LAH 0200 10/84 AN/LS/MD/NA/NT

NUCLEOTIDE ANALYSIS OF CANINE CARDIAC MUSCLE

USING A RADIAL-PAK™ RESOLVE™ C₁₈ COLUMN

Many investigators (1,2) have used the depletion of nucleotides and subsequent formation of degradation products associated with ischemia (suppression of blood flow to organ or tissue) to study changes in cardiac cell metabolism, structure and function. A recent method (3) provides excellent resolution of nucleotides and their major degradation products from totally ischemic canine cardiac muscle with minimal analysis time using an isocratic elution system.

The standard nucleotides used in this study were ATP, ADP, AMP, inosine, hypoxanthine and xanthine. They were dissolved in high purity water at appropriate concentrations. The separation was accomplished on a Radial-PAKTM RESOLVETM c_{18} cartridge (10 μ , 8 mm i.d. X 100 mm) held in an RCM-100R Radial Compression Module. Figure 1 shows a separation of six standard nucleotides accomplished in ~13 minutes. Figure 2 is a separation of a control canine cardiac tissue after a perchloric acid/potassium carbonate-potassium hydroxide extraction. Figure 3 is a sample of canine cardiac tissue after 60 minutes of total ischemia. Note the increase or decrease in the concentration of the nucleotides after this procedure.

Column:

Radial-PAKTM RESOLVETM C18

Mobile Phase:

cartridge (10 μ m, 8 mm i.d. x 100 mm) 0.1 M ammonium dihydrogen phosphate

buffer, pH = 5.5 with 3M ammonium

hydroxide.

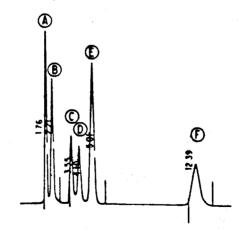
Flow Rate:

4.0 m1/min

Temperature:

Ambient

UV Detection at 254 nm.



- A. ATP
- B. ADP
- C. Hypoxanthine
- D. Xanthine
- E. AMP
- F. Inosine

FIGURE 1. Chromatogram showing the resolution of standard nucleotides.

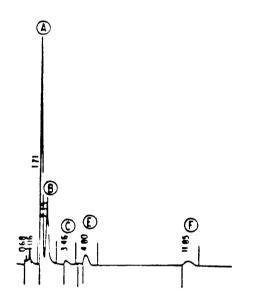


FIGURE 2. Control canine cardiac tissue. Peaks and chromatographic conditions as given in Figure 1.

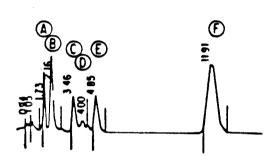


FIGURE 3. Canine cardiac tissue after 60 min. of total ischemia. Peaks and chromatographic conditions as given in Figure 1.

The ability to monitor nucleotides and their major metabolites is essential for delaying or reversing myocardial ischemic cell injury or death. The described method utilizes an inexpensive, easy to prepare, low viscosity and large buffering capacity mobile phase along with a Radial Compression Separation System. These two factors will assist investigators in easing the enormous volume of work needed in metabolic studies because they provide efficient, precise and economical results.

J. E. Lowe, R. B. Jennings and K. A. Reimer, <u>J. Mol. Cell Cardiol.</u>, <u>11</u> (1979) 1017.

J. E. Lowe, H. K. Hawkins and R. B. Jennings, <u>Surg. Forum</u>, <u>29</u> (1978) 247.
J. E. Lowe, R. G. Cummings and E. A. Hull-Ryde, <u>J. Chromatogr.</u>, <u>275</u> (1983) 411.