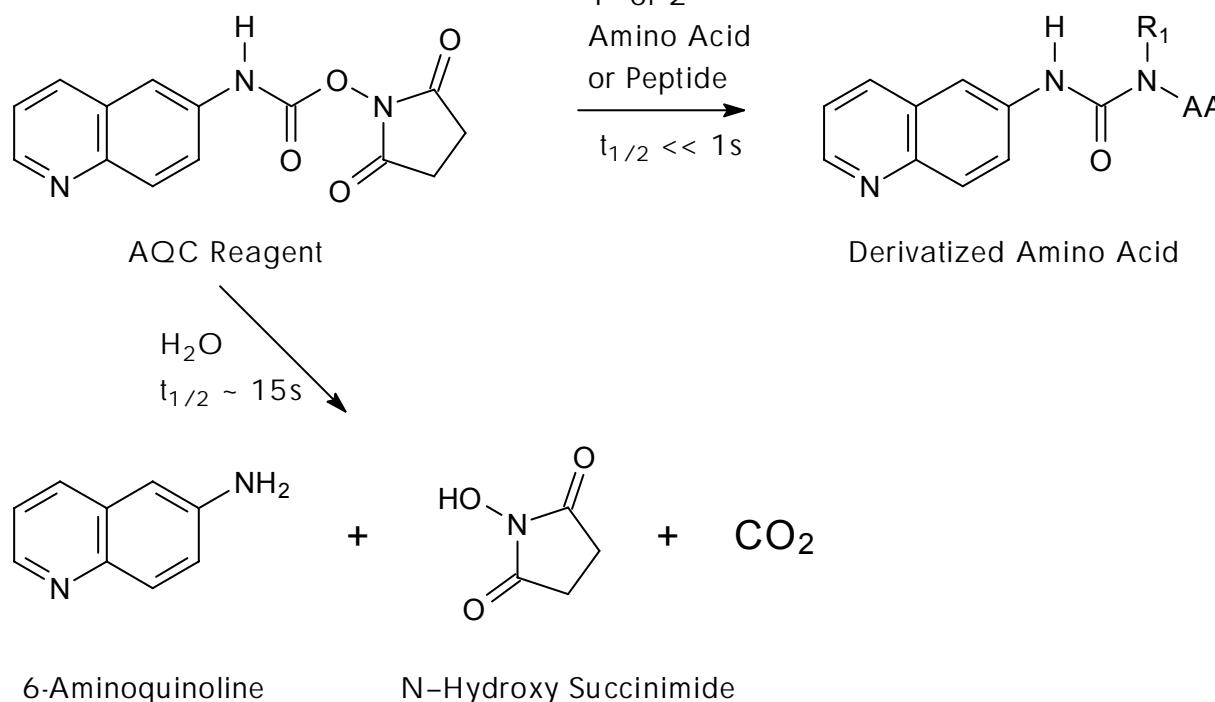






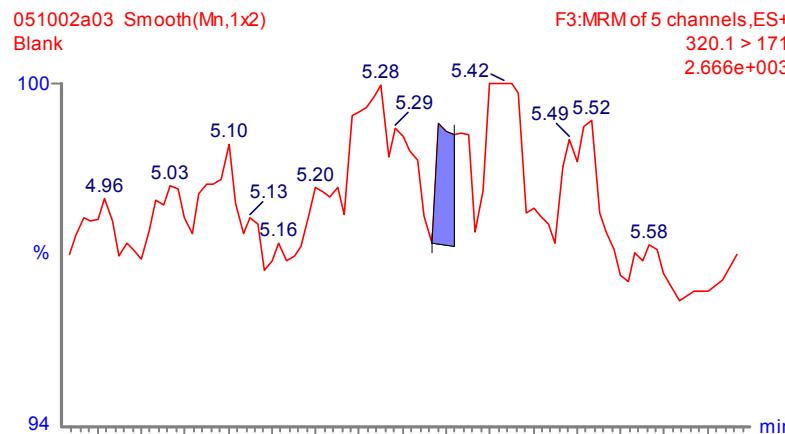
- These are typically very difficult to analyse by reversed-phase LC/MS, eluting at the solvent front and suffering ion suppression.
- Using alternative LC methodologies such as HILIC would mean that more than one analytical run is required.
- This would not necessarily ensure that the polar compound is detected.
- These compounds can be important biomarkers of toxicity, e.g. amino acids.

