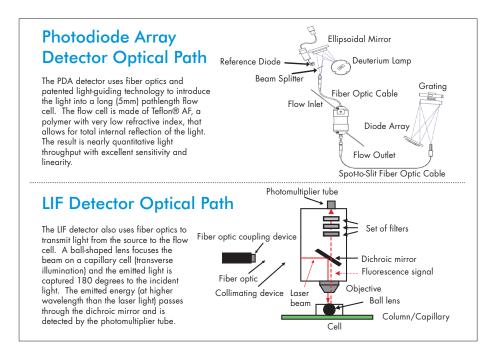
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## **Experimental Objectives**

- Study sensitivity limits in capillary chromatography
- Compare UV, laser-induced fluorescence (LIF) and mass spectrometry detection
- Explore the applicability of derivatization for enhancing amino acid and peptide detection in capillary chromatography



## Instrumental Setup

#### System 1: LC/UV/LIF Studies

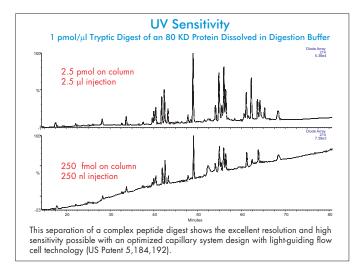
Waters CapLC System with Photodiode Array Detector, Liconix Helium Cadmium Laser and Picometrics Zetalif Fluorescence Detector •DNA adducts •Derivatized peptides •Derivatized amino acids

### System 2: LC/UV/MS Studies

Waters CapLC System plus Waters/Micromass ZMD Mass Spectrometer •DNA adducts •Derivatized peptides

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# LC/UV of Peptides



### Conditions for High Sensitivity UV Peptide Detection

#### Chromatographic Conditions

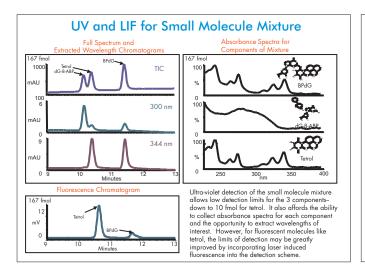
- Column: 0.32x150 mm Symmetry C<sub>18</sub>, 300 Å, 5 μm
- Eluent A: 0.02% TFA in H2O
- Eluent B: 0.017% TFA in MeCN
- Flow Rate: 5 μl/min
- Gradient: Hold at 2% B for 2 min, 2-40% B in 70 min, 40-60% B in 25 min, 60-100% B in 10 min, hold at 100% B for 10 min
- Total Run Time: 140 min

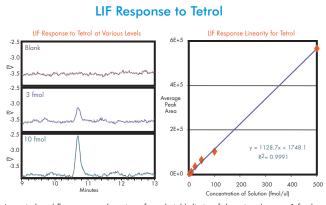
#### **Detection Parameters**

- Photodiode Array Detector with light guiding flow cell, 200 300 nm
- Sample
- Tryptic digest of an 80kD protein

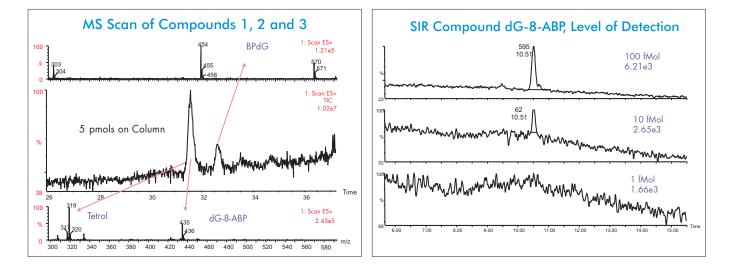
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# LC/UV/LIF of DNA-Type Adducts





Laser induced fluorescence detection of tetrol yields limits of detection down to 1 fmol, a 10-fold improvement over UV detection. The LIF response to tetrol was linear from 3 fmol/ul to 500 fmol/ul.



### Experimental Conditions for LC/UV/LIF Study of DNA-Type Adducts

#### Chromatographic Conditions

- Column: 0.32 x 15 mm Symmetry ® C18, 100 Å, 5μm
- Flow Rate: 10 μL/min
- Mobile Phase: A: 0.01% TFA in MilliQ Water, B: 100% MeOH
- Gradient: 30-70% B in 20 minutes
- Column Temperature: 50°C
- Injection Volume: 1 μl
- Total Run Time: 30 minutes

#### **Detection Parameters**

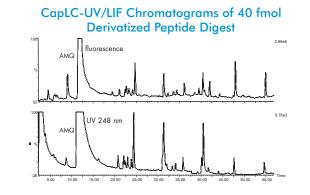
- PDA Detection from 220 to 400 nm
- Fluorescence Excitation at 325 nm, Emission Detection > 360 nm

#### Samples

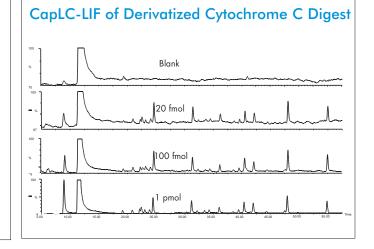
• Nucleotide adducts and precursor analog: deoxyguanosyl-amino-biphenyl (dG-8-ABP), tetrol and benzo[a]pyrene diol deoxyguanosine (BPdG), samples courtesy of Dr. Radoslav Goldman, NIH)

