

AIM

- To enhance precursor ion sensitivity on a Q-ToF instrument

METHOD

- T-Wave ion mobility separator used to replace scanning quadrupole
- ToF duty cycle increase using synchronization with T-Wave ion guide

RESULTS

- TOF duty cycle enhancement up to x10
- Combination with ion mobility separation of precursors provides up to x100 increase in response over precursor ion scanning on a tandem quadrupole

OVERVIEW

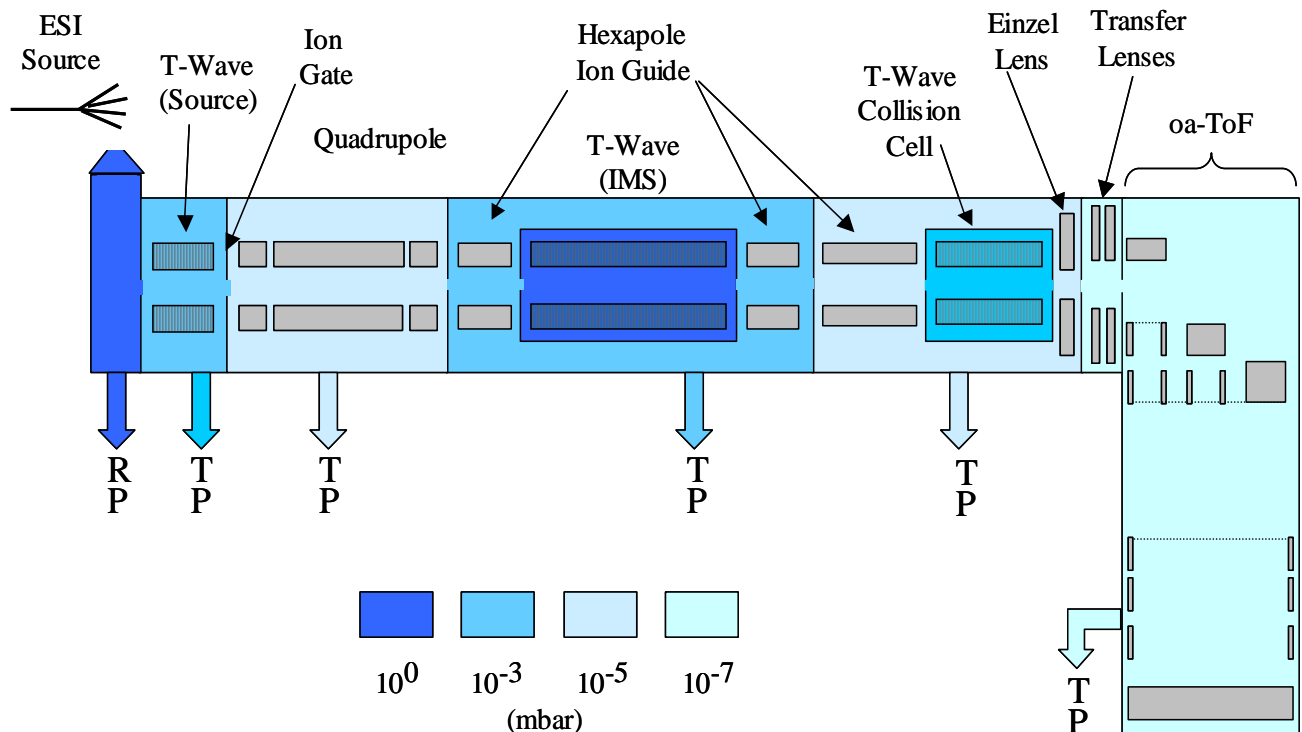


Figure 1 Schematic diagram of the modified Q-ToF Premier instrument

The mobility separated ions can be injected into the T-Wave collision cell with low energy to allow recording of the precursor ions, or with high energy to record fragment ions. The continually running T-Wave in the cell maintains the mobility profile in both low and high energy cases and thus enables correlation of fragment ions with associated precursors.

Enhancement of the ToF duty cycle (EDC) for a product ion is achieved by synchronising the ToF pusher with the ion packet release from the cell T-Wave and introducing a fixed delay to account for the time of flight of the ion of interest from the cell to the pusher region.

