

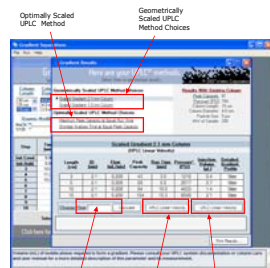
DIRECT SCALING OF HPLC SEPARATIONS TO UPLC® TECHNOLOGY FOR THE ANALYSIS OF POLAR COMPOUNDS

Jane Xu, Kenneth J. Fountain, Pamela C. Iraneta, Diane M. Diehl
Waters Corporation, Milford, MA 01757, USA

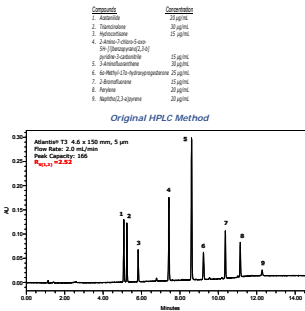
INTRODUCTION

Performing fast and efficient separations of polar compounds without compromising resolution is challenging. Waters ACQUITY UPLC® HSS T3 columns successfully meet this challenge because they are designed to retain and separate polar compounds in a UPLC environment. Just like Atlantis® T3 HPLC columns, HSS (high strength silica) T3 UPLC columns use a trifunctional C₁₈ alkyl phase bonded at a ligand density that enhances polar compound retention. As a result, these columns are capable of operating at 100% aqueous conditions with minimal dewetting.

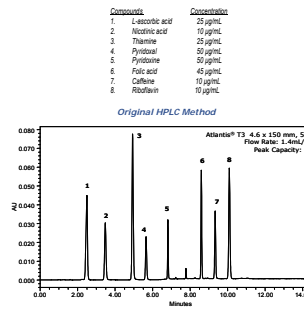
Since ACQUITY UPLC® HSS T3 columns benefit from the same lifetime, peak shape, and stability seen with Atlantis® T3 columns, methods are easily transferred between HPLC and UPLC technology. Possessing the ability to separate polar compounds faster with ACQUITY UPLC® HSS T3 columns allows for more efficient method development and chromatographic condition screening.



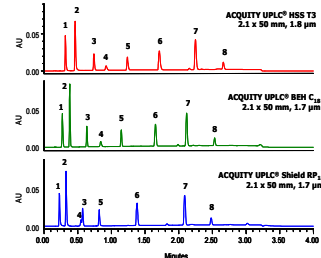
Polar and Non-polar Compounds



Water Soluble Vitamins

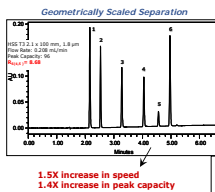
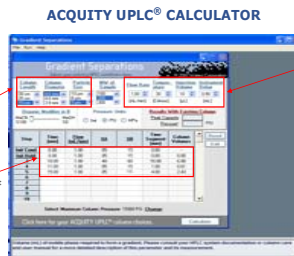
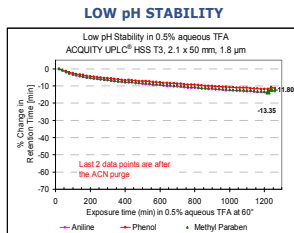


COMPARISON OF UPLC CHEMISTRIES FOR CARDIAC DRUG COMPOUND RETENTION

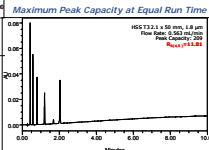
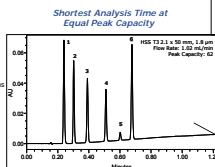
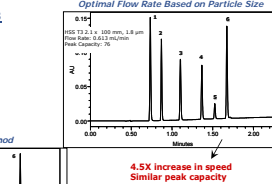


Mobile phase A was 10 mM ammonium formate in H₂O, pH 3.0. Mobile phase B was methanol. The gradient started at 15% B and increased to 75% B in 2.5 minutes, then held at 75% B for 0.5 minutes. Flow rate was 0.8 mL/min. UV @ 270 nm, 30 °C. Injection volume was 1.0 µL. Peaks: 1 = Procainamide, 2 = Procaine, 3 = Pindolol, 4 = Lidocaine, 5 = Disopyramide, 6 = Propranolol, 7 = Nifedipine, 8 = Nimodipine. The mixture was prepared in MeOH/10 mM ammonium formate in water, pH 3.0 (15/85).