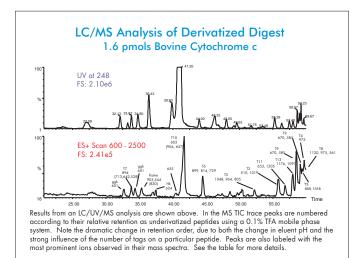
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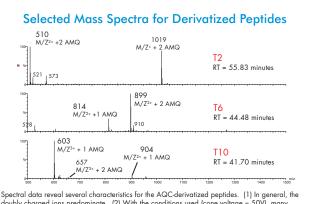
# LC/UV/LIF and LC/UV/MS of Derivatized Peptides



Detection of derivatized peptides by both UV and LIF provides excellent sensitivity with low femtomole limits. The large reagent peak (AMQ) is somewhat reduced in magnitude with the current detection system using a cutoff filter; a narrow bandpass filter(ca. 395nm) would greatly reduce the reagent peak size.







Spectral data reveal several characteristics for the AQC-derivalized peptides. (1) In general, the doubly charged ions predominate. (2) With the conditions used (cone voltage = 50Y), many peptides show fragmentation to lose one or more of the quinolyl tags. These m/z values are the same as those expected for partially tagged products, but they only appear in peaks containing the fully tagged peptides. Even with only appooximately 1.6 pmol of sample, the spectra show excellent signal-honise.

### **Experimental Conditions for Peptide Analysis**

#### Chromatographic Conditions

- Column: 0.32 x 150 mm Symmetry C18, 100 Å, 5 μm
- Eluent A: 50 mM ammonium acetate, pH 6.90; eluent B: 60% acetonitrile in water
- Gradient: 15%B to 60%B in 50 mins.
- Column temperature: 50 C.
- Samples injected: Sample 2 (20, 40, 100 fmol); Sample 1 (1 pmol

#### Detection

- PDA Detection from 220 to 400 nm
- LIF: Fluorescence Excitation at 325 nm, Emission Detection > 360 nm

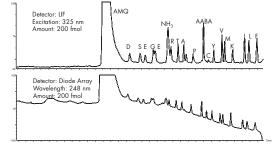
#### Samples

- Tryptic digestion of 1.25 mg/ml bovine cytochrome C with 12.5 ug/ml trypsin in 25 mM borate and 5 mM  ${\rm CaCl}_2$
- Dilute the digest 50 times with 25 mM borate
- Derivatized 160 ul of the dilute digest with 40 ul of 10 mM AQC (Sample 1)
- Dilute 20 times Sample 1 with 20% acetonitrile (Sample 2)

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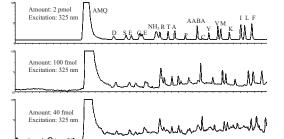
# LC/UV/LIF of Derivatized **Amino Acids**

Comparison of Capillary LC Separation of 18 Derivatized Amino Acids with LIF Detection and **Photodiode Array Detection** 



Both LIF and UV detection provide femtomole level detection for the AQC derivatized amino acids. Because a cutoff filter rather than a narrow bandpass filter was employed the reagent hydrolysis peak (AMQ) is larger than normally observed.





The AQC derivatized amino acids are highly fluorescent. Under the conditions used here, detection limits are in the low femtomole range. However, the excitation maximum for the derivatives is approximately 248 nm, not the 325 nm provided by the HeCd laser. We estimate that detection limits would improve by an order of magnitude with newer deep UV lasers that may be available within the next year.

#### **Experimental Conditions for Capillary LC** Separation of Derivatized Amino Acids

**Chromatographic Conditions:** Column: 0.32 x 150 mm Symmetry C18, 100 Å, 5 μm

- Flow rate: 5 μl/min
- Eluents: Eluent A: 140 mM sodium acetate, 17 mM triethylamine (TEA), pH 5.05
- containing 1 mM Calcium disodium EDTA; Eluent B: 60% ACN/40% water Gradient: 0 min=0%B: 0.5 min = 2% B: 15 min = 7%: 19 min = 10% B: 32 min=
- 33%B; 38 min = 100%B; 43 min= 0% B
- Column temperature: 37° C

#### **Detection parameters**

- LIF: Excitation power ~5 mW, Excitation: 325 nm, Emission cutoff filter > 365 nm
- PDA: 248 nm channel
- **Sample Preparation:**
- Place 10 μl of dilute amino acids standard in a vial
- Add 70 μ l of borate buffer; mix
- Add 20 µ l of 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate (AQC) reagent (3mg/ml in MeCNI
- Heat the vial in a reaction block or oven for 10 min at 55° C
- Inject 1 µl = 10 pmol

### **Summary and Conclusions**

- Capillary chromatography can routinely provide low to mid-femtomole level detection with common available detection methods
- With suitable derivatization techniques, LIF can enable low femtomole analysis or peptides and amino acids
- MS confirms single products for the reaction of AQC with peptides
- The next generation of lasers may permit subfemtomole analysis of AQC derivatized compounds

### **Acknowledgements**

- Amino acid separations Dr. Wensheng Xu, Waters
- Peptide separations Hongji Liu, Waters
- DNA Adducts Dr. Radoslav Goldman, NIH