- Amino acids are unretained on reversed-phase LC systems.
- Potential solution to use Waters AccQ-Tag with MS detection.
- Sample prep:- evaporate sample add buffer add derivatizing reagent.
- Chromatograph by gradient LC/MS/MS, injection volume 2 $\mu$ L. Using MRM analysis for 20 amino acids
- Run times 10 mins per sample.

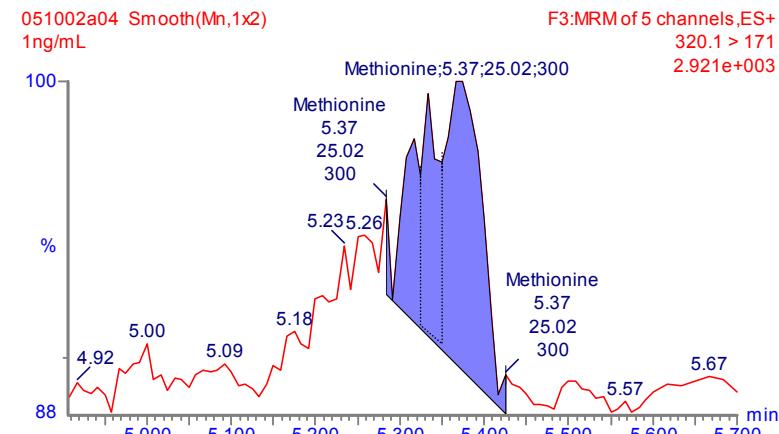


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# Derivitization of Methionine



