

# Performance of an Ultra Low Elution Volume 96-Well Plate

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Achieving both high throughput and low detection limits (LOQ's) is now the challenge of pharmaceutical analysis. Since the mid 90's, additional emphasis has been directed on the sample preparation at sub ng/mL levels and extraction speed. Classical extraction techniques, like liquid-liquid extraction (LLE) or protein precipitation (ppt), are not suitable formats for high throughput for good clean-up. For example, LLE gives better recovery and clean-up requires extensive manual

labor and is not easily automated. Ppt, on the other hand, is fast and can be automated, but the extract is not as clean as LLE or SPE techniques (potential loss of signal due to ion suppression). As more drug candidates shows increased levels of potency, less dosage is required, hence the need for lower LOQ's in order to define the pharmacokinetic profile. During pre-clinical trials of a drug candidate, sample volume is relatively low, most common in the 50  $\mu\text{L}$  range. Clearly, using solid phase extraction (SPE) in a 96-well plate format

(1,2) can keep up with these new demands for low sensitivity using less sample volume. With the assistance of robotic liquid handler, a 96-well plate can be prepared in less than a minute per sample (3). The performance of a new 96-well ultra low elution volume plate for LC/MS/MS applications will be summarized. Each well in the plate contains 2 mg of a high capacity solid phase extraction polymer (Oasis<sup>®</sup> HLB and MCX) (4) that has the capacity of extracting analytes from several hundred microliters of plasma. A novel tip design permits quantitative

quantitative recovery of the extracted analytes with as little as 25 microliters of elution solvent.

Therefore, the plate provides as much as 5-fold concentration of the analyte from plasma. The plate allows for simple dilution of the elution solution with water prior to LC/MS/MS analysis, thus avoiding an evaporation and reconstitution step.

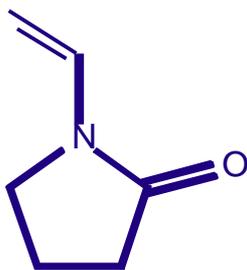
1- Zweigenbaum, T. Heing, K., Steinborner, S. Wachst, T., Henion, J., Anal. Chem., 1999, **71**, 2294

2- Show, W.Z., Jiang, X., Beato, B.D., Naidong, W., Rapid Commun. Mass Spectrom., 2001, **15**, 466

3- Jemal, M., Huang, M., Mao, Y., Whigan, D., Powell, M., Rapid Commun. Mass Spectrom., 2001, **15**, 994

4- Cheng, Y.F., Neue, W., Bean, L., J. Chromatogr. A, 1998, **828**, 273

# Hydrophilic-Lipophilic Balanced copolymer



N-Vinyl-Pyrrolidone



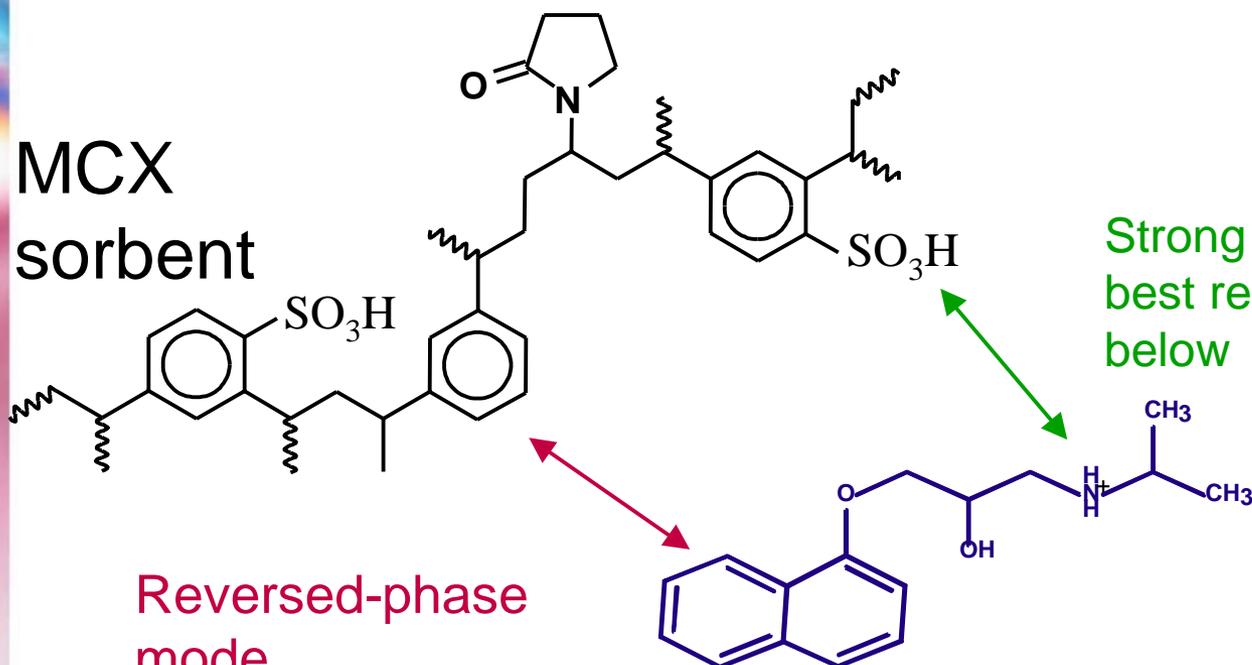
Di-Vinyl-Benzene

**Water Loving**  
Hydrophilic monomer

- Provide wetting properties
- No impact of sorbent drying

**Fat Loving**  
Lipophilic monomer

- Provide reverse phase property for analyte retention



Strong cation-exchange mode  
best retention at least 2 pH units  
below pKa

Reversed-phase  
mode

Propranolol  
Basic  
drug

Waters

# Oasis<sup>®</sup> HLB micro elution 96-well plate





Alliance<sup>®</sup> HT 2795



Micromass<sup>®</sup> Ultima  
Triple quadrupole

MS: Micromass<sup>®</sup> Quattro Ultima LC: Waters Alliance<sup>®</sup> HT 2795

Ion source: ESI (+)

Flow rate: 0.2 mL/min

Source temperature: 150 °C

Mobile phase A: Water + 0.5 % NH<sub>4</sub>OH

Gas cell: 2.0 e-3 bar Argon

Mobile phase B: ACN + 0.5 % NH<sub>4</sub>OH

Desolvation temperature: 350 °C Column: XTerra<sup>®</sup> MS C18

2.1 x 30 mm, 3.5 μm

Drying gas flow: 500 L/hr

LC conditions: 5% to 95% in 1 min.

Cone gas flow: 50 L/hr

Column temperature: ambient

Cone voltage: 25 volts

Capillary voltage: 3.5 Kv

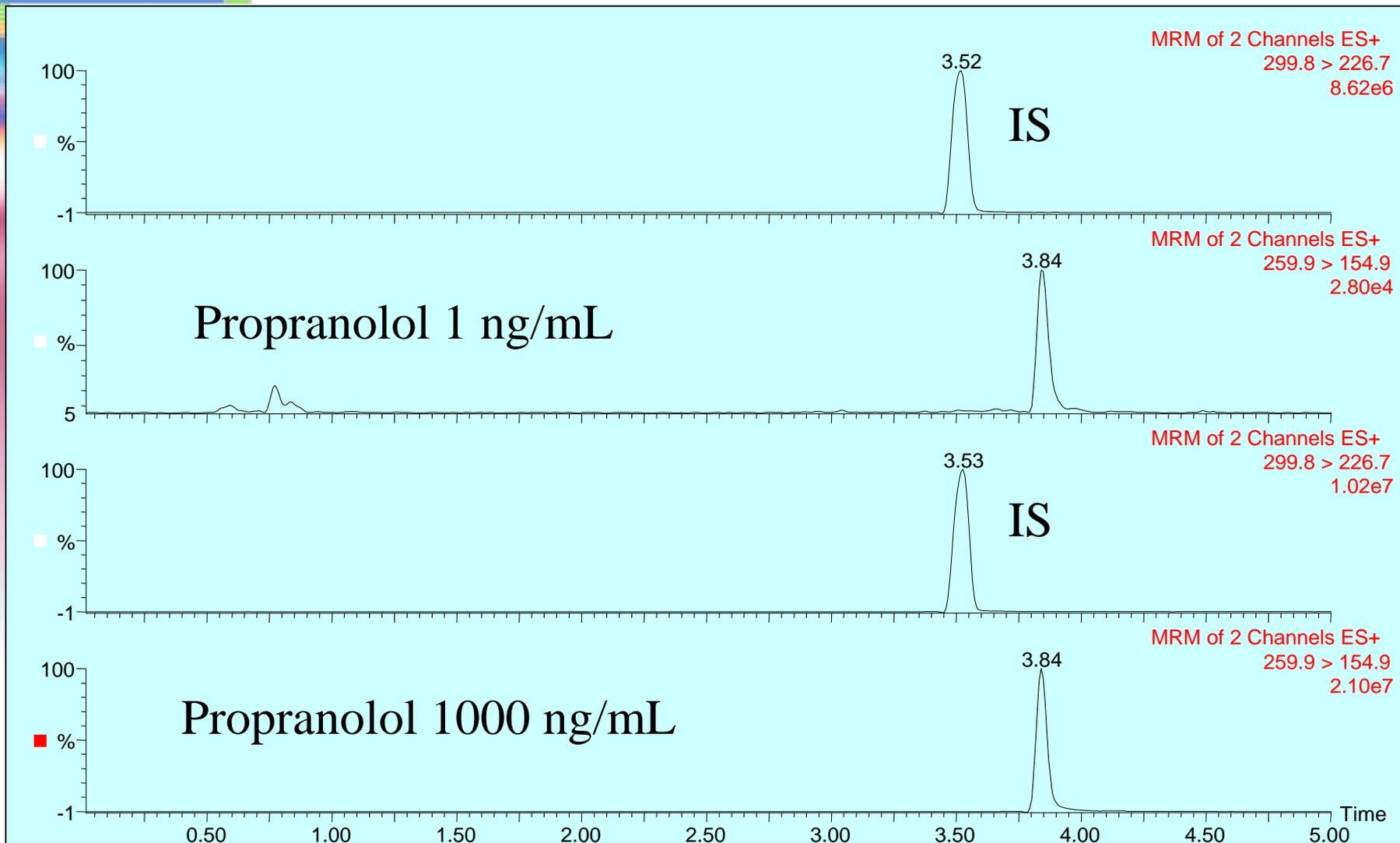
MRM transition: Metoclopramide (IS) m/z 299.8 → 226.7