Experimental Parameters

Nano ESI interface - samples infused at 400 nL/min

	Quattro Premier	Modified Q-ToF Premier	
		T-Wave	
	Collision Cell	IMS Cell	Collision Cell
Pressure	4.0x10 ⁻³ mbar Ar	1.0 mbar N ₂	2.0x10 ⁻² mbar N ₂
Pulse Height	5 V	21 V	3.5 V
Pulse Velocity	300 m/s	600 m/s	300 m/s
Acquisition	Precursor ion scan 50-1100 Da in 1 s	IMS ToF MS 0-1950 Da	
Time	30 s	30 s	

RESULTS

Figure 2 shows the mass spectra obtained by conventional tandem quadrupole precursor ion scanning for decreasing concentrations of verapamil in 70/30 acetonitrile/water (0.1% formic acid). From a comparison of rows 2 and 3 of Figure 2, it is reasonable to assume that the limit of detection (LOD) for this technique, defined by a continuous peak half-width, is approximately 1-2pg/μL.

The ion mobility spectra and associated mass spectra of Figure 3 illustrate the technique of enhanced precursor ion scanning on a modified Q-ToF Premier MS. Row 1 shows the low collision energy ion mobility separation and the summed mass spectrum for 500fg/μL of verapamil.

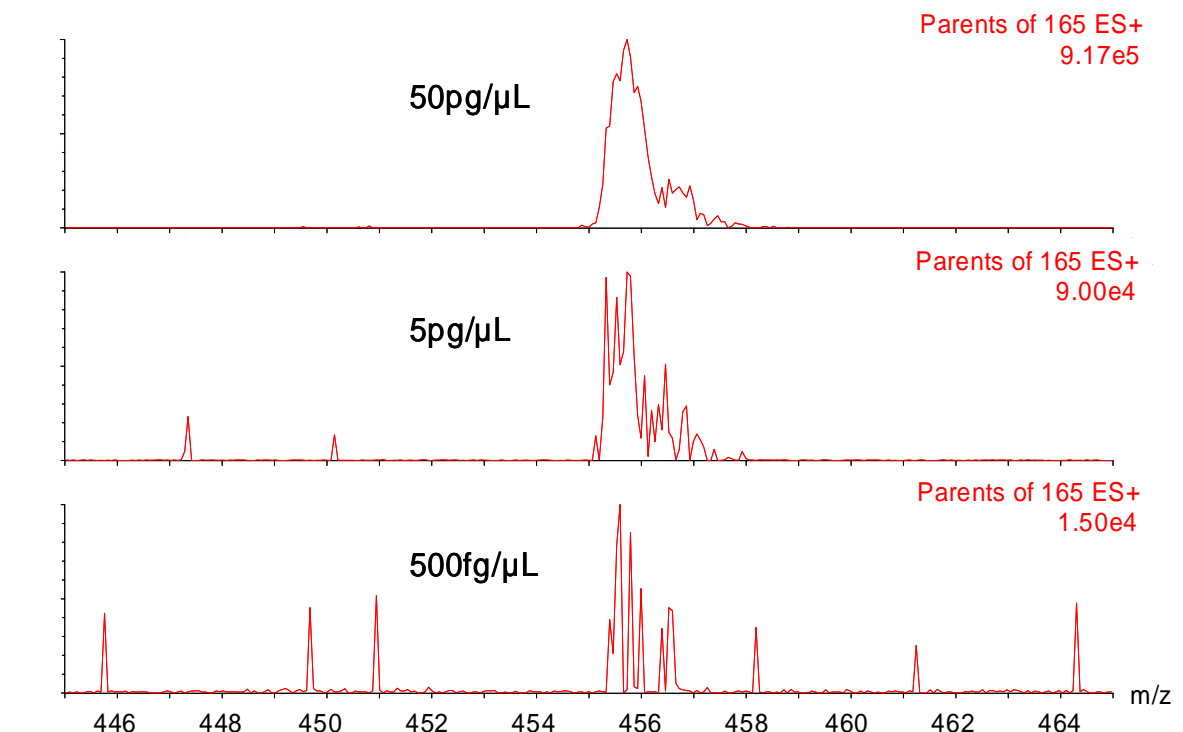


Figure 2 Conventional tandem quadrupole precursor ion scans for decreasing amounts of verapamil