Blank



1ng/mL



50ng/mL std

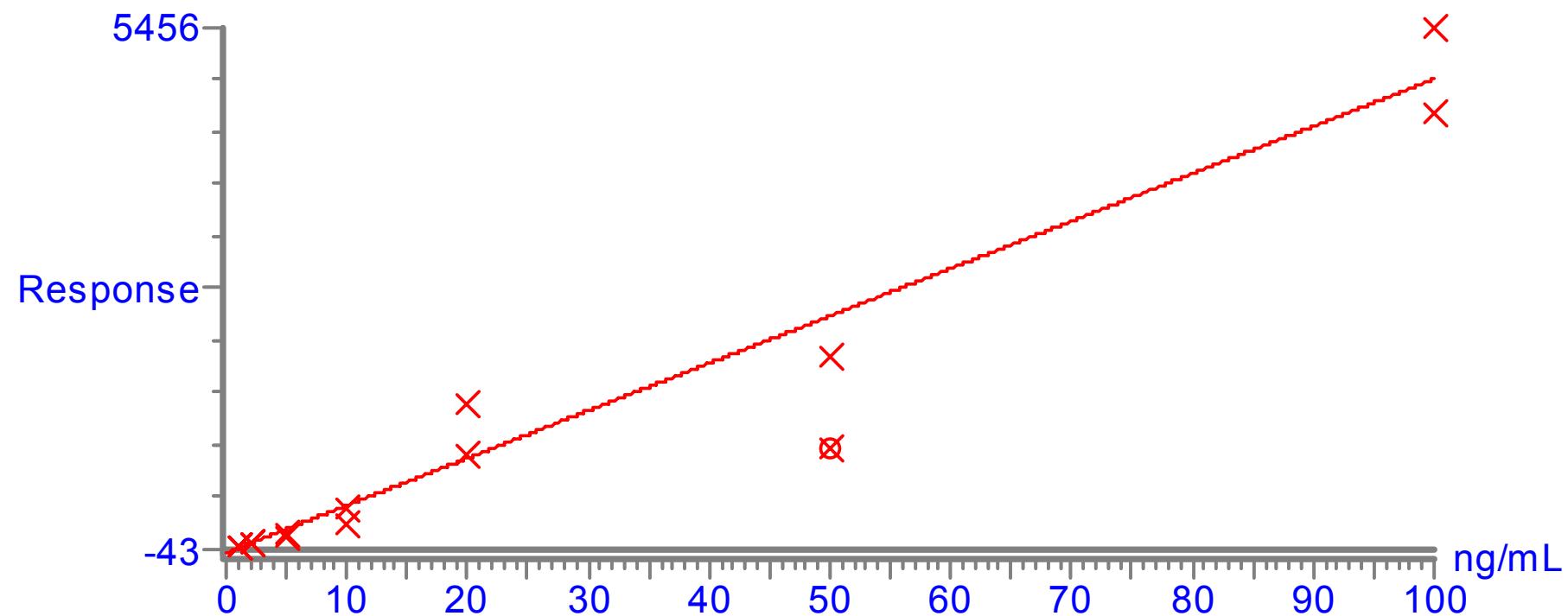
Compound name: Methionine

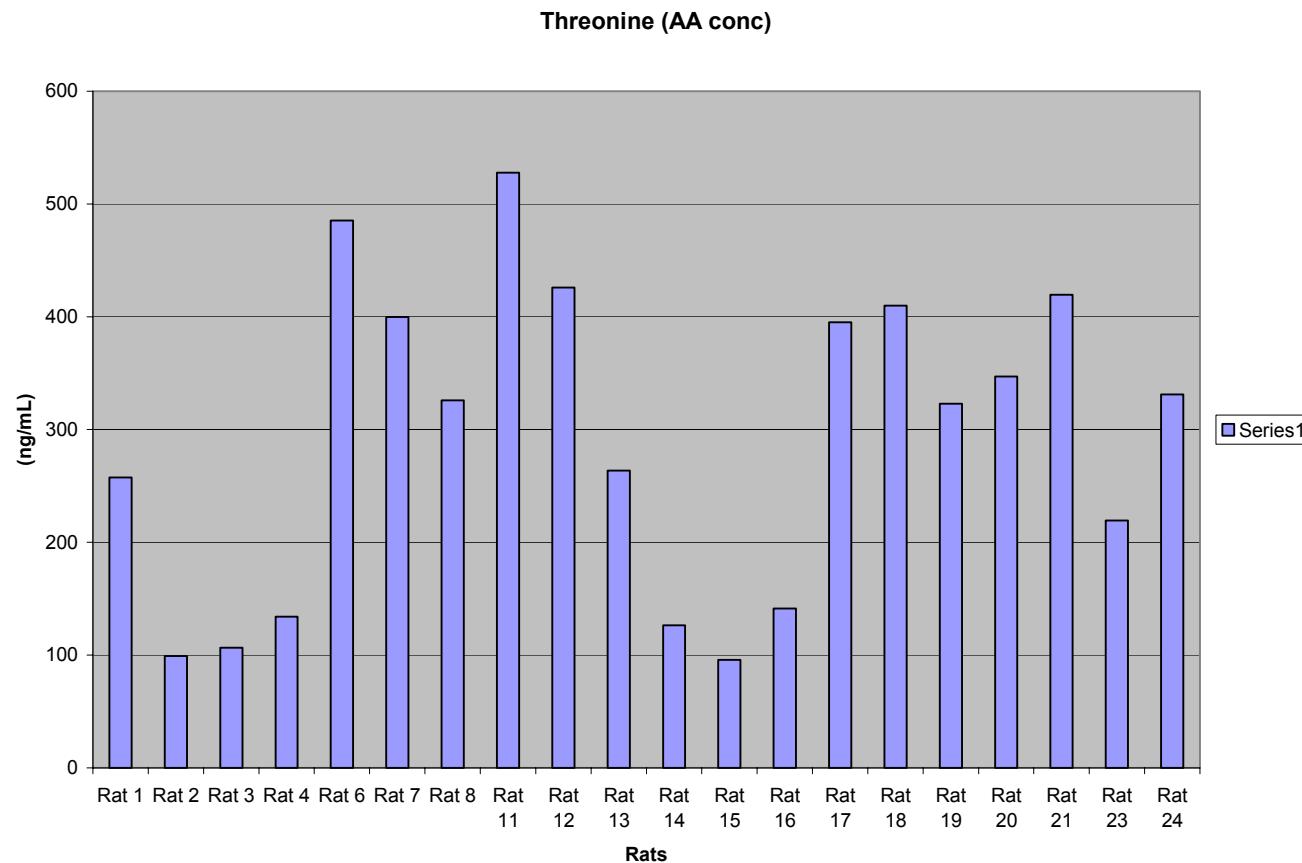
Coefficient of Determination: 0.976096