Propranolol m/z 259.9 → 154.9

Amitriptyline m/z 278.1 → 232.9

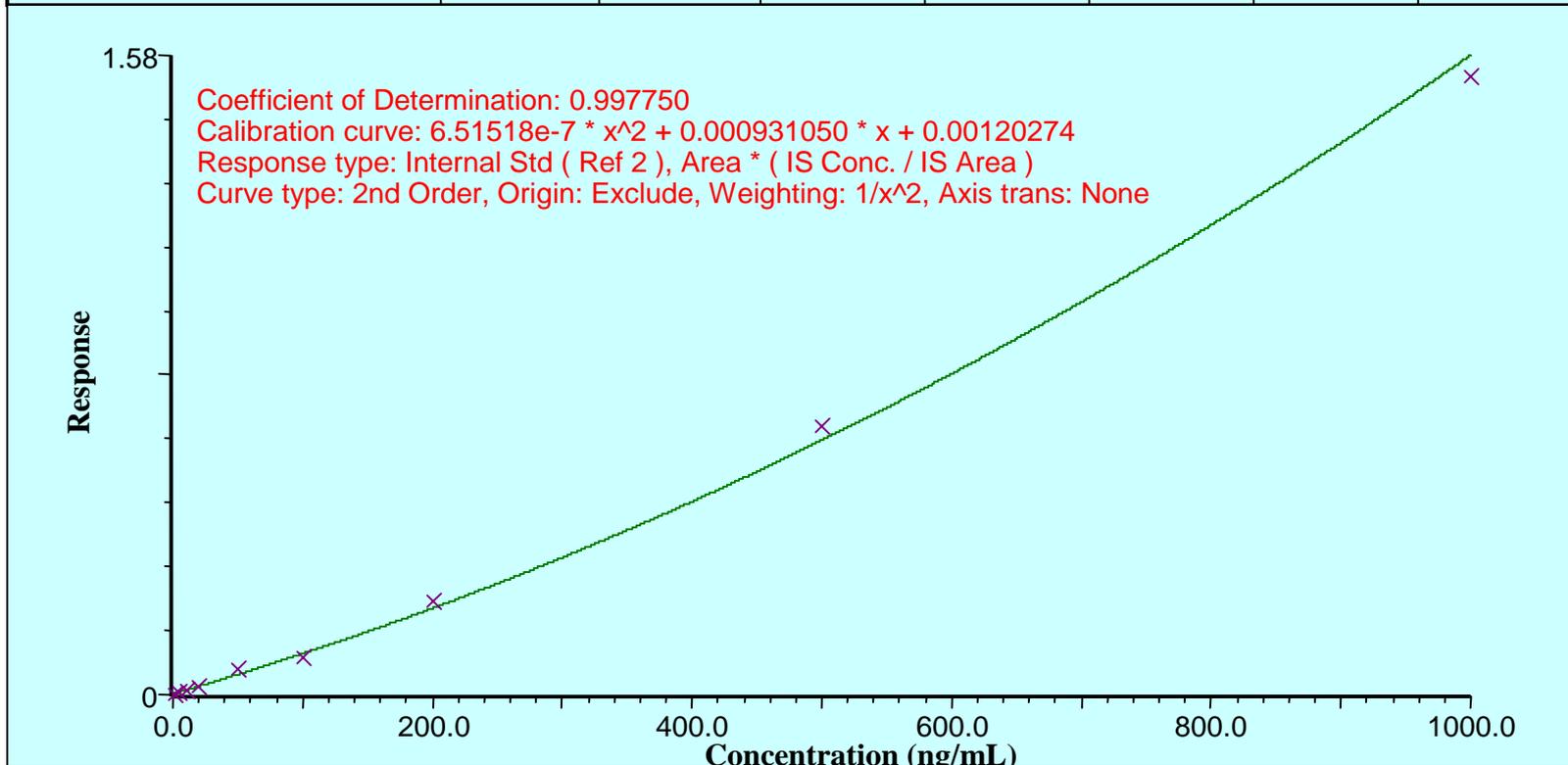
Nortriptyline m/z 263.9 → 190.8

<b>Condition</b>	200 $\mu$ L MeOH
<b>Equilibrate</b>	200 $\mu$ L Water
<b>Load</b>	50 $\mu$ L spiked rat plasma
<b>Load</b>	50 $\mu$ L IS (10 pg/ $\mu$ L) in water
<b>Wash</b>	200 $\mu$ L 5% MeOH in water
<b>Elute</b>	25 $\mu$ L ACN:IPA 40:60 + 2% FA
<b>Dilute</b>	50 $\mu$ L Water
<b>Inject</b>	20 $\mu$ L

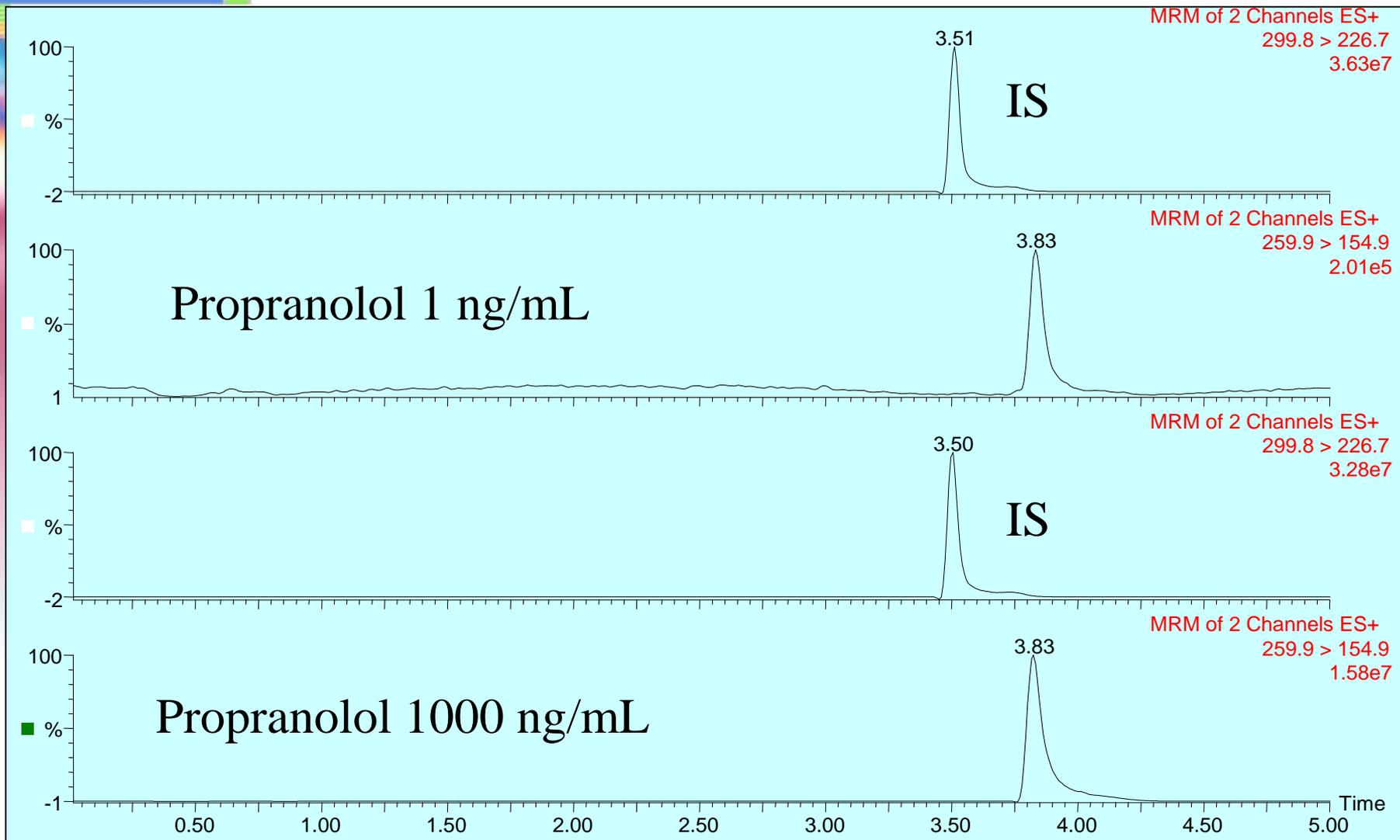


# Calibration curve of Propranolol on Oasis<sup>®</sup> HLB

<b>Conc. ng/mL N=6</b>	2.0	5.0	10.0	20.0	50.0	500.0	1000.0
<b>Average</b>	1.95	5.25	9.98	19.01	50.98	508.06	994.60
<b>Standard Deviation</b>	0.056	0.24	0.51	0.89	2.21	23.97	14.27
<b>RSD %</b>	2.8	4.7	5.1	4.7	4.3	4.7	1.4