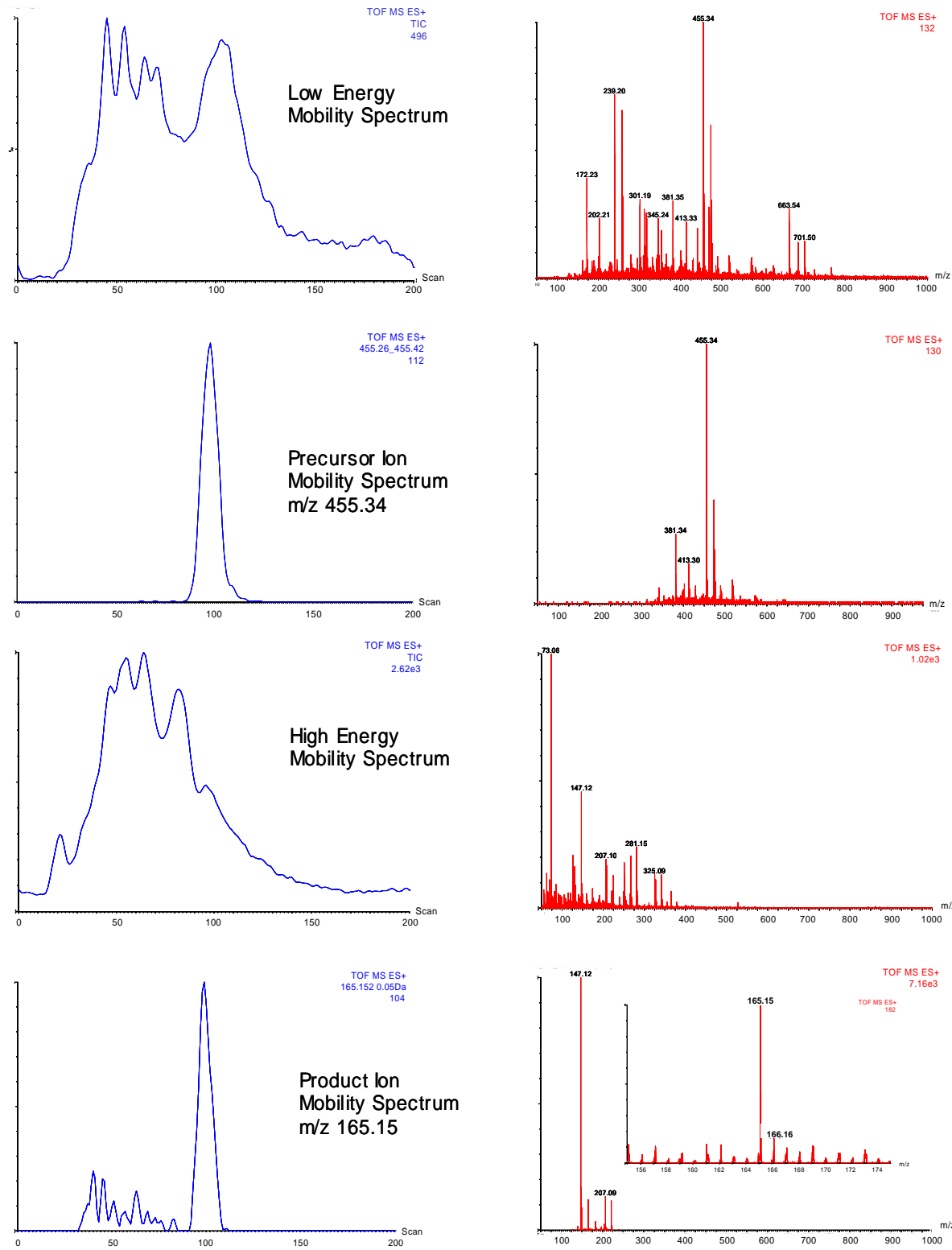


Figure 3 Ion mobility separations and their associated mass spectra obtained under low and high energy conditions for 500fg/μL verapamil