Calibration curve:  $49.6451 * x + -42.9708$

Response type: External Std, Area

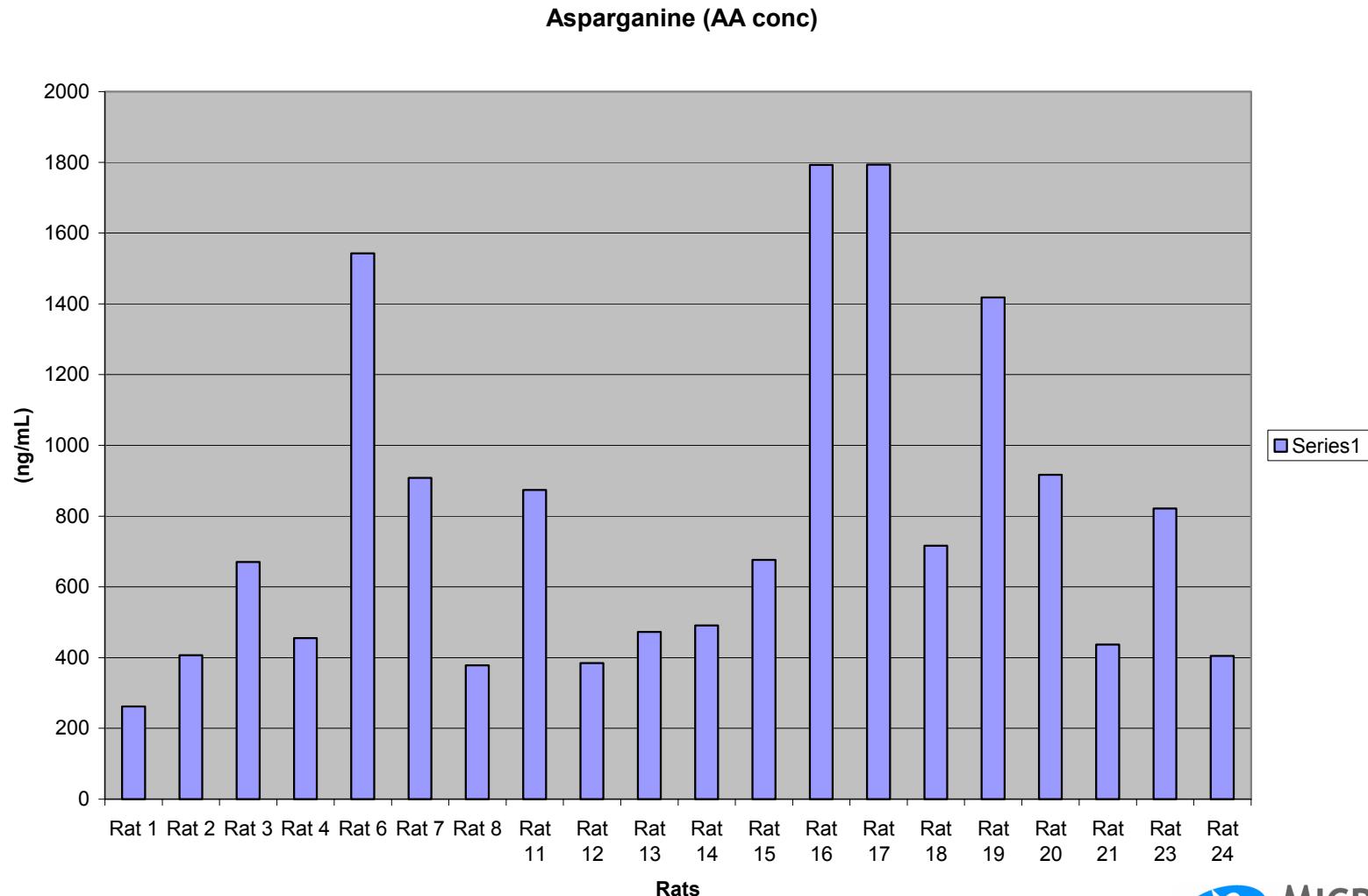
Curve type: Linear, Origin: Exclude, Weighting:  $1/x$ , Axis trans: None