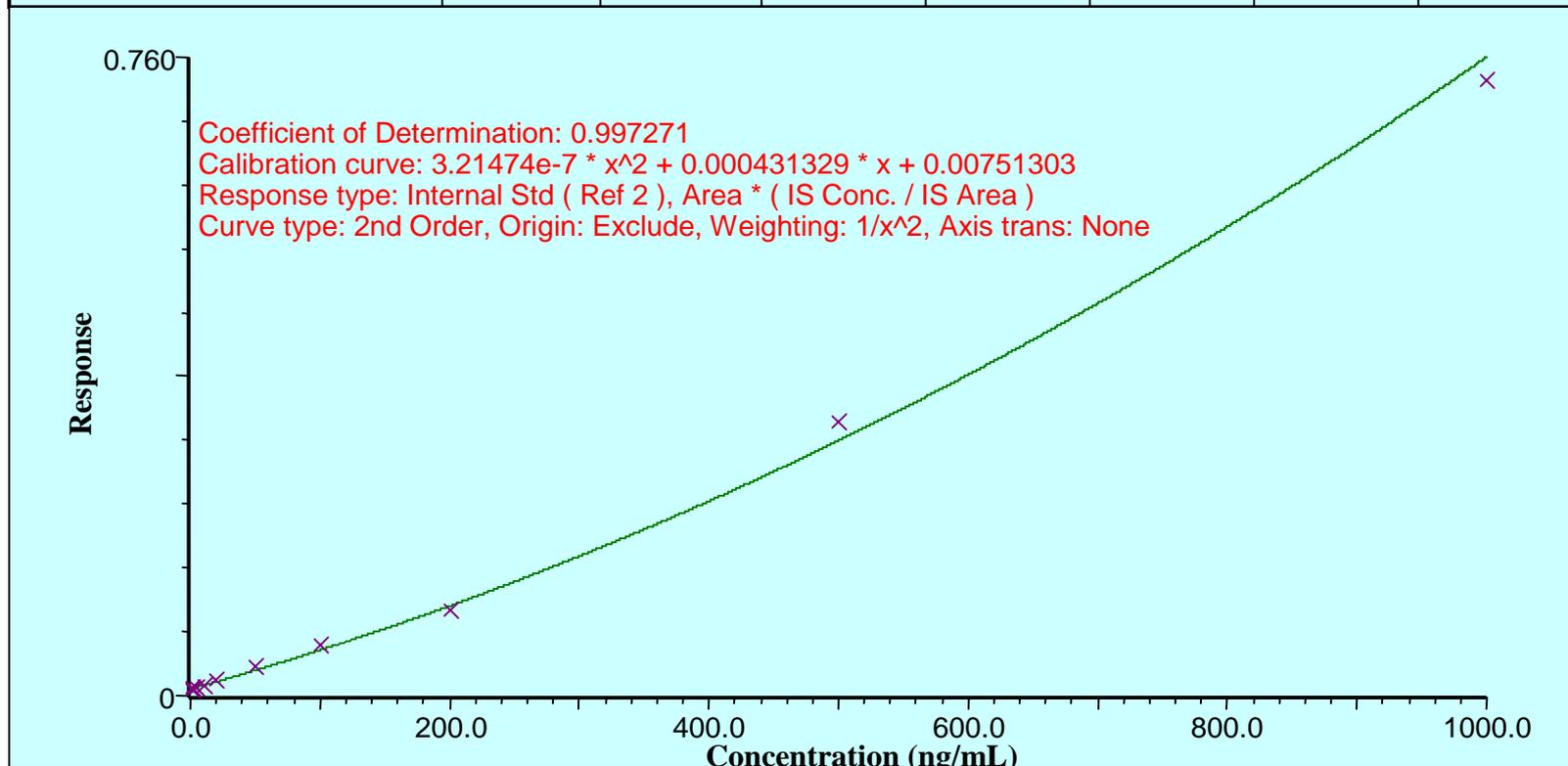


<b>Condition</b>	200 $\mu$ L MeOH
<b>Equilibrate</b>	200 $\mu$ L Water
<b>Load</b>	50 $\mu$ L spiked rat plasma
<b>Load</b>	50 $\mu$ L IS (10 pg/ $\mu$ L) in water
<b>Wash 1</b>	200 $\mu$ L Water + 2% FA
<b>Wash 2</b>	200 $\mu$ L MeOH
<b>Elute</b>	25 $\mu$ L ACN:IPA 40:60 + 2% NH <sub>4</sub> OH
<b>Dilute</b>	50 $\mu$ L water
<b>Inject</b>	20 $\mu$ L

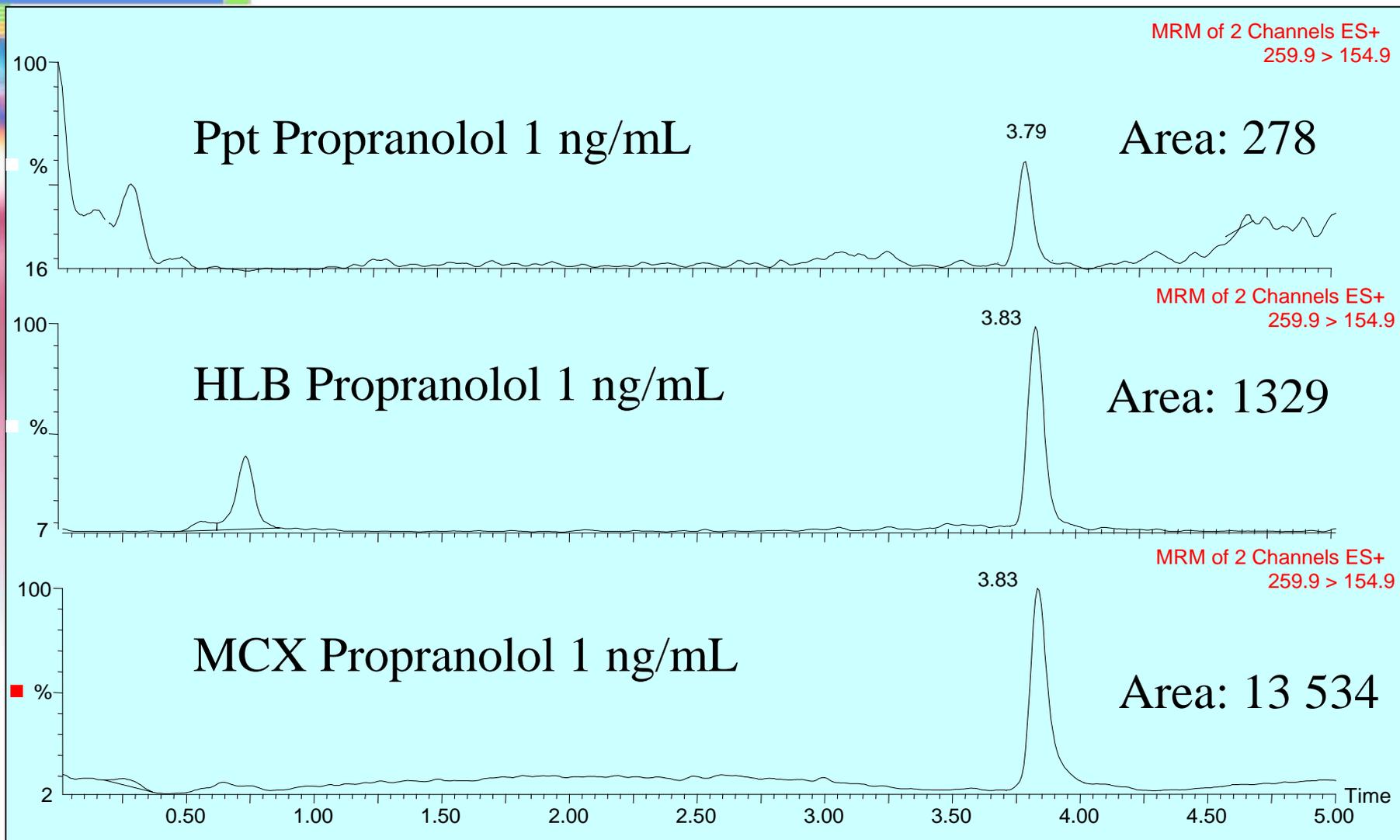


# Calibration curve of Propranolol on Oasis<sup>®</sup> MCX

<b>Conc. ng/mL N=6</b>	2.0	20	50.0	100.0	200.0	500.0	1000.0
<b>Average</b>	2.02	19.10	50.77	103.78	198.72	534.95	968.75
<b>Standard Deviation</b>	0.056	1.17	2.54	5.84	13.14	25.48	16.37
<b>RSD %</b>	2.8	6.1	5.0	5.6	6.6	4.7	1.6

