

Oligonucleotide synthesis yield

Although state-of-the-ort oligonucleotide synthesis is a fast and reliable process, the yield of the target product is limited. The full-length oligonucleotide (N) is typically contaminated with shorter failure products called N-1, N-2, N-3... Oligonucleotide purity decreases with the complexity of synthesis. The typical purity of crude 25mer product is ~ 75 % while the purity of a 60mer product is ~ 20 %. Occasionally the synthesis or post synthesis deprotection result in a product of inferior purity, which may compromise quality of the polymerase chain reaction (PCR) or other molecular biology assays.

XTerra[®] columns for efficient oligonucleotide separations

XTerra[®] MS C₁₈ columns are packed with porous 2.5 μ m, hybrid particles. The sorbent has extended stability at temperatures and pH's typically used for oligonucleotide separations (50-60° C; pH 7-9). The column yields high resolution efficiency required for successful analysis of oligonucleotides (*Figure 2*). Volatile ion-pairing mobile phases are utilized to separate oligonucleotides according to their length. The resolution performance rivals that seen with capillary gel electrophoresis (*Figure 1*). With XTerra[®] MS C₁₈ columns, baseline resolution of N from N-1 oligonucleotides is typically achieved for <30mer. Good resolution can be achieved for larger oligonucleotides (e.g. 60mer or larger: see (*Figures 2 & 3*)).







Figure 2. RP-HPLC quality control of synthetic oligonucleotides. Purity was measured as the % of peak area at UV_{260 nm}.

Triethylammonium acetate (TEAA) is an inexpensive volatile ion-pairing agent used for routine oligonucleotide analysis **(Figure** 2) and purification. Alternatively, triethylammonium-hexafluoroisopropanol buffer (TEA-HFIP) can be used for more efficient separation of long oligonucleotides (>45mer, **Figure 3**), and for applications using mass spectrometry detection.

Preparation of 0.1 M TEAA buffer: Dissolve 5.6 mm glacial acetic acid in ~950 ml of water. While mixing add 13.86 ml of TEA. Adjust pH with diluted acetic acid to ~ 7 and adjust volume to 1L with water.

Preparation of 16.3 mM TEA - 400 mM HFIP buffer: Dissolve 42.1 ml of HFIP in ~950 ml of water. While mixing vigorously add 2.28 ml of TEA. Adjust volume to 1L with water. The pH of solution should be close to 7.9.





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