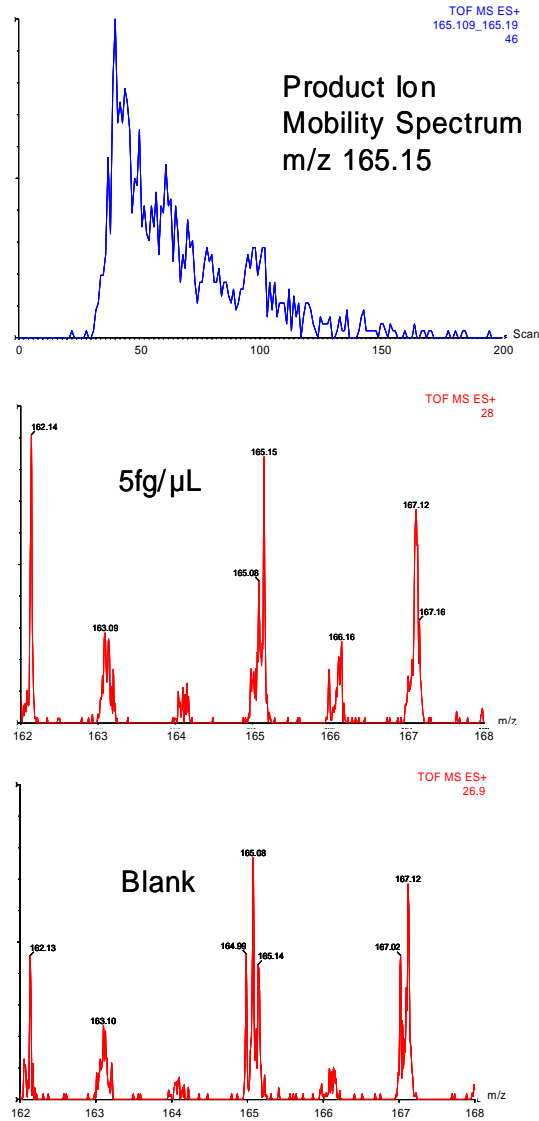


Figure 4 LOD for precursor ion scanning of 5fg/μL verapamil with the modified Q-ToF Premier

At a total digest concentration of 100fmol/μL (row 3), we can estimate that the doubly charged precursor ion (FQBEEQQQTEDELQDK) contains around 10 ions! In contrast, the 100fmol/μL data for the modified Q-ToF Premier, shown in Figure 6, demonstrates a high degree of precursor ion confirmation (rows 2 and 4). Furthermore, the first two isotopes for the dephosphorylated product ion spectrum (Figure 6, row 4) contain approximately 1000 ion counts (EDC factor ≈4x). These data reinforce the assertion that the modified Q-ToF premier precursor ion scanning technique can show x100 improvements in sensitivity when compared to a tandem quadrupole instrument.

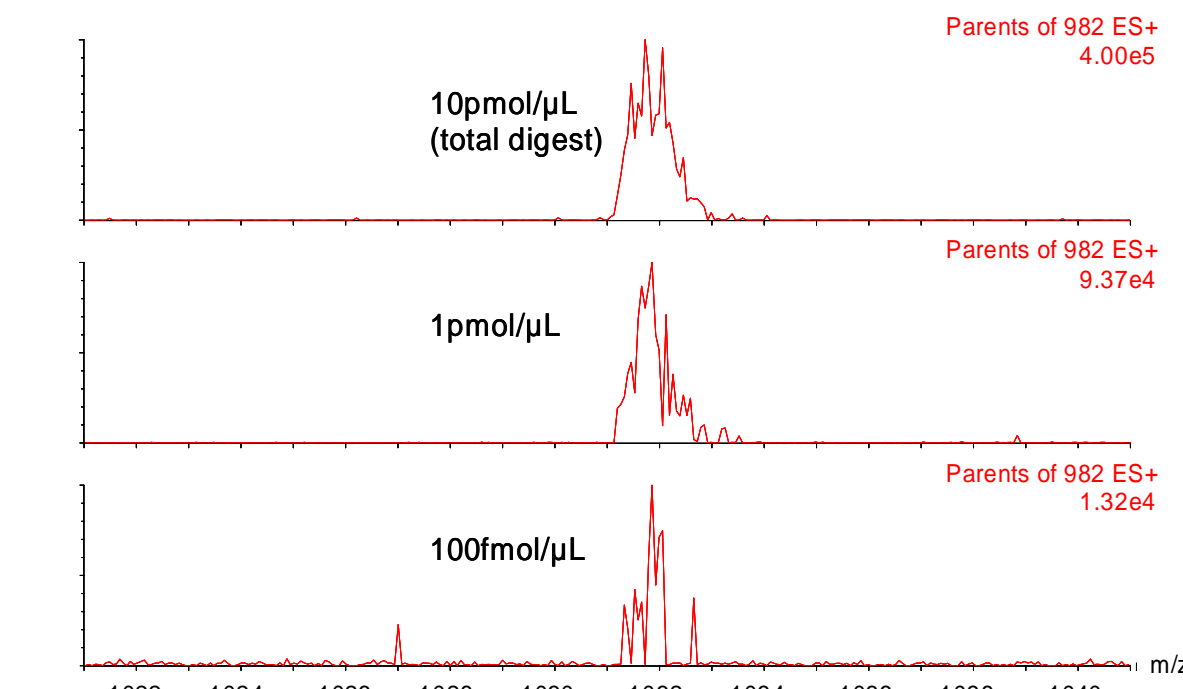


Figure 5 Conventional tandem quadrupole precursor ion scans for decreasing amounts of beta casein digest