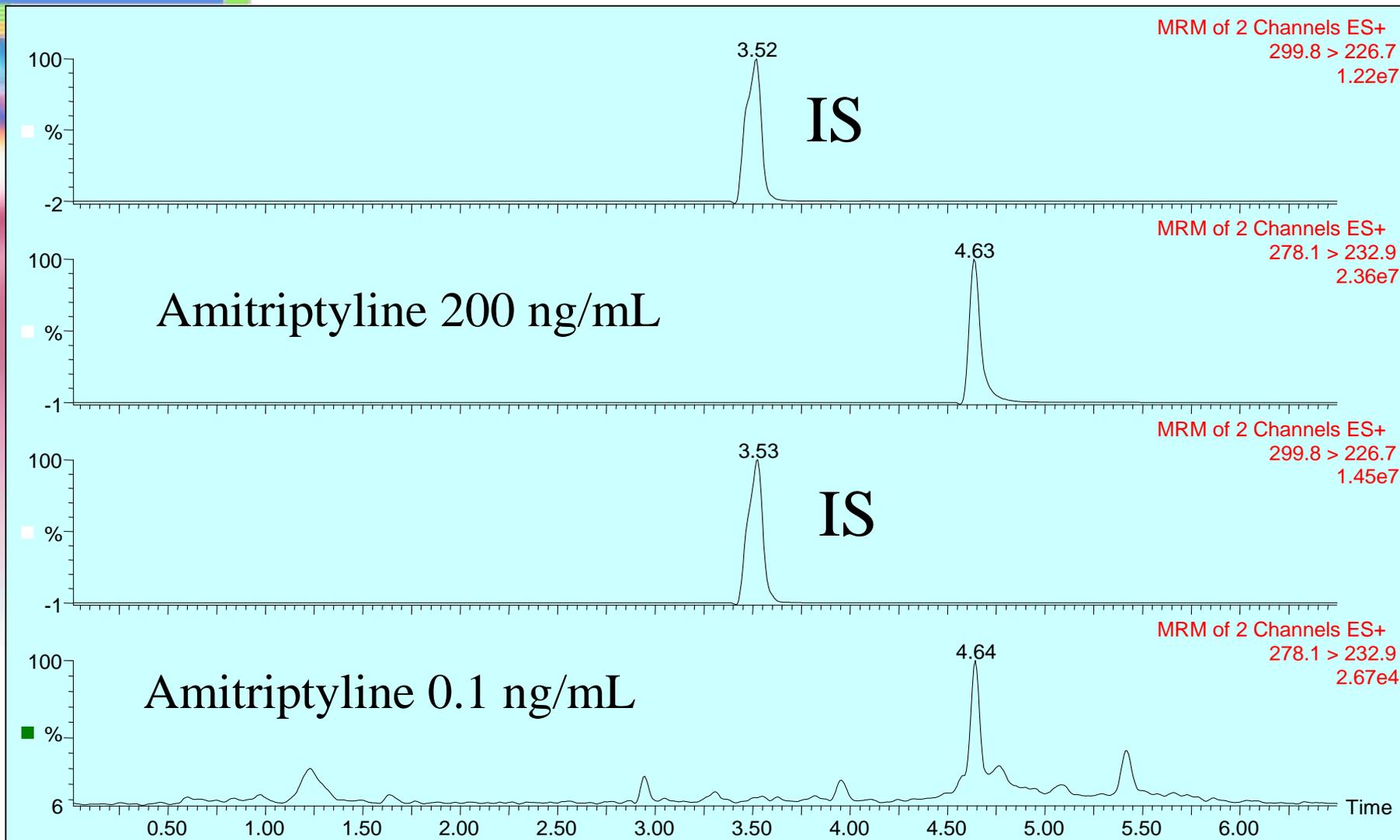


# Comparison Ppt vs HLB vs MCX



# LC/MS/MS analysis of Amitriptyline on Oasis<sup>®</sup> HLB

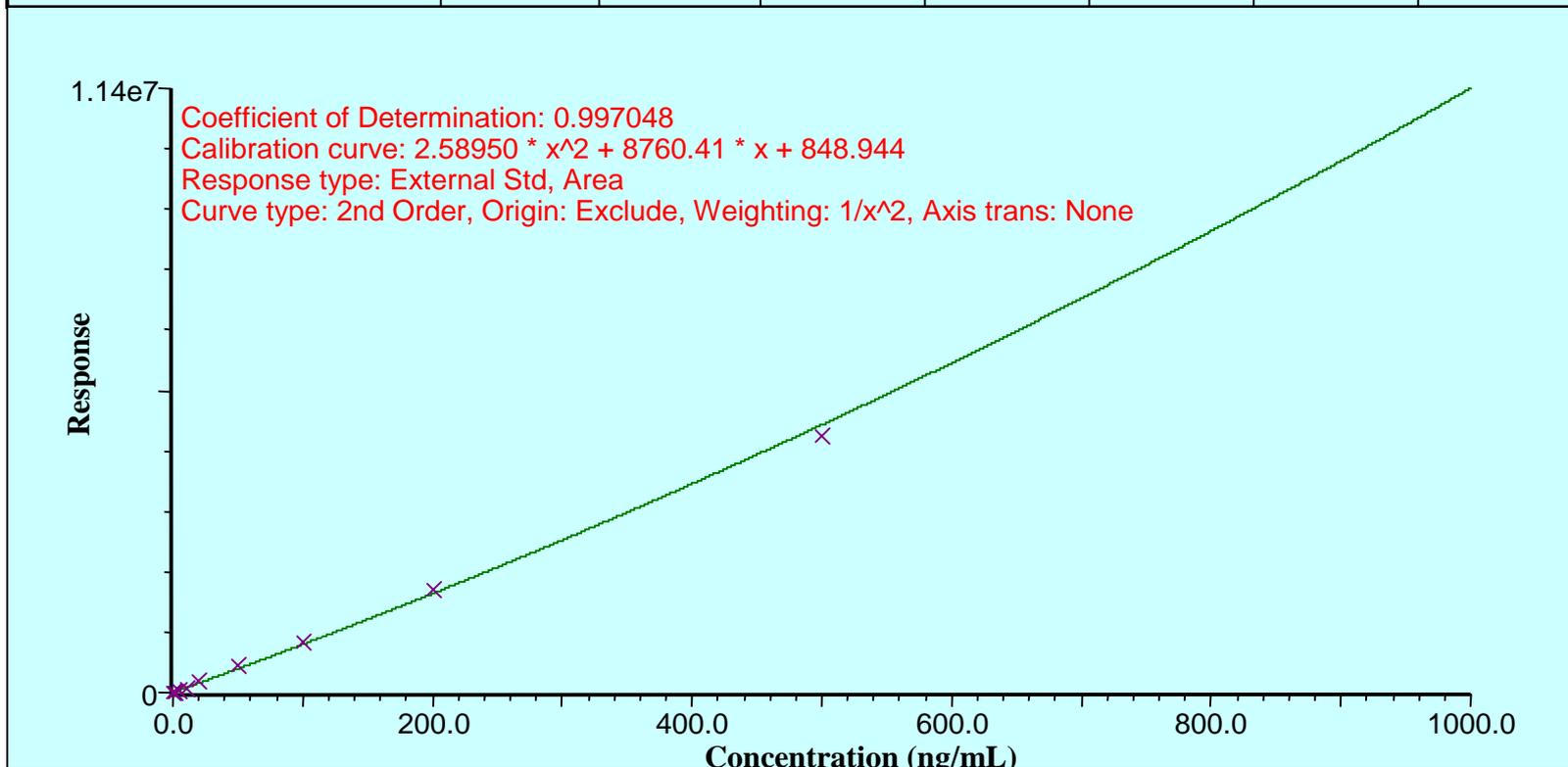
\* Elution with 40:60 ACN:IPA + 2% FA



# Calibration curve of Amitriptyline on Oasis<sup>®</sup> HLB

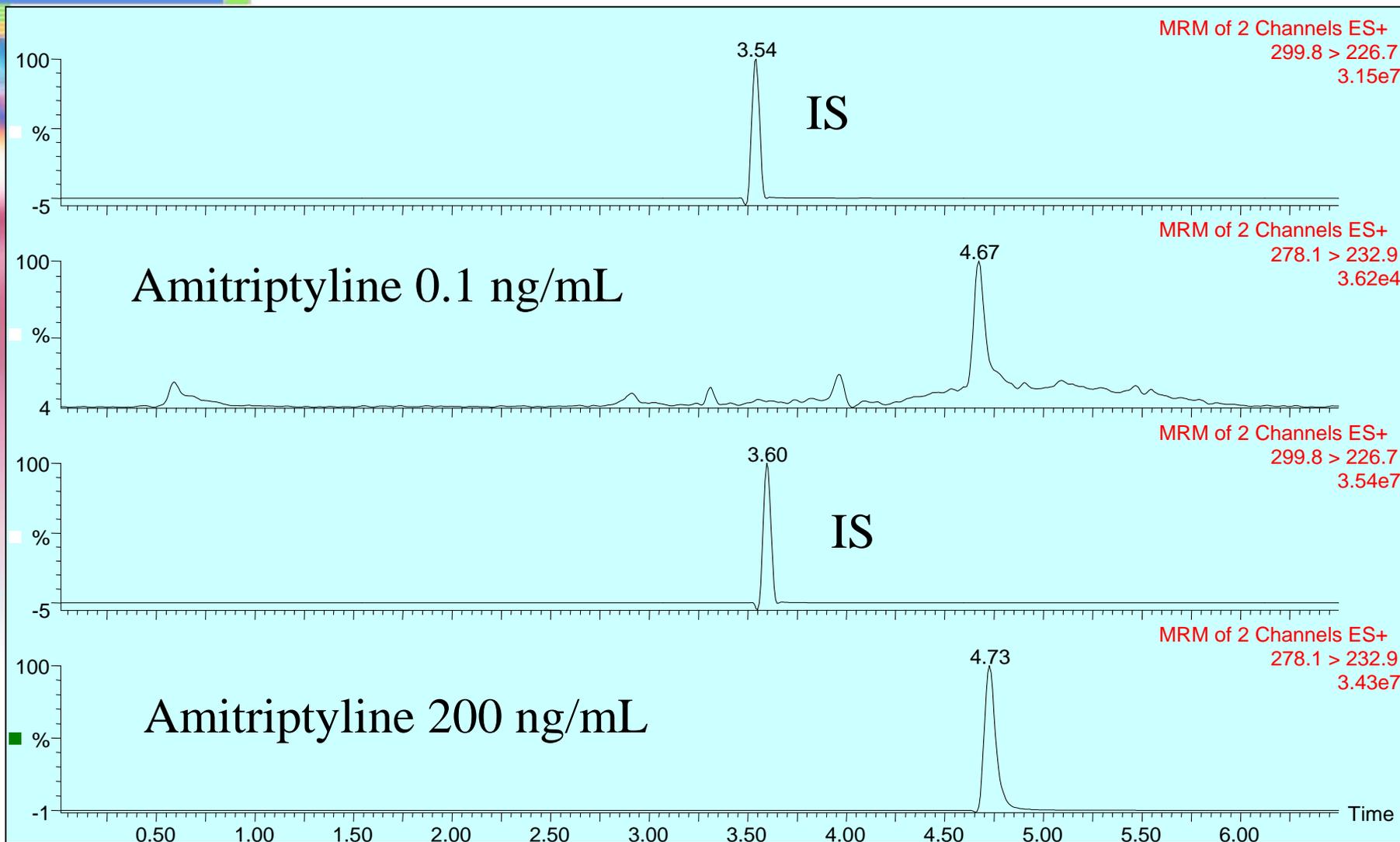
\* Elution with 40:60 ACN:IPA + 2 % FA

<b>Conc. ng/mL N=6</b>	0.1	0.2	0.5	5.0	10.0	20.0	50
<b>Average</b>	0.1	0.24	0.49	5.02	10.12	21.0	51.74
<b>Standard Deviation</b>	0.005	0.005	0.017	0.34	0.63	1.27	1.86
<b>RSD %</b>	5.0	2.1	3.6	6.9	6.2	6.0	3.6



# LC/MS/MS analysis of Amitriptyline on Oasis<sup>®</sup> HLB

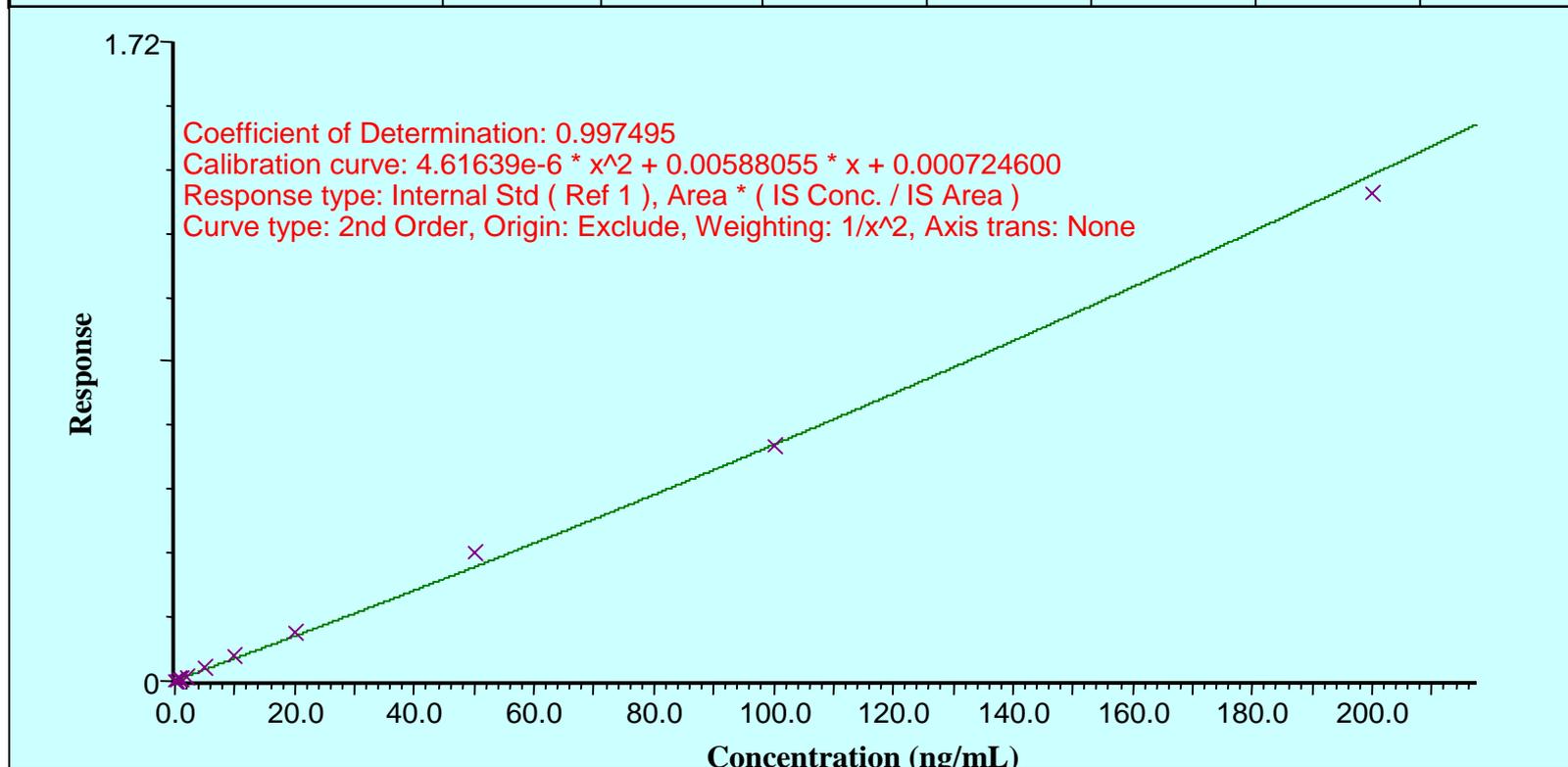
\* Elution with 40:60 ACN:IPA + 2 % NH<sub>4</sub>OH