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Asparganine



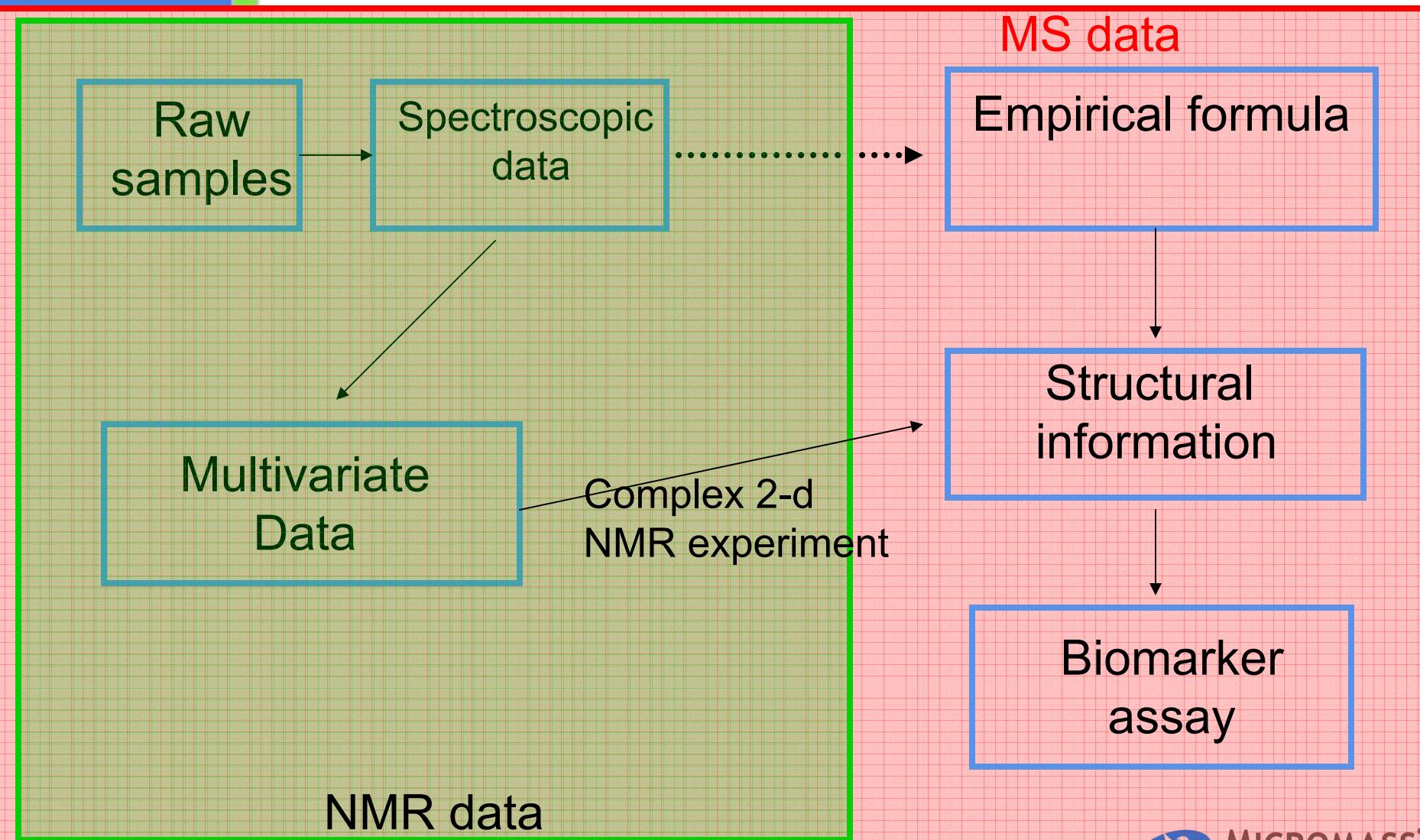
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# Amino acid summary

Amino Acids	Retention Window (min)	Max Conc (ng/mL)	M/Z
Glycine	3.25-3.9	820	246.1
Serine	3.1-3.85	340	276.1
Glutamine	3.5-4.0	2200	317.1
Histidine	3.5-4.0	810	326.1
Alanine	3.7-4.25	1450	260.1
Proline	4.125-4.5	1630	286.1
Threonine	3.5-4.125	540	290.1
Taurine	3.5-4.25	64000	296.1
Valine	4.85-5.25	325	288.1
Methionine	5.125-5.65	1800	320.1
Asparganine	5.125-5.5	1800	321.1
Tyrosine	4.75-5.25	270	352.1
Cystine	4.75-5.25	50	581.3
Leucine	5.5-6.0	460	302.1
Phenylalanine	6.0-6.5	850	336.2
Tryptophan	6.0-6.5	1150	375.1
Lysine	5.75-6.25	1650	487.2
Homocysteine	5.75-6.25	1.6	609.3

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Data flow



- *With a relatively simple reversed-phase LC/MS system it was possible to separate the control from dosed samples.*
- *m/z value of analytes responsible for the PCA separation were identified.*
- *Analytes responsible for PCA separation were identified by LC/MS/MS and TOF MS.*
- *AccQ-Tag<sup>TM</sup> was successfully employed to analyse amino acid in urine.*



## Acknowledgements

- *John Haselden, Safety Assessment, GlaxoSmithKline, Ware, UK,*
- *Gordon Dear, DMPK, GlaxoSmithKline, Ware, UK.*
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- *Hilary Major, Micromass, Manchester, UK.*