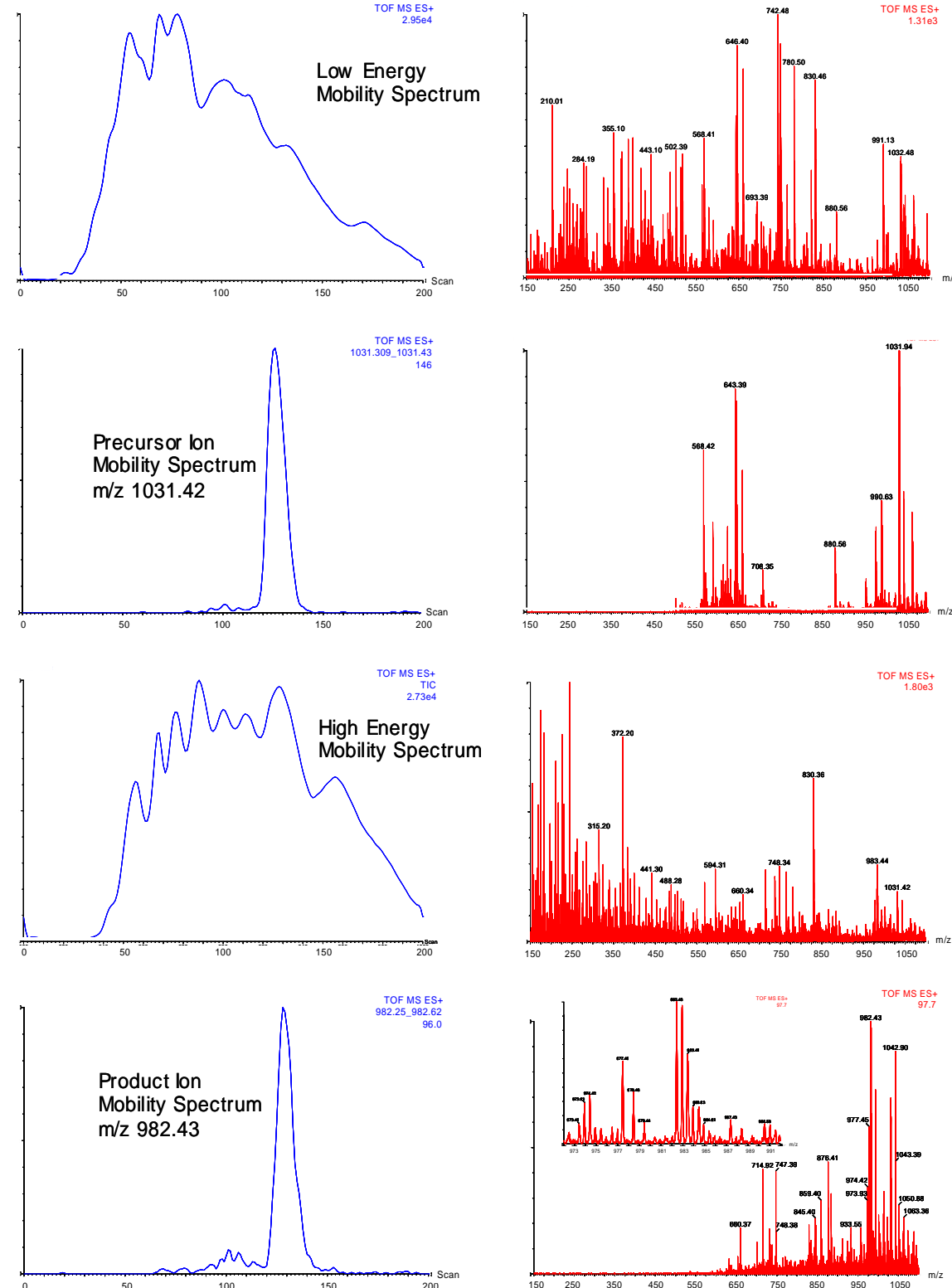


Figure 6 Ion mobility separations and their associated mass spectra obtained under low and high energy conditions for 100fmol/μL beta casein digest

CONCLUSIONS

- This study has demonstrated the feasibility of using a mobility-based separation of precursor ions to enhance duty cycle when compared with a scanning quadrupole.
- Further increases in performance have been demonstrated by using EDC acquisition on the oa-TOF analyser.
- This technique has been shown to enhance the sensitivity of precursor ion detection by at least a factor of 100 when compared with a scanning tandem quadrupole.

REFERENCES

- 1 'Applications of a Travelling Wave-Based Radio-Frequency-Only Stacked Ring Ion Guide', Giles K, Pringle SD, Worthington KR, Little D, Wildgoose JL and Bateman RH, *Rapid Commun. Mass Spectrom.*, **14** (2004) 2401