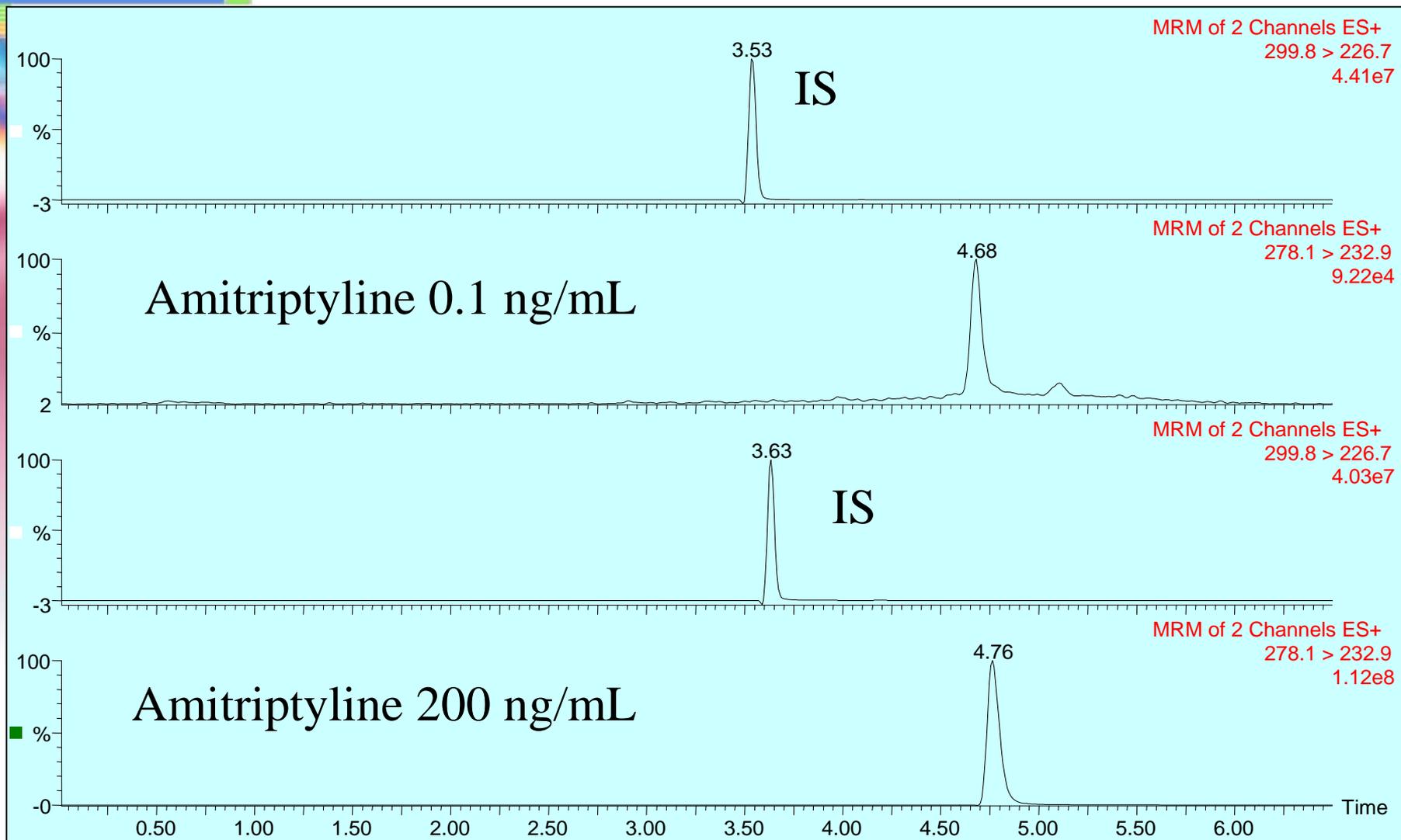


# Calibration curve of Amitriptyline on Oasis<sup>®</sup> HLB

Elution with 40:60 ACN:IPA + 2 % NH<sub>4</sub>OH

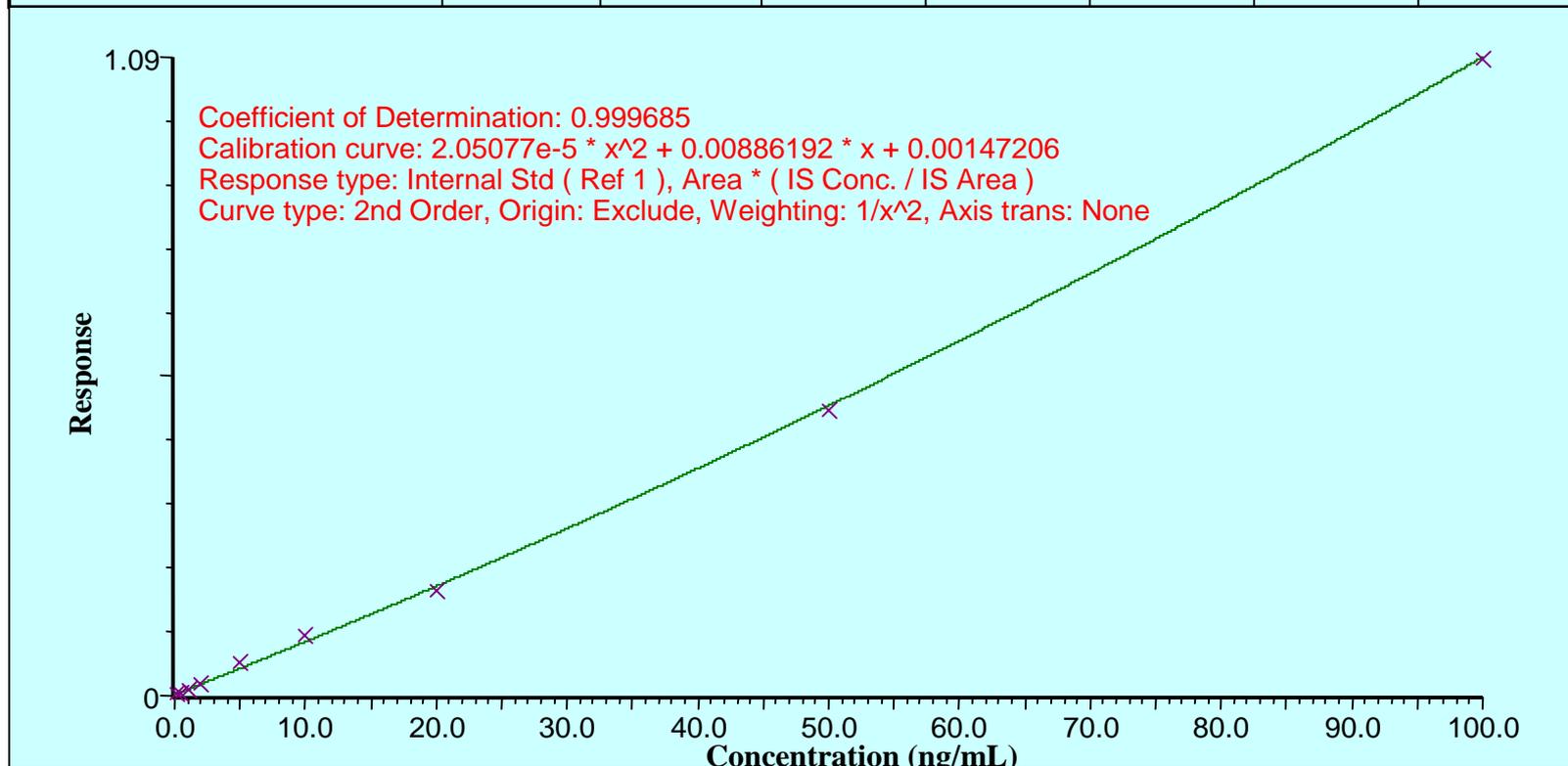
<b>Conc. ng/mL N=6</b>	0.1	0.5	1.0	5.0	10.0	50.0	100.0
<b>Average</b>	0.101	0.47	0.975	5.05	9.89	53.47	96.58
<b>Standard Deviation</b>	0.003	0.012	0.07	0.35	0.52	1.19	1.33
<b>RSD %</b>	3.6	2.5	7.2	7.0	5.2	2.2	1.3