APPLICATIONS

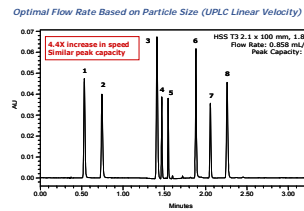
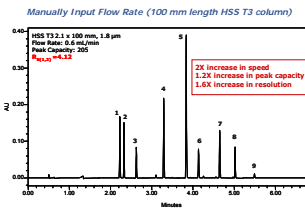
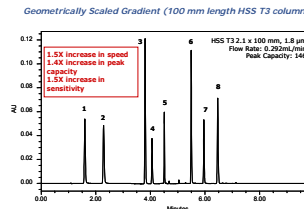
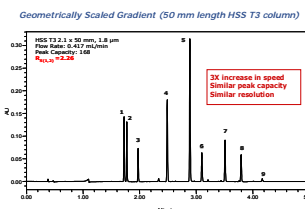


Analgesics



Comparison of ACQUITY UPLC® calculator options. Mobile phase A was 0.1% HCOOH in H₂O. Mobile phase B was 0.1% HCOOH in ACN. The gradient (HPLC) started with 15% B, then increased to 60% B in 10 minutes. Flow rate was 1.0 mL/min. UV @ 260 nm. The UPLC column was Atlantis® T3 4.6 x 150 mm, 5 µm. The mixture was prepared in water. The injection volume was 10 µL.

The gradient (UPLC Linear Velocity) started with 15% B, then increased to 60% B in 1.13 minutes. Flow rate was 0.613 mL/min. UV @ 260 nm, 30 °C. The UPLC column was ACQUITY UPLC® HSS T3 2.1 x 50 mm, 1.8 µm Q2 2.1 x 100 mm, 1.8 µm. The injection volume was 0.7 µL (50 nm) or 1.4 µL (100 nm).



Mobile phase A was 0.1% HCOOH in H₂O. Mobile phase B was 0.1% HCOOH in ACN. The gradient (HPLC) had an initial hold at 0% B for 0.5 minutes, then increased to 100% B at 10 minutes and hold 100% B for 5 minutes. Flow rate was 2.0 mL/min. UV @ 254 nm. The HPLC column was Atlantis® T3 4.6 x 150 mm, 5 µm. The injection volume was 10 µL.

The gradient (UPLC Linear Velocity) started with an initial hold at 0% B for 0.33 minutes, then increased to 100% B at 0.67 minutes and hold 100% B until 1.00 minutes. Flow rate was 0.477 mL/min. UV @ 254 nm. The UPLC column was ACQUITY UPLC® HSS T3 2.1 x 100 mm, 1.8 µm Q2 2.1 x 100 mm, 1.8 µm. The mixture was prepared in 0.1% HCOOH in water/0.1% HCOOH in ACN (50/50). The injection volume was 0.7 µL (50 nm) or 1.4 µL (100 nm).

Mobile phase A was 0.1% TFA in H₂O. Mobile phase B was ACN. The gradient (HPLC) started with 0% B, then increased to 2% B at 4 minutes, to 15% B at 8 minutes and 20% B at 15 minutes. Flow rate was 1.4 mL/min. UV @ 260 nm. The HPLC column was Atlantis® T3 4.6 x 150 mm, 5 µm. The mixture was prepared in water. The injection volume was 10 µL.

The gradient (UPLC Linear Velocity) started with 0% B, then increased to 2% B in 0.3 minutes, to 15% B at 1.3 minutes and 20% B at 3.40 minutes. Flow rate was 0.858 mL/min. UV @ 260 nm. The UPLC column was ACQUITY UPLC® HSS T3 2.1 x 100 mm, 1.8 µm. The injection volume was 1.4 µL.