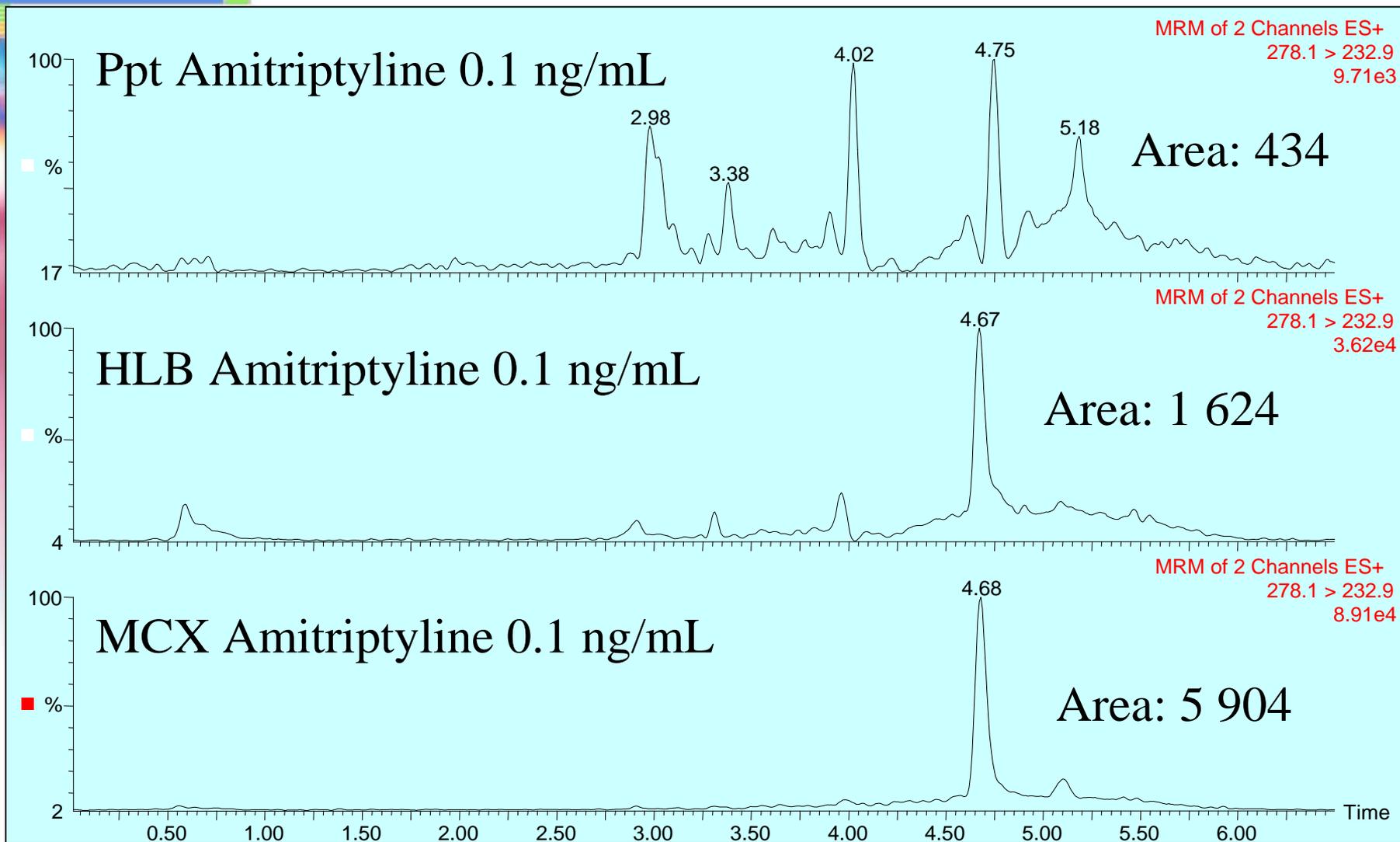


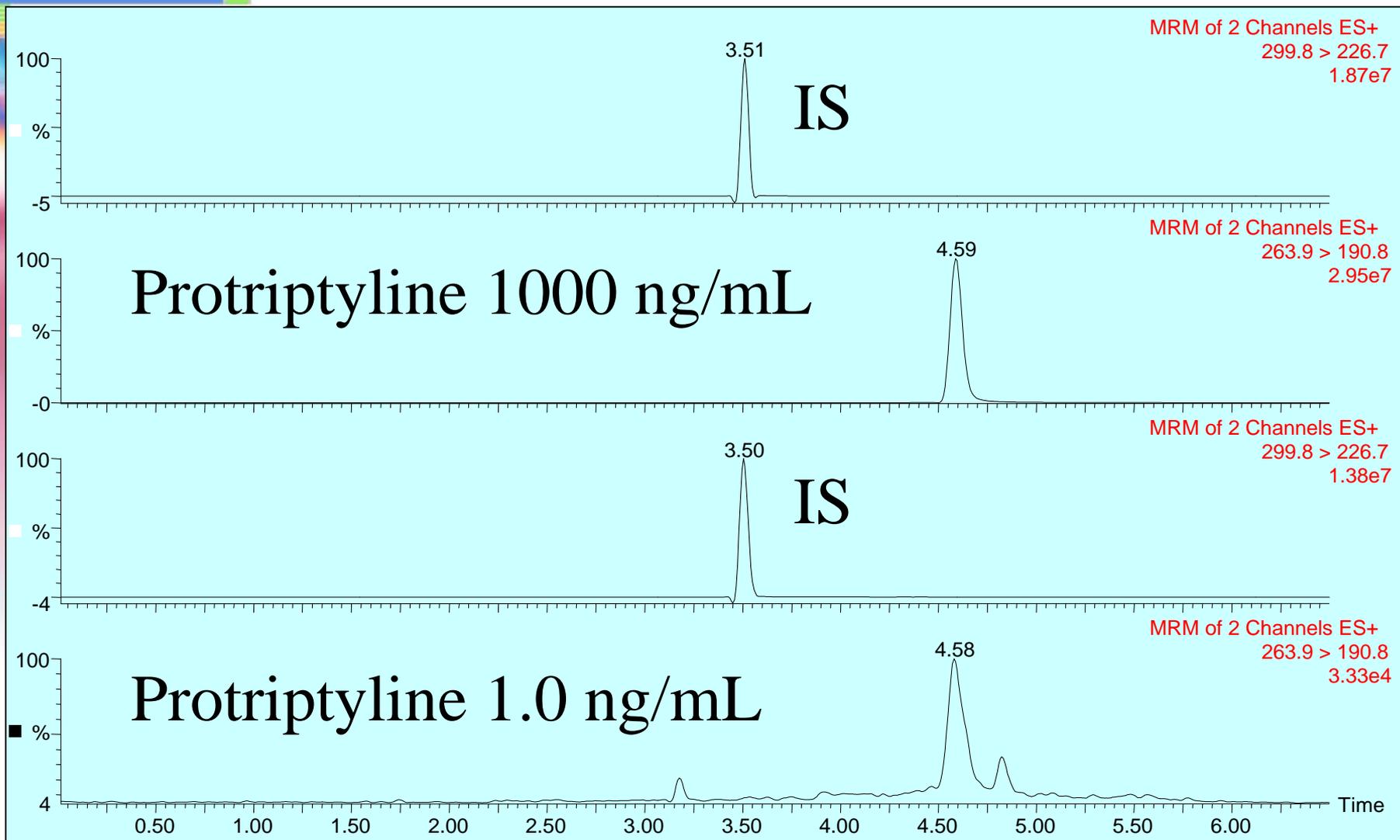
# Calibration curve of Amitriptyline on Oasis<sup>®</sup> MCX

<b>Conc. ng/mL N=6</b>	0.1	0.2	0.5	1.0	5.0	10.0	50.0
<b>Average</b>	0.105	0.188	0.481	1.023	5.104	10.59	52.0
<b>Standard Deviation</b>	0.005	0.009	0.027	0.019	0.23	0.15	2.92
<b>RSD %</b>	4.7	5.2	5.8	1.9	4.6	1.4	5.6



# Comparison of Ppt vs HLB vs MCX

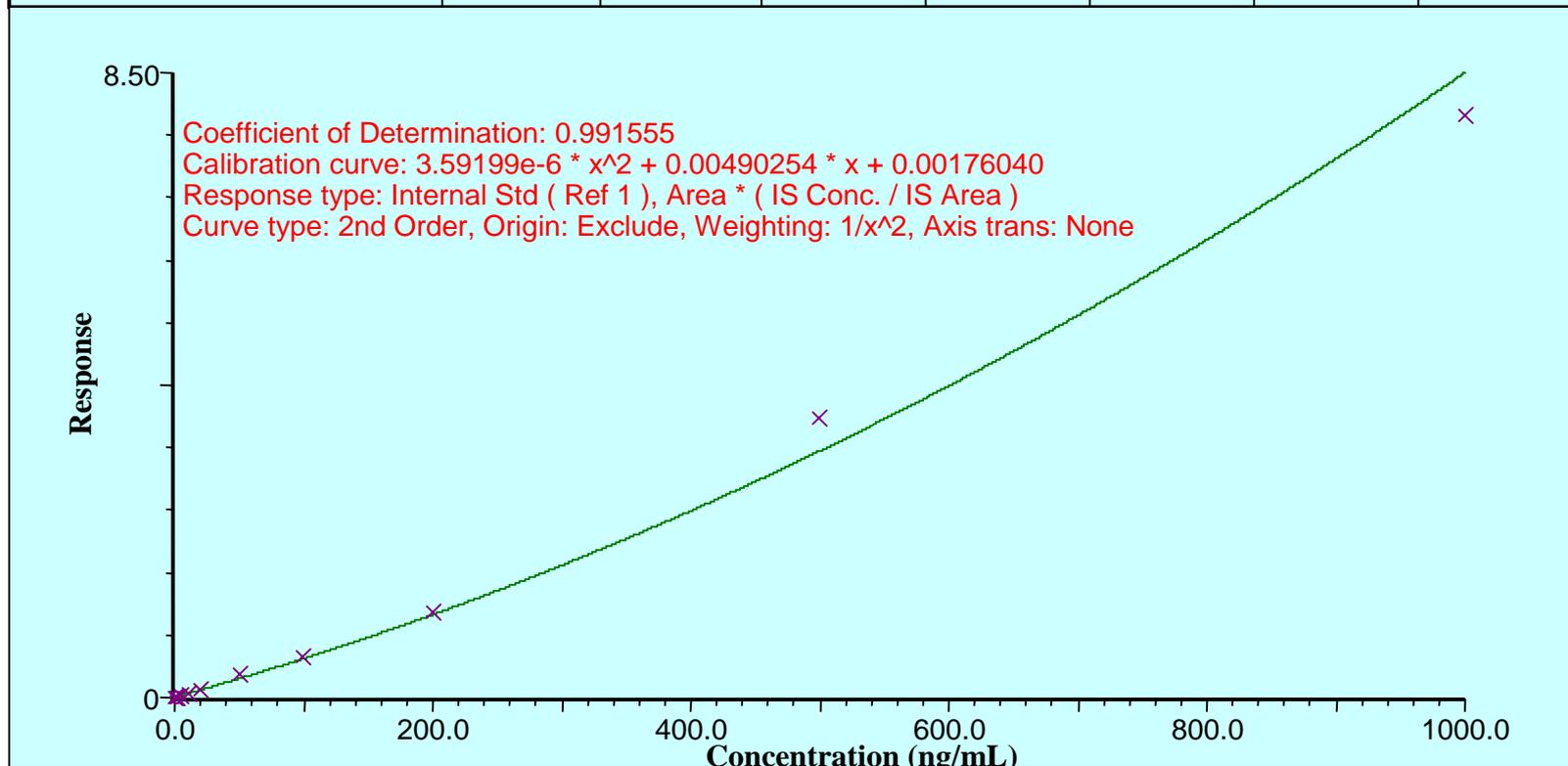


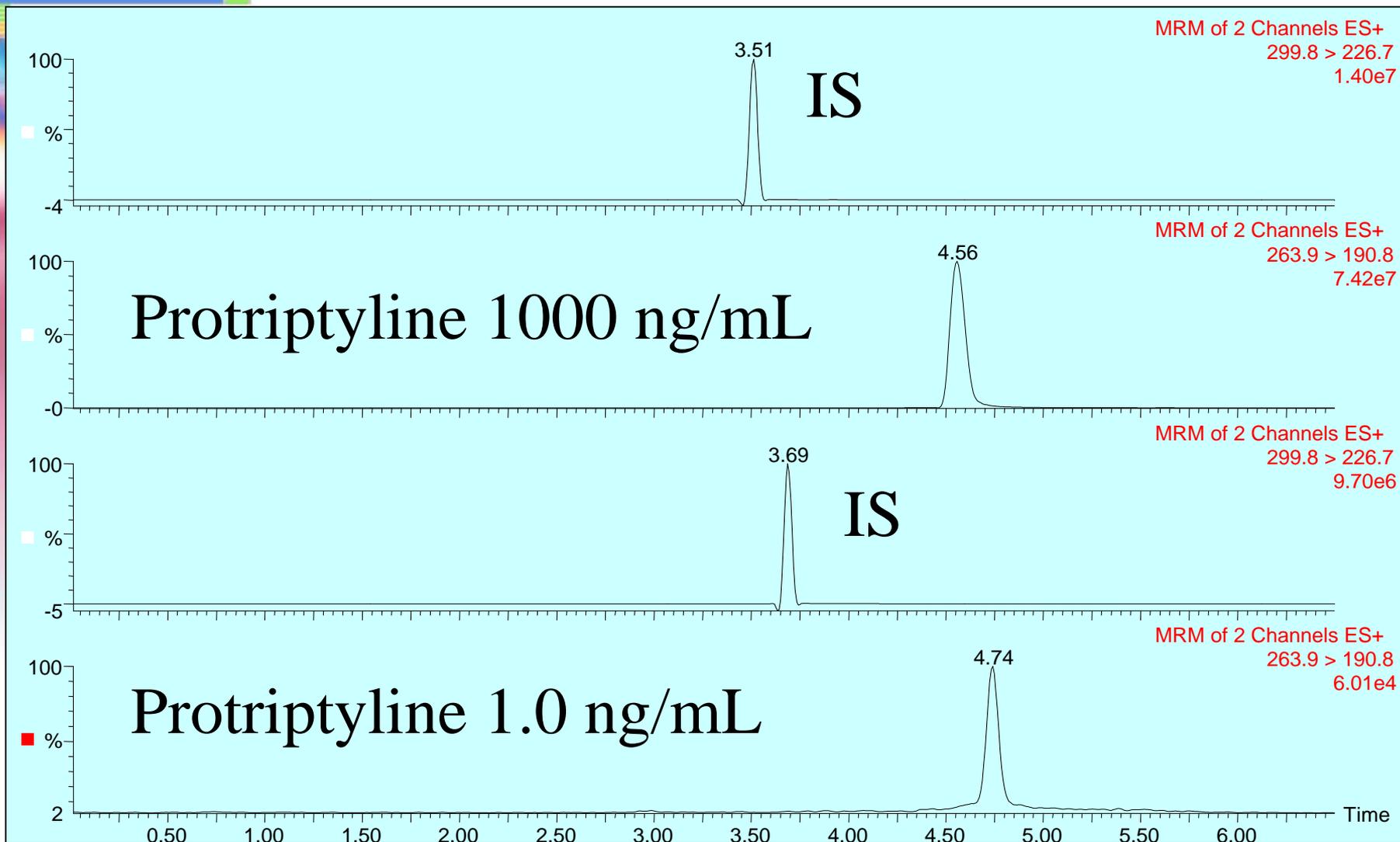


# Calibration curve of Protriptyline on Oasis<sup>®</sup> HLB

Elution with 40:60 ACN:IPA + 2 % NH<sub>4</sub>OH

<b>Conc. ng/mL N=6</b>	5.0	10.0	20.0	100.0	200.0	500.0	1000.0
<b>Average</b>	4.8	9.8	20.0	104.3	204.0	538.7	988.3
<b>Standard Deviation</b>	0.28	0.48	1.19	3.11	10.13	1.34	1.33
<b>RSD %</b>	5.8	4.9	5.9	2.9	4.9	0.2	1.3

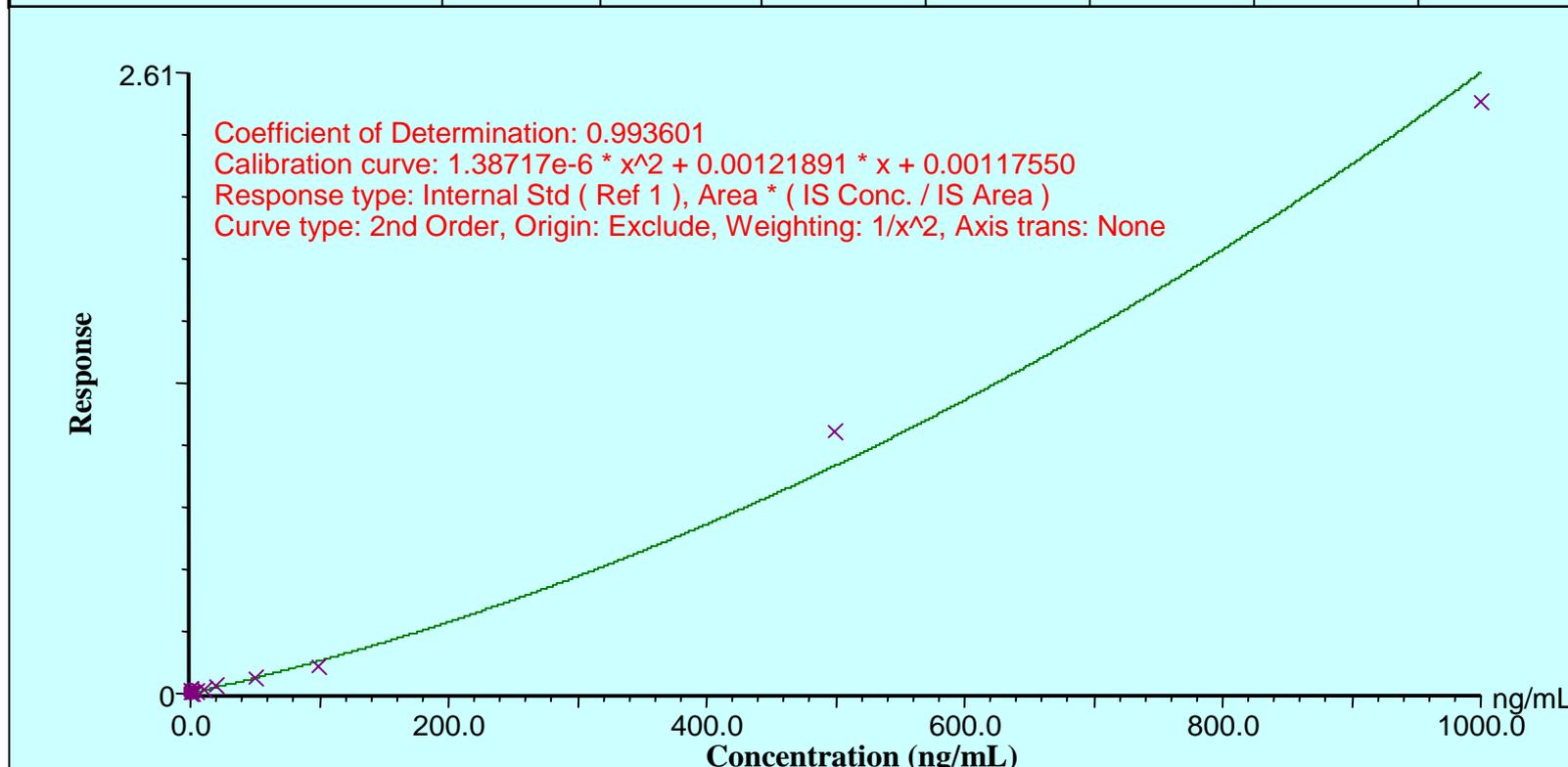




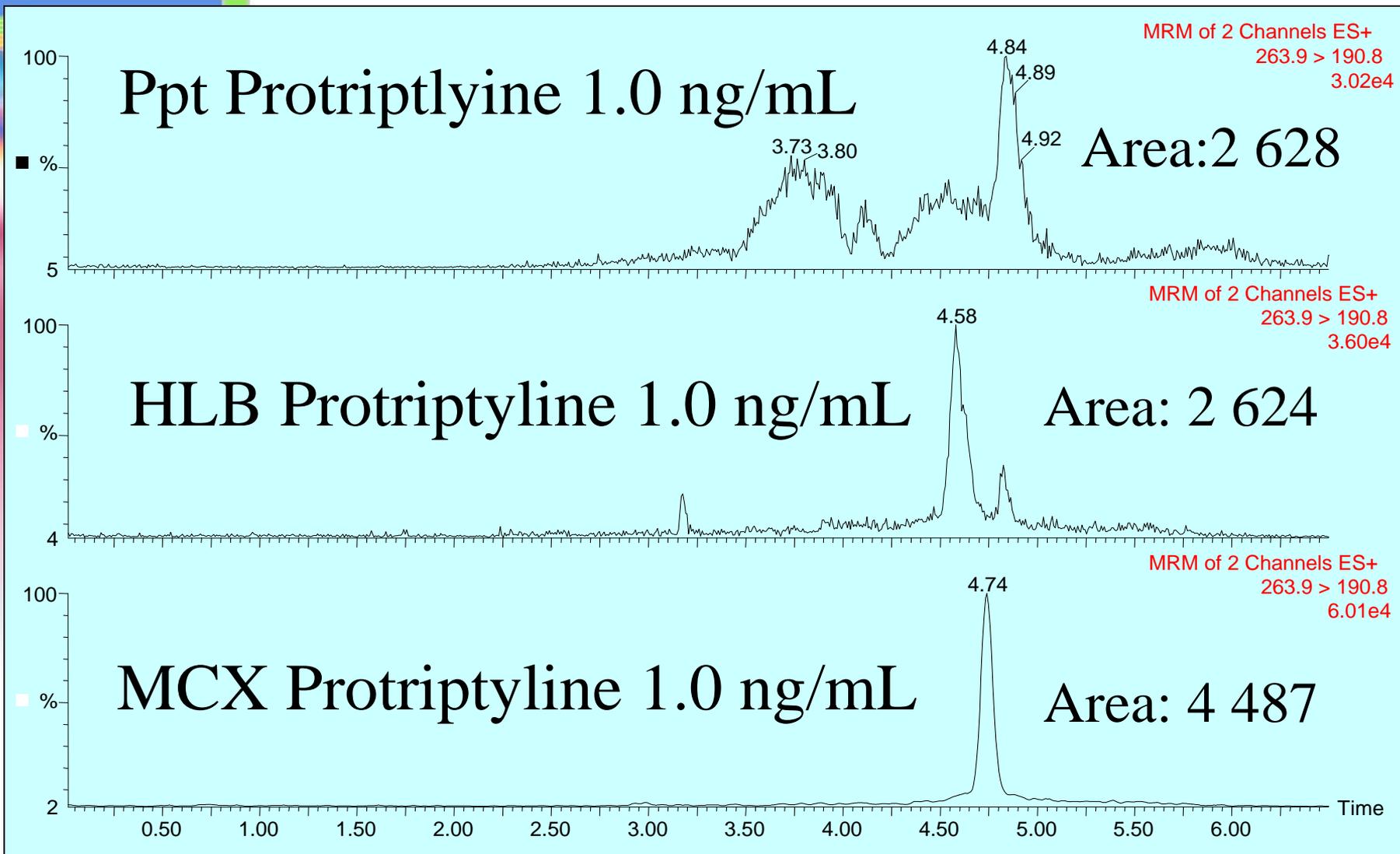
# Calibration curve of Protriptyline on Oasis<sup>®</sup> MCX

Elution with 40:60 ACN:IPA + 2 % NH<sub>4</sub>OH

<b>Conc. ng/mL N=6</b>	1.0	5.0	10.0	20.0	50.0	500.0	1000.0
<b>Average</b>	1.01	4.9	9.6	19.4	51.7	515.9	989.8
<b>Standard Deviation</b>	0.01	0.3	0.55	1.06	1.62	22.21	20.34
<b>RSD %</b>	1.5	6.1	5.7	5.4	3.1	4.3	2.0



# Comparison of Ppt vs HLB vs MCX



Wide volume range for loading (50 to 800  $\mu\text{L}$ )

Low elution volume (25 – 50  $\mu\text{L}$ )

Generic protocols for HLB and MCX

Pre-concentration factor up to 10x

Sub ng/mL quantitation limits

96-well plate format for high throughput